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(54) Title: NOVEL G PROTEIN-COUPLED RECEPTORS

(57) Abstract: The present invention provides a gene encoding a G protein-coupled receptor termed nGPCR-x; constructs and recombinant host cells incorporating the genes; the nGPCR-x polypeptides encoded by the gene; antibodies to the nGPCR-x polypeptides; and methods of making and using all of the foregoing.

NOVEL G PROTEIN-COUPLED RECEPTORS

CROSS-REFERENCE TO RELATED APPLICATIONS

The present application claims priority to Application Serial Numbers 60/184,305 filed February 23, 2000, 60/184,304 filed February 23, 2000, 60/184,303 filed February 23, 2000,
5 60/184,397 filed February 23, 2000, 60/184,247 filed February 23, 2000, 60/188,880 filed March 13, 2000, 60/217,369 filed July 11, 2000, 60/217,370 filed July 11, 2000, 60/219,492 filed July 20, 2000, 60/186,810 filed March 3, 2000, 60/188,064 filed March 9, 2000,
10 60/186,457 filed March 2, 2000, 60/213,861 filed June 23, 2000, 60/194,344 filed April 3, 2000, and 60/218,337 filed July 14, 2000, each of which is incorporated herein by reference in its entirety.

FIELD OF THE INVENTION

The present invention relates generally to the fields of genetics and cellular and molecular biology. More particularly, the invention relates to novel G protein coupled receptors, to polynucleotides that encode such novel receptors, to reagents such as antibodies, probes, 15 primers and kits comprising such antibodies, probes, primers related to the same, and to methods which use the novel G protein coupled receptors, polynucleotides or reagents.

BACKGROUND OF THE INVENTION

The G protein-coupled receptors (GPCRs) form a vast superfamily of cell surface receptors which are characterized by an amino-terminal extracellular domain, a carboxyl-terminal intracellular domain, and a serpentine structure that passes through the cell membrane seven times. Hence, such receptors are sometimes also referred to as seven transmembrane (7TM) receptors. These seven transmembrane domains define three extracellular loops and three intracellular loops, in addition to the amino- and carboxy-terminal domains. The extracellular portions of the receptor have a role in recognizing and binding one or more extracellular binding 25 partners (e.g., ligands), whereas the intracellular portions have a role in recognizing and communicating with downstream molecules in the signal transduction cascade.

The G protein-coupled receptors bind a variety of ligands including calcium ions, hormones, chemokines, neuropeptides, neurotransmitters, nucleotides, lipids, odorants, and even photons, and are important in the normal (and sometimes the aberrant) function of many cell types. (See generally Strosberg, *Eur. J. Biochem.* 196:1-10 (1991) and Bohm *et al.*, *Biochem J.* 322:1-18 (1997)). When a specific ligand binds to its corresponding receptor, the ligand typically stimulates the receptor to activate a specific heterotrimeric guanine-nucleotide-binding regulatory protein (G-protein) that is coupled to the intracellular portion of the receptor. The G protein in turn transmits a signal to an effector molecule within the cell, by either stimulating or

inhibiting the activity of that effector molecule. These effector molecules include adenylate cyclase, phospholipases and ion channels. Adenylate cyclase and phospholipases are enzymes that are involved in the production of the second messenger molecules cAMP, inositol triphosphate and diacyglycerol. It is through this sequence of events that an extracellular ligand 5 stimuli exerts intracellular changes through a G protein-coupled receptor. Each such receptor has its own characteristic primary structure, expression pattern, ligand-binding profile, and intracellular effector system.

Because of the vital role of G protein-coupled receptors in the communication between cells and their environment, such receptors are attractive targets for therapeutic intervention, for 10 example by activating or antagonizing such receptors. For receptors having a known ligand, the identification of agonists or antagonists may be sought specifically to enhance or inhibit the action of the ligand. Some G protein-coupled receptors have roles in disease pathogenesis (e.g., certain chemokine receptors that act as HIV co-receptors may have a role in AIDS pathogenesis), and are attractive targets for therapeutic intervention even in the absence of 15 knowledge of the natural ligand of the receptor. Other receptors are attractive targets for therapeutic intervention by virtue of their expression pattern in tissues or cell types that are themselves attractive targets for therapeutic intervention. Examples of this latter category of receptors include receptors expressed in immune cells, which can be targeted to either inhibit autoimmune responses or to enhance immune responses to fight pathogens or cancer; and 20 receptors expressed in the brain or other neural organs and tissues, which are likely targets in the treatment of mental disorder, depression, schizophrenia, bipolar disease, or other neurological disorders. This latter category of receptor is also useful as a marker for identifying and/or purifying (e.g., via fluorescence-activated cell sorting) cellular subtypes that express the 25 receptor. Unfortunately, only a limited number of G protein receptors from the central nervous system (CNS) are known. Thus, a need exists for G protein-coupled receptors that have been identified and show promise as targets for therapeutic intervention in a variety of animals, including humans.

SUMMARY OF THE INVENTION

The present invention relates to an isolated nucleic acid molecule that comprises a 30 nucleotide sequence that encodes a polypeptide comprising an amino acid sequence homologous to sequences selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, or a fragment thereof. In some embodiments, the nucleic acid molecule encodes at least a portion of SEQ ID NO:111 to SEQ ID NO:120. In some embodiments, the nucleic acid molecule comprises a sequence that encodes a polypeptide comprising a sequence selected from the group

consisting of SEQ ID NO:61 to SEQ ID NO:120, or a fragment thereof. In some embodiments, the nucleic acid molecule comprises a sequence homologous to a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, or a fragment thereof. In some embodiments, the nucleic acid molecule comprises a sequence selected from the group 5 consisting of SEQ ID NO:1 to SEQ ID NO:60, or fragments thereof.

According to some embodiments, the present invention provides vectors which comprise the nucleic acid molecule of the invention. In some embodiments, the vector is an expression vector.

According to some embodiments, the present invention provides host cells which 10 comprise the vectors of the invention. In some embodiments, the host cells comprise expression vectors.

The present invention provides an isolated nucleic acid molecule comprising a nucleotide sequence complementary to at least a portion of a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, said portion comprising at least 10 nucleotides.

15 The present invention provides a method of producing a polypeptide comprising a sequence selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, or a homolog or fragment thereof. The method comprising the steps of introducing a recombinant expression vector that includes a nucleotide sequence that encodes the polypeptide into a compatible host cell, growing the host cell under conditions for expression of the polypeptide 20 and recovering the polypeptide.

The present invention provides an isolated antibody which binds to an epitope on a polypeptide comprising a sequence selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, or a homolog or fragment thereof.

25 The present invention provides a method of inducing an immune response in a mammal against a polypeptide comprising a sequence selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, or a homolog or fragment thereof. The method comprises administering to a mammal an amount of the polypeptide sufficient to induce said immune response.

30 The present invention provides a method for identifying a compound which binds nGPCR-x. The method comprises the steps of contacting nGPCR-x with a compound and determining whether the compound binds nGPCR-x.

The present invention provides a method for identifying a compound which binds a nucleic acid molecule encoding nGPCR-x. The method comprises the steps of contacting said

nucleic acid molecule encoding nGPCR-x with a compound and determining whether said compound binds said nucleic acid molecule.

The present invention provides a method for identifying a compound which modulates the activity of nGPCR-x. The method comprises the steps of contacting nGPCR-x with a compound and determining whether nGPCR-x activity has been modulated.

The present invention provides a method of identifying an animal homolog of nGPCR-x. The method comprises the steps screening a nucleic acid database of the animal with a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, or a portion thereof and determining whether a portion of said library or database is homologous to said sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, or portion thereof.

The present invention provides a method of identifying an animal homolog of nGPCR-x. The methods comprises the steps screening a nucleic acid library of the animal with a nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, or a portion thereof and determining whether a portion of said library or database is homologous to said sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, or a portion thereof.

Another aspect of the present invention relates to methods of screening a human subject to diagnose a disorder affecting the brain or genetic predisposition therefor. The methods comprise the steps of assaying nucleic acid of a human subject to determine a presence or an absence of a mutation altering an amino acid sequence, expression, or biological activity of at least one nGPCR that is expressed in the brain. The nGPCR comprise an amino acid sequence selected from the group consisting of SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120, and allelic variants thereof. A diagnosis of the disorder or predisposition is made from the presence or absence of the mutation. The presence of a mutation altering the amino acid sequence, expression, or biological activity of the nGPCR in the nucleic acid correlates with an increased risk of developing the disorder.

The present invention further relates to methods of screening for a hereditary mental disorder genotype related to nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 in a human patient. The methods comprise the steps of providing a biological sample comprising nucleic acid from the patient, in which the nucleic acid includes sequences corresponding to alleles of nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70. The presence of one or more mutations in the nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 allele is detected indicative of a hereditary mental disorder genotype.

The present invention provides kits for screening a human subject to diagnose mental disorder or a genetic predisposition therefor. The kits include an oligonucleotide useful as a

probe for identifying polymorphisms in a human nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 gene. The oligonucleotide comprises 6-50 nucleotides in a sequence that is identical or complementary to a sequence of a wild type human nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 gene sequence or nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 coding sequence, except for one 5 sequence difference selected from the group consisting of a nucleotide addition, a nucleotide deletion, or nucleotide substitution. The kit also includes a media packaged with the oligonucleotide. The media contains information for identifying polymorphisms that correlate with mental disorder or a genetic predisposition therefor, the polymorphisms being identifiable using the oligonucleotide as a probe.

10 The present invention further relates to methods of identifying nGPCR allelic variants that correlates with mental disorders. The methods comprise the steps of providing biological samples that comprise nucleic acid from a human patient diagnosed with a mental disorder, or from the patient's genetic progenitors or progeny, and detecting in the nucleic acid the presence of one or more mutations in an nGPCR that is expressed in the brain. The nGPCR comprises an 15 amino acid sequence selected from the group consisting of SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120, and allelic variants thereof. The nucleic acid includes sequences corresponding to the gene or genes encoding nGPCR. The one or more mutations detected indicate an allelic variant that correlates with a mental disorder.

20 The present invention further relates to purified polynucleotides comprising nucleotide sequences encoding alleles of nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 from a human with mental disorder. The polynucleotide hybridizes to the complement of SEQ ID Numbers 1, 2, 8, 31, 34, 36, 37, 39, 40, and 51-60 under the following hybridization conditions: (a) hybridization for 16 hours at 42C in a hybridization solution comprising 50% formamide, 1% SDS, 1 M NaCl, 10% dextran sulfate and (b) washing 2 times for 30 minutes at 60C in a wash solution 25 comprising 0.1x SSC and 1% SDS. The polynucleotide that encodes nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 amino acid sequence of the human differs from SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120 by at least one residue.

30 The present invention also provides methods for identifying a modulator of biological activity of nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 comprising the steps of contacting a cell that expresses nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 in the presence and in the absence of a putative modulator compound and measuring nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 biological activity in the cell. The decreased or increased nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 biological activity in the presence versus absence of the putative modulator is indicative of a modulator of biological activity.

The present invention further provides methods to identify compounds useful for the treatment of mental disorders. The methods comprise the steps of contacting a composition comprising nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 with a compound suspected of binding nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70. The binding between nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 and the compound suspected of binding nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 is detected. Compounds identified as binding nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 are candidate compounds useful for the treatment of mental disorder. Compounds identified as binding nGPCR-42, 46, 48, 49, 51, 52, 61, 63, 70, or other nGPCRs can be further tested in other assays including, but not limited to, *in vivo* models, in order to confirm or quantitate their activity.

The present invention further provides methods for identifying a compound useful as a modulator of binding between nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 and a binding partner of nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70. The methods comprise the steps of contacting the binding partner and a composition comprising nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 in the presence and in the absence of a putative modulator compound and detecting binding between the binding partner and nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70. Decreased or increased binding between the binding partner and nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 in the presence of the putative modulator, as compared to binding in the absence of the putative modulator is indicative a modulator compound useful for the treatment of a related disease or disorder. Compounds identified as modulating binding between nGPCR-42, 46, 48, 49, 51, 52, 61, 63, 70, or other nGPCRs and an nGPCR-x binding partner can be further tested in other assays including, but not limited to, *in vivo* models, in order to confirm or quantitate their activity as modulators.

Another aspect of the present invention relates to methods of purifying a G protein from a sample containing a G protein. The methods comprise the steps of contacting the sample with an nGPCR for a time sufficient to allow the G protein to form a complex with the nGPCR, isolating the complex from remaining components of the sample, maintaining the complex under conditions which result in dissociation of the G protein from the nGPCR, and isolating said G protein from the nGPCR.

Another aspect of the present invention relates to methods of identifying a compound that binds to or modulates nGPCR-51. The methods comprise contacting a composition comprising nGPCR-51 and Peptide A with a test compound, or a plurality of test compounds, and determining whether the test compound or compounds compete with Peptide A for binding to nGPCR-51.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS**Definitions**

Various definitions are made throughout this document. Most words have the meaning that would be attributed to those words by one skilled in the art. Words specifically defined either below or elsewhere in this document have the meaning provided in the context of the present invention as a whole and as are typically understood by those skilled in the art.

“Synthesized” as used herein and understood in the art, refers to polynucleotides produced by purely chemical, as opposed to enzymatic, methods. “Wholly” synthesized DNA sequences are therefore produced entirely by chemical means, and “partially” synthesized DNAs embrace those wherein only portions of the resulting DNA were produced by chemical means.

By the term “region” is meant a physically contiguous portion of the primary structure of a biomolecule. In the case of proteins, a region is defined by a contiguous portion of the amino acid sequence of that protein.

The term “domain” is herein defined as referring to a structural part of a biomolecule that contributes to a known or suspected function of the biomolecule. Domains may be co-extensive with regions or portions thereof; domains may also incorporate a portion of a biomolecule that is distinct from a particular region, in addition to all or part of that region. Examples of GPCR protein domains include, but are not limited to, the extracellular (*i.e.*, N-terminal), transmembrane and cytoplasmic (*i.e.*, C-terminal) domains, which are co-extensive with like-named regions of GPCRs; each of the seven transmembrane segments of a GPCR; and each of the loop segments (both extracellular and intracellular loops) connecting adjacent transmembrane segments.

As used herein, the term “activity” refers to a variety of measurable indicia suggesting or revealing binding, either direct or indirect; affecting a response, *i.e.* having a measurable affect in response to some exposure or stimulus, including, for example, the affinity of a compound for directly binding a polypeptide or polynucleotide of the invention, or, for example, measurement of amounts of upstream or downstream proteins or other similar functions after some stimulus or event.

Unless indicated otherwise, as used herein, the abbreviation in lower case (gpcr) refers to a gene, cDNA, RNA or nucleic acid sequence, while the upper case version (GPCR) refers to a protein, polypeptide, peptide, oligopeptide, or amino acid sequence. The term “nGPCR-x” refers to any of the nGPCRs taught herein, while specific reference to a nGPCR (for example nGPCR-63) refers only to that specific nGPCR.

As used herein, the term "antibody" is meant to refer to complete, intact antibodies, and Fab, Fab', F(ab)2, and other fragments thereof. Complete, intact antibodies include monoclonal antibodies such as murine monoclonal antibodies, chimeric antibodies and humanized antibodies.

5 As used herein, the term "binding" means the physical or chemical interaction between two proteins or compounds or associated proteins or compounds or combinations thereof. Binding includes ionic, non-ionic, Hydrogen bonds, Van der Waals, hydrophobic interactions, etc. The physical interaction, the binding, can be either direct or indirect, indirect being through or due to the effects of another protein or compound. Direct binding refers to interactions that do 10 not take place through or due to the effect of another protein or compound but instead are without other substantial chemical intermediates. Binding may be detected in many different manners. As a non-limiting example, the physical binding interaction between a nGPCR-x of the invention and a compound can be detected using a labeled compound. Alternatively, functional evidence of binding can be detected using, for example, a cell transfected with and expressing a 15 nGPCR-x of the invention. Binding of the transfected cell to a ligand of the nGPCR that was transfected into the cell provides functional evidence of binding. Other methods of detecting binding are well-known to those of skill in the art.

20 As used herein, the term "compound" means any identifiable chemical or molecule, including, but not limited to, small molecule, peptide, protein, sugar, nucleotide, or nucleic acid, and such compound can be natural or synthetic.

As used herein, the term "complementary" refers to Watson-Crick basepairing between nucleotide units of a nucleic acid molecule.

25 As used herein, the term "contacting" means bringing together, either directly or indirectly, a compound into physical proximity to a polypeptide or polynucleotide of the invention. The polypeptide or polynucleotide can be in any number of buffers, salts, solutions etc. Contacting includes, for example, placing the compound into a beaker, microtiter plate, cell culture flask, or a microarray, such as a gene chip, or the like, which contains the nucleic acid molecule, or polypeptide encoding the nGPCR or fragment thereof.

30 As used herein, the phrase "homologous nucleotide sequence," or "homologous amino acid sequence," or variations thereof, refers to sequences characterized by a homology, at the nucleotide level or amino acid level, of at least the specified percentage. Homologous nucleotide sequences include those sequences coding for isoforms of proteins. Such isoforms can be expressed in different tissues of the same organism as a result of, for example, alternative splicing of RNA. Alternatively, isoforms can be encoded by different genes. Homologous

nucleotide sequences include nucleotide sequences encoding for a protein of a species other than humans, including, but not limited to, mammals. Homologous nucleotide sequences also include, but are not limited to, naturally occurring allelic variations and mutations of the nucleotide sequences set forth herein. A homologous nucleotide sequence does not, however, 5 include the nucleotide sequence encoding other known GPCRs. Homologous amino acid sequences include those amino acid sequences which contain conservative amino acid substitutions and which polypeptides have the same binding and/or activity. A homologous amino acid sequence does not, however, include the amino acid sequence encoding other known GPCRs. Percent homology can be determined by, for example, the Gap program (Wisconsin 10 Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University Research Park, Madison WI), using the default settings, which uses the algorithm of Smith and Waterman (*Adv. Appl. Math.*, 1981, 2, 482-489, which is incorporated herein by reference in its entirety).

As used herein, the term "isolated" nucleic acid molecule refers to a nucleic acid 15 molecule (DNA or RNA) that has been removed from its native environment. Examples of isolated nucleic acid molecules include, but are not limited to, recombinant DNA molecules contained in a vector, recombinant DNA molecules maintained in a heterologous host cell, partially or substantially purified nucleic acid molecules, and synthetic DNA or RNA molecules.

As used herein, the terms "modulates" or "modifies" means an increase or decrease in 20 the amount, quality, or effect of a particular activity or protein.

As used herein, the term "oligonucleotide" refers to a series of linked nucleotide residues which has a sufficient number of bases to be used in a polymerase chain reaction (PCR). This short sequence is based on (or designed from) a genomic or cDNA sequence and is used to amplify, confirm, or reveal the presence of an identical, similar or complementary DNA or RNA 25 in a particular cell or tissue. Oligonucleotides comprise portions of a DNA sequence having at least about 10 nucleotides and as many as about 50 nucleotides, preferably about 15 to 30 nucleotides. They are chemically synthesized and may be used as probes.

As used herein, the term "probe" refers to nucleic acid sequences of variable length, preferably between at least about 10 and as many as about 6,000 nucleotides, depending on use. 30 They are used in the detection of identical, similar, or complementary nucleic acid sequences. Longer length probes are usually obtained from a natural or recombinant source, are highly specific and much slower to hybridize than oligomers. They may be single- or double-stranded and carefully designed to have specificity in PCR, hybridization membrane-based, or ELISA-like technologies.

The term "preventing" refers to decreasing the probability that an organism contracts or develops an abnormal condition.

The term "treating" refers to having a therapeutic effect and at least partially alleviating or abrogating an abnormal condition in the organism.

5 The term "therapeutic effect" refers to the inhibition or activation factors causing or contributing to the abnormal condition. A therapeutic effect relieves to some extent one or more of the symptoms of the abnormal condition. In reference to the treatment of abnormal conditions, a therapeutic effect can refer to one or more of the following: (a) an increase in the proliferation, growth, and/or differentiation of cells; (b) inhibition (*i.e.*, slowing or stopping) of 10 cell death; (c) inhibition of degeneration; (d) relieving to some extent one or more of the symptoms associated with the abnormal condition; and (e) enhancing the function of the affected population of cells. Compounds demonstrating efficacy against abnormal conditions can be identified as described herein.

15 The term "abnormal condition" refers to a function in the cells or tissues of an organism that deviates from their normal functions in that organism. An abnormal condition can relate to cell proliferation, cell differentiation, cell signaling, or cell survival. An abnormal condition may also include obesity, diabetic complications such as retinal degeneration, and irregularities in glucose uptake and metabolism, and fatty acid uptake and metabolism.

20 Abnormal cell proliferative conditions include cancers such as fibrotic and mesangial disorders, abnormal angiogenesis and vasculogenesis, wound healing, psoriasis, diabetes mellitus, and inflammation.

25 Abnormal differentiation conditions include, but are not limited to, neurodegenerative disorders, slow wound healing rates, and slow tissue grafting healing rates. Abnormal cell signaling conditions include, but are not limited to, psychiatric disorders involving excess neurotransmitter activity.

Abnormal cell survival conditions may also relate to conditions in which programmed cell death (apoptosis) pathways are activated or abrogated. A number of protein kinases are associated with the apoptosis pathways. Aberrations in the function of any one of the protein kinases could lead to cell immortality or premature cell death.

30 The term "administering" relates to a method of incorporating a compound into cells or tissues of an organism. The abnormal condition can be prevented or treated when the cells or tissues of the organism exist within the organism or outside of the organism. Cells existing outside the organism can be maintained or grown in cell culture dishes. For cells harbored within the organism, many techniques exist in the art to administer compounds, including (but

not limited to) oral, parenteral, dermal, injection, and aerosol applications. For cells outside of the organism, multiple techniques exist in the art to administer the compounds, including (but not limited to) cell microinjection techniques, transformation techniques and carrier techniques.

The abnormal condition can also be prevented or treated by administering a compound to 5 a group of cells having an aberration in a signal transduction pathway to an organism. The effect of administering a compound on organism function can then be monitored. The organism is preferably a mouse, rat, rabbit, guinea pig or goat, more preferably a monkey or ape, and most preferably a human.

By "amplification" it is meant increased numbers of DNA or RNA in a cell compared 10 with normal cells. "Amplification" as it refers to RNA can be the detectable presence of RNA in cells, since in some normal cells there is no basal expression of RNA. In other normal cells, a basal level of expression exists, therefore in these cases amplification is the detection of at least 1 to 2-fold, and preferably more, compared to the basal level.

As used herein, the phrase "stringent hybridization conditions" or "stringent conditions" 15 refers to conditions under which a probe, primer, or oligonucleotide will hybridize to its target sequence, but to no other sequences. Stringent conditions are sequence-dependent and will be different in different circumstances. Longer sequences hybridize specifically at higher temperatures. Generally, stringent conditions are selected to be about 5C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the 20 temperature (under defined ionic strength, pH and nucleic acid concentration) at which 50% of the probes complementary to the target sequence hybridize to the target sequence at equilibrium. Since the target sequences are generally present in excess, at T_m , 50% of the probes are occupied 25 at equilibrium. Typically, stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, typically about 0.01 to 1.0 M sodium ion (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30C for short probes, primers or oligonucleotides (e.g. 10 to 50 nucleotides) and at least about 60C for longer probes, primers or oligonucleotides. Stringent conditions may also be achieved with the addition of destabilizing agents, such as formamide.

The amino acid sequences are presented in the amino to carboxy direction, from left to 30 right. The amino and carboxy groups are not presented in the sequence. The nucleotide sequences are presented by single strand only, in the 5' to 3' direction, from left to right. Nucleotides and amino acids are represented in the manner recommended by the IUPAC-IUB Biochemical Nomenclature Commission or (for amino acids) by three letters code.

Polynucleotides

The present invention provides purified and isolated polynucleotides (e.g., DNA sequences and RNA transcripts, both sense and complementary antisense strands, both single- and double-stranded, including splice variants thereof) that encode unknown G protein-coupled receptors heretofore termed novel GPCRs, or nGPCRs. These genes are described herein and designated herein collectively as nGPCR-x (where x is 42, 44, 45, 46, 47, 48, 49, 50, 51, 52, 61, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 2001, 2002, 2003, 2004, 2005, 2006, 2007, 2008, 2009, 2010, 2011, 2012, 2013, 2014, 2015, 2016, 2017, 2018, 2019, 2020, 2021, 2022, 2024, 2025, 2026, 2027, 2028, 2029, and 2030). That is, these genes are described herein and designated herein as nGPCR-42, nGPCR-44, etc. Table 1 below identifies the novel gene sequence 5 nGPCR-x designation, the SEQ ID NO: of the gene sequence, the SEQ ID NO: of the polypeptide encoded thereby, and the U.S. Provisional Application in which the gene sequence 10 has been disclosed.

Table 1

nGPCR	Nucleotide Sequence (SEQ ID NO:)	Amino acid Sequence (SEQ ID NO:)	Originally filed in:
0042	31	91	D
0042	54	114	I
0044	32	92	D
0045	33	93	D
0046	34	94	D
0046	55	115	J
0047	35	95	D
0048	36	96	D
0048	56	116	K
0049	37	97	D
0049	59	119	N
0050	38	98	D
0051	39	99	D
0051	57	117	L
0052	40	100	D
0052	58	118	M
0061	1	61	A
0061	60	120	O
0063	2	62	A
0063	53	113	H
0063	51	111	F
0064	3	63	A
0065	4	64	A
0066	5	65	A
0067	6	66	A
0068	7	67	A
0069	10	70	A
0070	8	68	A
0070	52	112	G
0071	9	69	A
0072	43	103	E
2001	21	81	C
2002	22	82	C
2003	23	83	C
2004	24	84	C
2005	25	85	C
2006	26	86	C
2007	27	87	C
2008	28	88	C

2009	29	89	C
2010	30	90	C
2011	11	71	B
2012	12	72	B
2013	13	73	B
2014	14	74	B
2015	15	75	B
2016	16	76	B
2017	17	77	B
2018	18	78	B
2019	19	79	B
2020	20	80	B
2021	41	101	E
2022	42	102	E
2024	44	104	E
2025	45	105	E
2026	46	106	E
2027	47	107	E
2028	48	108	E
2029	49	109	E
2030	50	110	E

Legend

A= Ser. No. 60/184,305

B= Ser. No. 60/184,304

C= Ser. No. 60/184,303

D= Ser. No. 60/184,397

E= Ser. No. 60/184,247

F= Ser. No. 60/188,880

5

G= Ser. No. 60/217,369

H= Ser. No. 60/219,492

I= Ser. No. 60/186,810

J= Ser. No. 60/188,064

K= Ser. No. 60/186,457

L= Ser. No. 60/213,861

M= Ser. No. 60/194,344

N= Ser. No. 60/218,337

O= Ser. No. 60/217,370

10

When a specific nGPCR is identified (for example nGPCR-63), it is understood that only that specific nGPCR is being referred to.

As described in Example 5 below, the genes encoding nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 have been detected in brain tissue indicating that these nGPCR proteins are 15 neuroreceptors.

20

The invention provides purified and isolated polynucleotides (e.g., cDNA, genomic DNA, synthetic DNA, RNA, or combinations thereof, whether single- or double-stranded) that comprise a nucleotide sequence encoding the amino acid sequence of the polypeptides of the invention. Such polynucleotides are useful for recombinantly expressing the receptor and also for detecting expression of the receptor in cells (e.g., using Northern hybridization and *in situ* hybridization assays). Such polynucleotides also are useful in the design of antisense and other molecules for the suppression of the expression of nGPCR-x in a cultured cell, a tissue, or an animal; for therapeutic purposes; or to provide a model for diseases or conditions characterized by aberrant nGPCR-x expression. Specifically excluded from the definition of polynucleotides 25 of the invention are entire isolated, non-recombinant native chromosomes of host cells. A preferred polynucleotide has a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, which correspond to naturally occurring nGPCR-x sequences. It will be appreciated that numerous other polynucleotide sequences exist that also encode nGPCR-x

having the sequence selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, due to the well-known degeneracy of the universal genetic code.

The invention also provides a purified and isolated polynucleotide comprising a nucleotide sequence that encodes a mammalian polypeptide, wherein the polynucleotide 5 hybridizes to a polynucleotide having the sequence set forth in sequences selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, or the non-coding strand complementary thereto, under the following hybridization conditions: (a) hybridization for 16 hours at 42C in a hybridization solution comprising 50% formamide, 1% SDS, 1 M NaCl, 10% dextran sulfate; and (b) washing 2 times for 30 minutes each at 60C in a wash solution comprising 0.1% SSC, 10 1% SDS. Polynucleotides that encode a human allelic variant are highly preferred.

The present invention relates to molecules which comprise the gene sequences that encode the nGPCRs; constructs and recombinant host cells incorporating the gene sequences; the novel GPCR polypeptides encoded by the gene sequences; antibodies to the polypeptides and homologs; kits employing the polynucleotides and polypeptides, and methods of making 15 and using all of the foregoing. In addition, the present invention relates to homologs of the gene sequences and of the polypeptides and methods of making and using the same.

Genomic DNA of the invention comprises the protein-coding region for a polypeptide of the invention and is also intended to include allelic variants thereof. It is widely understood that, for many genes, genomic DNA is transcribed into RNA transcripts that undergo one or more 20 splicing events wherein intron (*i.e.*, non-coding regions) of the transcripts are removed, or “spliced out.” RNA transcripts that can be spliced by alternative mechanisms, and therefore be subject to removal of different RNA sequences but still encode a nGPCR-x polypeptide, are referred to in the art as splice variants which are embraced by the invention. Splice variants comprehended by the invention therefore are encoded by the same original genomic DNA sequences but arise from distinct mRNA transcripts. Allelic variants are modified forms of a wild-type gene sequence, the modification resulting from recombination during chromosomal 25 segregation or exposure to conditions which give rise to genetic mutation. Allelic variants, like wild type genes, are naturally occurring sequences (as opposed to non-naturally occurring variants that arise from *in vitro* manipulation).

30 The invention also comprehends cDNA that is obtained through reverse transcription of an RNA polynucleotide encoding nGPCR-x (conventionally followed by second strand synthesis of a complementary strand to provide a double-stranded DNA).

Preferred DNA sequences encoding human nGPCR-x polypeptides are selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60. A preferred DNA of the invention

comprises a double stranded molecule along with the complementary molecule (the “non-coding strand” or “complement”) having a sequence unambiguously deducible from the coding strand according to Watson-Crick base-pairing rules for DNA. Also preferred are other polynucleotides encoding the nGPCR-x polypeptide selected from the group consisting of SEQ ID NO:61 to 5 SEQ ID NO:120, which differ in sequence from the polynucleotides selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, by virtue of the well-known degeneracy of the universal nuclear genetic code.

The invention further embraces other species, preferably mammalian, homologs of the human nGPCR-x DNA. Species homologs, sometimes referred to as “orthologs,” in general, 10 share at least 35%, at least 40%, at least 45%, at least 50%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98%, or at least 99% homology with human DNA of the invention. Generally, percent sequence “homology” with respect to polynucleotides of the invention may be calculated as the percentage of nucleotide bases in the candidate sequence that are identical to nucleotides in the nGPCR-x 15 sequence set forth in sequences selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity.

Polynucleotides of the invention permit identification and isolation of polynucleotides encoding related nGPCR-x polypeptides, such as human allelic variants and species homologs, 20 by well-known techniques including Southern and/or Northern hybridization, and polymerase chain reaction (PCR). Examples of related polynucleotides include human and non-human genomic sequences, including allelic variants, as well as polynucleotides encoding polypeptides homologous to nGPCR-x and structurally related polypeptides sharing one or more biological, immunological, and/or physical properties of nGPCR-x. Non-human species genes encoding 25 proteins homologous to nGPCR-x can also be identified by Southern and/or PCR analysis and are useful in animal models for nGPCR-x disorders. Knowledge of the sequence of a human nGPCR-x DNA also makes possible through use of Southern hybridization or polymerase chain reaction (PCR) the identification of genomic DNA sequences encoding nGPCR-x expression control regulatory sequences such as promoters, operators, enhancers, repressors, and the like. 30 Polynucleotides of the invention are also useful in hybridization assays to detect the capacity of cells to express nGPCR-x. Polynucleotides of the invention may also provide a basis for diagnostic methods useful for identifying a genetic alteration(s) in a nGPCR-x locus that underlies a disease state or states, which information is useful both for diagnosis and for selection of therapeutic strategies.

According to the present invention, the nGPCR-x nucleotide sequences disclosed herein may be used to identify homologs of the nGPCR-x, in other animals, including but not limited to humans and other mammals, and invertebrates. Any of the nucleotide sequences disclosed herein, or any portion thereof, can be used, for example, as probes to screen databases or nucleic acid libraries, such as, for example, genomic or cDNA libraries, to identify homologs, using screening procedures well known to those skilled in the art. Accordingly, homologs having at least 50%, more preferably at least 60%, more preferably at least 70%, more preferably at least 80%, more preferably at least 90%, more preferably at least 95%, and most preferably at least 100% homology with nGPCR-x sequences can be identified.

The disclosure herein of full-length polynucleotides encoding nGPCR-x polypeptides makes readily available to the worker of ordinary skill in the art every possible fragment of the full-length polynucleotide.

One preferred embodiment of the present invention provides an isolated nucleic acid molecule comprising a sequence homologous sequences selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, and fragments thereof. Another preferred embodiment provides an isolated nucleic acid molecule comprising a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, and fragments thereof.

As used in the present invention, fragments of nGPCR-x-encoding polynucleotides comprise at least 10, and preferably at least 12, 14, 16, 18, 20, 25, 50, or 75 consecutive nucleotides of a polynucleotide encoding nGPCR-x. Preferably, fragment polynucleotides of the invention comprise sequences unique to the nGPCR-x-encoding polynucleotide sequence, and therefore hybridize under highly stringent or moderately stringent conditions only (*i.e.*, “specifically”) to polynucleotides encoding nGPCR-x (or fragments thereof). Polynucleotide fragments of genomic sequences of the invention comprise not only sequences unique to the coding region, but also include fragments of the full-length sequence derived from introns, regulatory regions, and/or other non-translated sequences. Sequences unique to polynucleotides of the invention are recognizable through sequence comparison to other known polynucleotides, and can be identified through use of alignment programs routinely utilized in the art, *e.g.*, those made available in public sequence databases. Such sequences also are recognizable from Southern hybridization analyses to determine the number of fragments of genomic DNA to which a polynucleotide will hybridize. Polynucleotides of the invention can be labeled in a manner that permits their detection, including radioactive, fluorescent, and enzymatic labeling.

Fragment polynucleotides are particularly useful as probes for detection of full-length or fragments of nGPCR-x polynucleotides. One or more polynucleotides can be included in kits

that are used to detect the presence of a polynucleotide encoding nGPCR-x, or used to detect variations in a polynucleotide sequence encoding nGPCR-x.

The invention also embraces DNAs encoding nGPCR-x polypeptides that hybridize under moderately stringent or high stringency conditions to the non-coding strand, or 5 complement, of the polynucleotides set forth in sequences selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60.

Exemplary highly stringent hybridization conditions are as follows: hybridization at 42C in a hybridization solution comprising 50% formamide, 1% SDS, 1 M NaCl, 10% Dextran sulfate, and washing twice for 30 minutes at 60C in a wash solution comprising 0.1 X SSC and 10 1% SDS. It is understood in the art that conditions of equivalent stringency can be achieved through variation of temperature and buffer, or salt concentration as described Ausubel *et al.*

10 (Eds.), *Protocols in Molecular Biology*, John Wiley & Sons (1994), pp. 6.0.3 to 6.4.10. Modifications in hybridization conditions can be empirically determined or precisely calculated based on the length and the percentage of guanosine/cytosine (GC) base pairing of the probe. 15 The hybridization conditions can be calculated as described in Sambrook, *et al.*, (Eds.), *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press: Cold Spring Harbor, New York (1989), pp. 9.47 to 9.51.

With the knowledge of the nucleotide sequence information disclosed in the present invention, one skilled in the art can identify and obtain nucleotide sequences which encode 20 nGPCR-x from different sources (*i.e.*, different tissues or different organisms) through a variety of means well known to the skilled artisan and as disclosed by, for example, Sambrook *et al.*, "Molecular cloning: a laboratory manual", Second Edition, Cold Spring Harbor Press, Cold Spring Harbor, NY (1989), which is incorporated herein by reference in its entirety.

For example, DNA that encodes nGPCR-x may be obtained by screening of mRNA, 25 cDNA, or genomic DNA with oligonucleotide probes generated from the nGPCR-x gene sequence information provided herein. Probes may be labeled with a detectable group, such as a fluorescent group, a radioactive atom or a chemiluminescent group in accordance with procedures known to the skilled artisan and used in conventional hybridization assays, as described by, for example, Sambrook *et al.*

30 A nucleic acid molecule comprising any of the nGPCR-x nucleotide sequences described above can alternatively be synthesized by use of the polymerase chain reaction (PCR) procedure, with the PCR oligonucleotide primers produced from the nucleotide sequences provided herein. See U.S. Patent Numbers 4,683,195 to Mullis *et al.* and 4,683,202 to Mullis. The PCR reaction provides a method for selectively increasing the concentration of a particular

nucleic acid sequence even when that sequence has not been previously purified and is present only in a single copy in a particular sample. The method can be used to amplify either single- or double-stranded DNA. The essence of the method involves the use of two oligonucleotide probes to serve as primers for the template-dependent, polymerase mediated replication of a 5 desired nucleic acid molecule.

A wide variety of alternative cloning and *in vitro* amplification methodologies are well known to those skilled in the art. Examples of these techniques are found in, for example, Berger *et al.*, *Guide to Molecular Cloning Techniques*, Methods in Enzymology 152, Academic Press, Inc., San Diego, CA (Berger), which is incorporated herein by reference in its entirety.

10 Automated sequencing methods can be used to obtain or verify the nucleotide sequence of nGPCR-x. The nGPCR-x nucleotide sequences of the present invention are believed to be 100% accurate. However, as is known in the art, nucleotide sequence obtained by automated methods may contain some errors. Nucleotide sequences determined by automation are typically at least about 90%, more typically at least about 95% to at least about 99.9% identical to the 15 actual nucleotide sequence of a given nucleic acid molecule. The actual sequence may be more precisely determined using manual sequencing methods, which are well known in the art. An error in a sequence which results in an insertion or deletion of one or more nucleotides may result in a frame shift in translation such that the predicted amino acid sequence will differ from that which would be predicted from the actual nucleotide sequence of the nucleic acid molecule, 20 starting at the point of the mutation.

The nucleic acid molecules of the present invention, and fragments derived therefrom, are useful for screening for restriction fragment length polymorphism (RFLP) associated with certain disorders, as well as for genetic mapping.

25 The polynucleotide sequence information provided by the invention makes possible large-scale expression of the encoded polypeptide by techniques well known and routinely practiced in the art.

Vectors

Another aspect of the present invention is directed to vectors, or recombinant expression 30 vectors, comprising any of the nucleic acid molecules described above. Vectors are used herein either to amplify DNA or RNA encoding nGPCR-x and/or to express DNA which encodes nGPCR-x. Preferred vectors include, but are not limited to, plasmids, phages, cosmids, episomes, viral particles or viruses, and integratable DNA fragments (*i.e.*, fragments integratable into the host genome by homologous recombination). Preferred viral particles include, but are not limited to, adenoviruses, baculoviruses, parvoviruses, herpesviruses, poxviruses, adeno-

associated viruses, Semliki Forest viruses, vaccinia viruses, and retroviruses. Preferred expression vectors include, but are not limited to, pcDNA3 (Invitrogen) and pSVL (Pharmacia Biotech). Other expression vectors include, but are not limited to, pSPORT™ vectors, pGEM™ vectors (Promega), pPROEXvectors™ (LTI, Bethesda, MD), Bluescript™ vectors (Stratagene), 5 pQE™ vectors (Qiagen), pSE420™ (Invitrogen), and pYES2™(Invitrogen).

Expression constructs preferably comprise GPCR-x-encoding polynucleotides operatively linked to an endogenous or exogenous expression control DNA sequence and a transcription terminator. Expression control DNA sequences include promoters, enhancers, operators, and regulatory element binding sites generally, and are typically selected based on the 10 expression systems in which the expression construct is to be utilized. Preferred promoter and enhancer sequences are generally selected for the ability to increase gene expression, while operator sequences are generally selected for the ability to regulate gene expression. Expression constructs of the invention may also include sequences encoding one or more selectable markers that permit identification of host cells bearing the construct. Expression constructs may also 15 include sequences that facilitate, and preferably promote, homologous recombination in a host cell. Preferred constructs of the invention also include sequences necessary for replication in a host cell.

Expression constructs are preferably utilized for production of an encoded protein, but may also be utilized simply to amplify a nGPCR-x-encoding polynucleotide sequence. In 20 preferred embodiments, the vector is an expression vector wherein the polynucleotide of the invention is operatively linked to a polynucleotide comprising an expression control sequence. Autonomously replicating recombinant expression constructs such as plasmid and viral DNA vectors incorporating polynucleotides of the invention are also provided. Preferred expression vectors are replicable DNA constructs in which a DNA sequence encoding nGPCR-x is 25 operably linked or connected to suitable control sequences capable of effecting the expression of the nGPCR-x in a suitable host. DNA regions are operably linked or connected when they are functionally related to each other. For example, a promoter is operably linked or connected to a coding sequence if it controls the transcription of the sequence. Amplification vectors do not require expression control domains, but rather need only the ability to replicate in a host, usually 30 conferred by an origin of replication, and a selection gene to facilitate recognition of transformants. The need for control sequences in the expression vector will vary depending upon the host selected and the transformation method chosen. Generally, control sequences include a transcriptional promoter, an optional operator sequence to control transcription, a

sequence encoding suitable mRNA ribosomal binding and sequences which control the termination of transcription and translation.

Preferred vectors preferably contain a promoter that is recognized by the host organism. The promoter sequences of the present invention may be prokaryotic, eukaryotic or viral.

5 Examples of suitable prokaryotic sequences include the P_R and P_L promoters of bacteriophage lambda (The bacteriophage Lambda, Hershey, A. D., Ed., Cold Spring Harbor Press, Cold Spring Harbor, NY (1973), which is incorporated herein by reference in its entirety; Lambda II, Hendrix, R. W., Ed., Cold Spring Harbor Press, Cold Spring Harbor, NY (1980), which is incorporated herein by reference in its entirety); the *trp*, *recA*, heat shock, and *lacZ* promoters of 10 *E. coli* and the SV40 early promoter (Benoist *et al.* *Nature*, 1981, 290, 304-310, which is incorporated herein by reference in its entirety). Additional promoters include, but are not limited to, mouse mammary tumor virus, long terminal repeat of human immunodeficiency virus, maloney virus, cytomegalovirus immediate early promoter, Epstein Barr virus, Rous sarcoma virus, human actin, human myosin, human hemoglobin, human muscle creatine, and 15 human metallothionein.

Additional regulatory sequences can also be included in preferred vectors. Preferred examples of suitable regulatory sequences are represented by the Shine-Dalgarno of the replicase gene of the phage MS-2 and of the gene cII of bacteriophage lambda. The Shine-Dalgarno sequence may be directly followed by DNA encoding nGPCR-x and result in the 20 expression of the mature nGPCR-x protein.

Moreover, suitable expression vectors can include an appropriate marker that allows the screening of the transformed host cells. The transformation of the selected host is carried out using any one of the various techniques well known to the expert in the art and described in Sambrook *et al.*, *supra*.

25 An origin of replication can also be provided either by construction of the vector to include an exogenous origin or may be provided by the host cell chromosomal replication mechanism. If the vector is integrated into the host cell chromosome, the latter may be sufficient. Alternatively, rather than using vectors which contain viral origins of replication, one skilled in the art can transform mammalian cells by the method of co-transformation with a 30 selectable marker and nGPCR-x DNA. An example of a suitable marker is dihydrofolate reductase (DHFR) or thymidine kinase (see, U.S. Patent No. 4,399,216).

Nucleotide sequences encoding GPCR-x may be recombined with vector DNA in accordance with conventional techniques, including blunt-ended or staggered-ended termini for ligation, restriction enzyme digestion to provide appropriate termini, filling in of cohesive ends

as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and ligation with appropriate ligases. Techniques for such manipulation are disclosed by Sambrook et al., *supra* and are well known in the art. Methods for construction of mammalian expression vectors are disclosed in, for example, Okayama et al., *Mol. Cell. Biol.*, 1983, 3, 280, Cosman et al., *Mol. Immunol.*, 1986, 23, 935, Cosman et al., *Nature*, 1984, 312, 768, EP-A-0367566, and WO 91/18982, each of which is incorporated herein by reference in its entirety.

5 Host cells

According to another aspect of the invention, host cells are provided, including prokaryotic and eukaryotic cells, comprising a polynucleotide of the invention (or vector of the invention) in a manner that permits expression of the encoded nGPCR-x polypeptide. Polynucleotides of the invention may be introduced into the host cell as part of a circular plasmid, or as linear DNA comprising an isolated protein coding region or a viral vector. Methods for introducing DNA into the host cell that are well known and routinely practiced in the art include transformation, transfection, electroporation, nuclear injection, or fusion with carriers such as liposomes, micelles, ghost cells, and protoplasts. Expression systems of the invention include bacterial, yeast, fungal, plant, insect, invertebrate, vertebrate, and mammalian cells systems.

10 The invention provides host cells that are transformed or transfected (stably or transiently) with polynucleotides of the invention or vectors of the invention. As stated above, such host cells are useful for amplifying the polynucleotides and also for expressing the nGPCR-x polypeptide or fragment thereof encoded by the polynucleotide.

In still another related embodiment, the invention provides a method for producing a nGPCR-x polypeptide (or fragment thereof) comprising the steps of growing a host cell of the invention in a nutrient medium and isolating the polypeptide or variant thereof from the cell or the medium. Because nGPCR-x is a seven transmembrane receptor, it will be appreciated that, for some applications, such as certain activity assays, the preferable isolation may involve isolation of cell membranes containing the polypeptide embedded therein, whereas for other applications a more complete isolation may be preferable.

According to some aspects of the present invention, transformed host cells having an expression vector comprising any of the nucleic acid molecules described above are provided. Expression of the nucleotide sequence occurs when the expression vector is introduced into an appropriate host cell. Suitable host cells for expression of the polypeptides of the invention include, but are not limited to, prokaryotes, yeast, and eukaryotes. If a prokaryotic expression vector is employed, then the appropriate host cell would be any prokaryotic cell capable of

expressing the cloned sequences. Suitable prokaryotic cells include, but are not limited to, bacteria of the genera *Escherichia*, *Bacillus*, *Salmonella*, *Pseudomonas*, *Streptomyces*, and *Staphylococcus*.

If an eukaryotic expression vector is employed, then the appropriate host cell would be
5 any eukaryotic cell capable of expressing the cloned sequence. Preferably, eukaryotic cells are cells of higher eukaryotes. Suitable eukaryotic cells include, but are not limited to, non-human mammalian tissue culture cells and human tissue culture cells. Preferred host cells include, but are not limited to, insect cells, HeLa cells, Chinese hamster ovary cells (CHO cells), African green monkey kidney cells (COS cells), human 293 cells, and murine 3T3 fibroblasts.
10 Propagation of such cells in cell culture has become a routine procedure (see, *Tissue Culture*, Academic Press, Kruse and Patterson, eds. (1973), which is incorporated herein by reference in its entirety).

In addition, a yeast host may be employed as a host cell. Preferred yeast cells include, but are not limited to, the genera *Saccharomyces*, *Pichia*, and *Kluveromyces*. Preferred yeast hosts are *S. cerevisiae* and *P. pastoris*. Preferred yeast vectors can contain an origin of replication sequence from a 2T yeast plasmid, an autonomously replication sequence (ARS), a promoter region, sequences for polyadenylation, sequences for transcription termination, and a selectable marker gene. Shuttle vectors for replication in both yeast and *E. coli* are also included herein.
15

20 Alternatively, insect cells may be used as host cells. In a preferred embodiment, the polypeptides of the invention are expressed using a baculovirus expression system (see, Luckow *et al.*, *Bio/Technology*, 1988, 6, 47, *Baculovirus Expression Vectors: A Laboratory Manual*, O’Rielly *et al.* (Eds.), W.H. Freeman and Company, New York, 1992, and U.S. Patent No. 4,879,236, each of which is incorporated herein by reference in its entirety). In addition, the 25 MAXBAC™ complete baculovirus expression system (Invitrogen) can, for example, be used for production in insect cells.

Host cells of the invention are a valuable source of immunogen for development of antibodies specifically immunoreactive with nGPCR-x. Host cells of the invention are also useful in methods for the large-scale production of nGPCR-x polypeptides wherein the cells are 30 grown in a suitable culture medium and the desired polypeptide products are isolated from the cells, or from the medium in which the cells are grown, by purification methods known in the art, *e.g.*, conventional chromatographic methods including immunoaffinity chromatography, receptor affinity chromatography, hydrophobic interaction chromatography, lectin affinity chromatography, size exclusion filtration, cation or anion exchange chromatography, high

pressure liquid chromatography (HPLC), reverse phase HPLC, and the like. Still other methods of purification include those methods wherein the desired protein is expressed and purified as a fusion protein having a specific tag, label, or chelating moiety that is recognized by a specific binding partner or agent. The purified protein can be cleaved to yield the desired protein, or can be left as an intact fusion protein. Cleavage of the fusion component may produce a form of the desired protein having additional amino acid residues as a result of the cleavage process.

Knowledge of nGPCR-x DNA sequences allows for modification of cells to permit, or increase, expression of endogenous nGPCR-x. Cells can be modified (e.g., by homologous recombination) to provide increased expression by replacing, in whole or in part, the naturally occurring nGPCR-x promoter with all or part of a heterologous promoter so that the cells express nGPCR-x at higher levels. The heterologous promoter is inserted in such a manner that it is operatively linked to endogenous nGPCR-x encoding sequences. (See, for example, PCT International Publication No. WO 94/12650, PCT International Publication No. WO 92/20808, and PCT International Publication No. WO 91/09955.) It is also contemplated that, in addition to heterologous promoter DNA, amplifiable marker DNA (e.g., ada, dhfr, and the multifunctional CAD gene which encodes carbamoyl phosphate synthase, aspartate transcarbamylase, and dihydroorotase) and/or intron DNA may be inserted along with the heterologous promoter DNA. If linked to the nGPCR-x coding sequence, amplification of the marker DNA by standard selection methods results in co-amplification of the nGPCR-x coding sequences in the cells.

Knock-outs

The DNA sequence information provided by the present invention also makes possible the development (e.g., by homologous recombination or "knock-out" strategies; see Capecchi, *Science* 244:1288-1292 (1989), which is incorporated herein by reference in its entirety) of animals that fail to express functional nGPCR-x or that express a variant of nGPCR-x. Such animals (especially small laboratory animals such as rats, rabbits, and mice) are useful as models for studying the *in vivo* activities of nGPCR-x and modulators of nGPCR-x.

Antisense

Also made available by the invention are anti-sense polynucleotides that recognize and hybridize to polynucleotides encoding nGPCR-x. Full-length and fragment anti-sense polynucleotides are provided. Fragment antisense molecules of the invention include (i) those that specifically recognize and hybridize to nGPCR-x RNA (as determined by sequence comparison of DNA encoding nGPCR-x to DNA encoding other known molecules). Identification of sequences unique to nGPCR-x encoding polynucleotides can be deduced

through use of any publicly available sequence database, and/or through use of commercially available sequence comparison programs. After identification of the desired sequences, isolation through restriction digestion or amplification using any of the various polymerase chain reaction techniques well known in the art can be performed. Anti-sense polynucleotides are particularly relevant to regulating expression of nGPCR-x by those cells expressing nGPCR-x mRNA.

5 Antisense nucleic acids (preferably 10 to 30 base-pair oligonucleotides) capable of specifically binding to nGPCR-x expression control sequences or nGPCR-x RNA are introduced into cells (e.g., by a viral vector or colloidal dispersion system such as a liposome). The antisense nucleic acid binds to the nGPCR-x target nucleotide sequence in the cell and prevents 10 transcription and/or translation of the target sequence. Phosphorothioate and methylphosphonate antisense oligonucleotides are specifically contemplated for therapeutic use by the invention. The antisense oligonucleotides may be further modified by adding poly-L-lysine, transferrin polylysine, or cholesterol moieties at their 5' end. Suppression of nGPCR-x expression at either 15 the transcriptional or translational level is useful to generate cellular or animal models for diseases/conditions characterized by aberrant nGPCR-x expression.

Antisense oligonucleotides, or fragments of sequences selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, or sequences complementary or homologous thereto, derived from the nucleotide sequences of the present invention encoding nGPCR-x are useful as diagnostic tools for probing gene expression in various tissues. For example, tissue can 20 be probed *in situ* with oligonucleotide probes carrying detectable groups by conventional autoradiography techniques to investigate native expression of this enzyme or pathological conditions relating thereto. Antisense oligonucleotides are preferably directed to regulatory 25 regions of sequences selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, or mRNA corresponding thereto, including, but not limited to, the initiation codon, TATA box, enhancer sequences, and the like.

Transcription factors

The nGPCR-x sequences taught in the present invention facilitate the design of novel transcription factors for modulating nGPCR-x expression in native cells and animals, and cells transformed or transfected with nGPCR-x polynucleotides. For example, the Cys₂-His₂ zinc 30 finger proteins, which bind DNA via their zinc finger domains, have been shown to be amenable to structural changes that lead to the recognition of different target sequences. These artificial zinc finger proteins recognize specific target sites with high affinity and low dissociation constants, and are able to act as gene switches to modulate gene expression. Knowledge of the particular nGPCR-x target sequence of the present invention facilitates the engineering of zinc

finger proteins specific for the target sequence using known methods such as a combination of structure-based modeling and screening of phage display libraries (Segal *et al.*, *Proc. Natl. Acad. Sci. USA* 96:2758-2763 (1999); Liu *et al.*, *Proc. Natl. Acad. Sci. USA* 94:5525-5530 (1997); Greisman *et al.*, *Science* 275:657-661 (1997); Choo *et al.*, *J. Mol. Biol.* 273:525-532 (1997)). Each zinc finger domain usually recognizes three or more base pairs. Since a recognition sequence of 18 base pairs is generally sufficient in length to render it unique in any known genome, a zinc finger protein consisting of 6 tandem repeats of zinc fingers would be expected to ensure specificity for a particular sequence (Segal *et al.*). The artificial zinc finger repeats, designed based on nGPCR-x sequences, are fused to activation or repression domains to promote or suppress nGPCR-x expression (Liu *et al.*). Alternatively, the zinc finger domains can be fused to the TATA box-binding factor (TBP) with varying lengths of linker region between the zinc finger peptide and the TBP to create either transcriptional activators or repressors (Kim *et al.*, *Proc. Natl. Acad. Sci. USA* 94:3616-3620 (1997)). Such proteins and polynucleotides that encode them, have utility for modulating nGPCR-x expression *in vivo* in both native cells, animals and humans; and/or cells transfected with nGPCR-x-encoding sequences. The novel transcription factor can be delivered to the target cells by transfecting constructs that express the transcription factor (gene therapy), or by introducing the protein. Engineered zinc finger proteins can also be designed to bind RNA sequences for use in therapeutics as alternatives to antisense or catalytic RNA methods (McColl *et al.*, *Proc. Natl. Acad. Sci. USA* 96:9521-9526 (1997); Wu *et al.*, *Proc. Natl. Acad. Sci. USA* 92:344-348 (1995)). The present invention contemplates methods of designing such transcription factors based on the gene sequence of the invention, as well as customized zinc finger proteins, that are useful to modulate nGPCR-x expression in cells (native or transformed) whose genetic complement includes these sequences.

Polypeptides

The invention also provides purified and isolated mammalian nGPCR-x polypeptides encoded by a polynucleotide of the invention. Presently preferred is a human nGPCR-x polypeptide comprising the amino acid sequence set out in sequences selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, or fragments thereof comprising an epitope specific to the polypeptide. By "epitope specific to" is meant a portion of the nGPCR receptor that is recognizable by an antibody that is specific for the nGPCR, as defined in detail below.

Although the sequences provided are particular human sequences, the invention is intended to include within its scope other human allelic variants; non-human mammalian forms of nGPCR-x, and other vertebrate forms of nGPCR-x.

It will be appreciated that extracellular epitopes are particularly useful for generating and screening for antibodies and other binding compounds that bind to receptors such as nGPCR-x. Thus, in another preferred embodiment, the invention provides a purified and isolated polypeptide comprising at least one extracellular domain (e.g., the N-terminal extracellular domain or one of the three extracellular loops) of nGPCR-x. Purified and isolated polypeptides comprising the N-terminal extracellular domain of nGPCR-x are highly preferred. Also preferred is a purified and isolated polypeptide comprising a nGPCR-x fragment selected from the group consisting of the N-terminal extracellular domain of nGPCR-x, transmembrane domains of nGPCR-x, an extracellular loop connecting transmembrane domains of nGPCR-x, 5 an intracellular loop connecting transmembrane domains of nGPCR-x, the C-terminal cytoplasmic region of nGPCR-x, and fusions thereof. Such fragments may be continuous portions of the native receptor. However, it will also be appreciated that knowledge of the nGPCR-x gene and protein sequences as provided herein permits recombining of various domains that are not contiguous in the native protein. Using a FORTRAN computer program 10 called "tmrest.all" (Parodi *et al.*, *Comput. Appl. Biosci.* 5:527-535 (1994)), nGPCR-x was 15 shown to contain transmembrane-spanning domains.

The invention also embraces polypeptides that have at least 99%, at least 95%, at least 90%, at least 85%, at least 80%, at least 75%, at least 70%, at least 65%, at least 60%, at least 55% or at least 50% identity and/or homology to the preferred polypeptide of the invention. 20 Percent amino acid sequence "identity" with respect to the preferred polypeptide of the invention is defined herein as the percentage of amino acid residues in the candidate sequence that are identical with the residues in the nGPCR-x sequence after aligning both sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not 25 considering any conservative substitutions as part of the sequence identity. Percent sequence "homology" with respect to the preferred polypeptide of the invention is defined herein as the percentage of amino acid residues in the candidate sequence that are identical with the residues in the nGPCR-x sequence after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and also considering any conservative substitutions as part of the sequence identity.

30 In one aspect, percent homology is calculated as the percentage of amino acid residues in the smaller of two sequences which align with identical amino acid residue in the sequence being compared, when four gaps in a length of 100 amino acids may be introduced to maximize alignment (Dayhoff, in *Atlas of Protein Sequence and Structure*, Vol. 5, p. 124, National

Biochemical Research Foundation, Washington, D.C. (1972), incorporated herein by reference in its entirety).

5 Polypeptides of the invention may be isolated from natural cell sources or may be chemically synthesized, but are preferably produced by recombinant procedures involving host cells of the invention. Use of mammalian host cells is expected to provide for such post-translational modifications (*e.g.*, glycosylation, truncation, lipidation, and phosphorylation) as may be needed to confer optimal biological activity on recombinant expression products of the invention. Glycosylated and non-glycosylated forms of nGPCR-x polypeptides are embraced by the invention.

10 The invention also embraces variant (or analog) nGPCR-x polypeptides. In one example, insertion variants are provided wherein one or more amino acid residues supplement a nGPCR-x amino acid sequence. Insertions may be located at either or both termini of the protein, or may be positioned within internal regions of the nGPCR-x amino acid sequence. Insertional variants with additional residues at either or both termini can include, for example, fusion proteins and 15 proteins including amino acid tags or labels.

Insertion variants include nGPCR-x polypeptides wherein one or more amino acid residues are added to a nGPCR-x acid sequence or to a biologically active fragment thereof.

20 Variant products of the invention also include mature nGPCR-x products, *i.e.*, nGPCR-x products wherein leader or signal sequences are removed, with additional amino terminal residues. The additional amino terminal residues may be derived from another protein, or may include one or more residues that are not identifiable as being derived from specific proteins. 25 nGPCR-x products with an additional methionine residue at position -1 (Met⁻¹-nGPCR-x) are contemplated, as are variants with additional methionine and lysine residues at positions -2 and -1 (Met⁻²-Lys⁻¹-nGPCR-x). Variants of nGPCR-x with additional Met, Met-Lys, Lys residues (or one or more basic residues in general) are particularly useful for enhanced recombinant protein production in bacterial host cells.

30 The invention also embraces nGPCR-x variants having additional amino acid residues that result from use of specific expression systems. For example, use of commercially available vectors that express a desired polypeptide as part of a glutathione-S-transferase (GST) fusion product provides the desired polypeptide having an additional glycine residue at position -1 after cleavage of the GST component from the desired polypeptide. Variants that result from expression in other vector systems are also contemplated.

Insertional variants also include fusion proteins wherein the amino terminus and/or the carboxy terminus of nGPCR-x is/are fused to another polypeptide..

In another aspect, the invention provides deletion variants wherein one or more amino acid residues in a nGPCR-x polypeptide are removed. Deletions can be effected at one or both termini of the nGPCR-x polypeptide, or with removal of one or more non-terminal amino acid residues of nGPCR-x. Deletion variants, therefore, include all fragments of a nGPCR-x polypeptide.

5 The invention also embraces polypeptide fragments of sequences selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, wherein the fragments maintain biological (e.g., ligand binding and/or intracellular signaling) immunological properties of a nGPCR-x polypeptide.

10 In one preferred embodiment of the invention, an isolated nucleic acid molecule comprises a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence homologous to sequences selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, and fragments thereof, wherein the nucleic acid molecule encoding at least a portion of nGPCR-x. In a more preferred embodiment, the isolated nucleic acid molecule comprises a sequence that encodes a polypeptide comprising sequences selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, and fragments thereof.

15 20 As used in the present invention, polypeptide fragments comprise at least 5, 10, 15, 20, 25, 30, 35, or 40 consecutive amino acids of sequences selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120. Preferred polypeptide fragments display antigenic properties unique to, or specific for, human nGPCR-x and its allelic and species homologs. Fragments of the invention having the desired biological and immunological properties can be prepared by any of the methods well known and routinely practiced in the art.

25 In still another aspect, the invention provides substitution variants of nGPCR-x polypeptides. Substitution variants include those polypeptides wherein one or more amino acid residues of a nGPCR-x polypeptide are removed and replaced with alternative residues. In one aspect, the substitutions are conservative in nature; however, the invention embraces substitutions that are also non-conservative. Conservative substitutions for this purpose may be defined as set out in Tables 2, 3, or 4 below.

30 Variant polypeptides include those wherein conservative substitutions have been introduced by modification of polynucleotides encoding polypeptides of the invention. Amino acids can be classified according to physical properties and contribution to secondary and tertiary protein structure. A conservative substitution is recognized in the art as a substitution of one amino acid for another amino acid that has similar properties. Exemplary conservative

substitutions are set out in Table 2 (from WO 97/09433, page 10, published March 13, 1997 (PCT/GB96/02197, filed 9/6/96), immediately below.

5

Table 2
Conservative Substitutions I

<u>SIDE CHAIN CHARACTERISTIC</u>	<u>AMINO ACID</u>
Aliphatic	
Non-polar	G A P I L V
Polar - uncharged	C S T M N Q
Polar - charged	D E K R
Aromatic	H F W Y
Other	N Q D E

Alternatively, conservative amino acids can be grouped as described in Lehninger, (*Biochemistry*, Second Edition; Worth Publishers, Inc. NY, NY (1975), pp.71-77) as set out in Table 3, below.

10

Table 3
Conservative Substitutions II

<u>SIDE CHAIN CHARACTERISTIC</u>	<u>AMINO ACID</u>
Non-polar (hydrophobic)	
A. Aliphatic:	A L I V P
B. Aromatic:	F W
C. Sulfur-containing:	M
D. Borderline:	G
Uncharged-polar	
A. Hydroxyl:	S T Y
B. Amides:	N Q
C. Sulfhydryl:	C
D. Borderline:	G
Positively Charged (Basic):	K R H
Negatively Charged (Acidic):	D E

15

As still another alternative, exemplary conservative substitutions are set out in Table 4, below.

Table 4
Conservative Substitutions III

<u>Original Residue</u>	<u>Exemplary Substitution</u>
Ala (A)	Val, Leu, Ile

Arg (R)	Lys, Gln, Asn
Asn (N)	Gln, His, Lys, Arg
Asp (D)	Glu
Cys (C)	Ser
Gln (Q)	Asn
Glu (E)	Asp
His (H)	Asn, Gln, Lys, Arg
Ile (I)	Leu, Val, Met, Ala, Phe,
Leu (L)	Ile, Val, Met, Ala, Phe
Lys (K)	Arg, Gln, Asn
Met (M)	Leu, Phe, Ile
Phe (F)	Leu, Val, Ile, Ala
Pro (P)	Gly
Ser (S)	Thr
Thr (T)	Ser
Trp (W)	Tyr
Tyr (Y)	Trp, Phe, Thr, Ser
Val (V)	Ile, Leu, Met, Phe, Ala

It should be understood that the definition of polypeptides of the invention is intended to include polypeptides bearing modifications other than insertion, deletion, or substitution of amino acid residues. By way of example, the modifications may be covalent in nature, and include for example, chemical bonding with polymers, lipids, other organic, and inorganic 5 moieties. Such derivatives may be prepared to increase circulating half-life of a polypeptide, or may be designed to improve the targeting capacity of the polypeptide for desired cells, tissues, or organs. Similarly, the invention further embraces nGPCR-x polypeptides that have been covalently modified to include one or more water-soluble polymer attachments such as polyethylene glycol, polyoxyethylene glycol, or polypropylene glycol. Variants that display 10 ligand binding properties of native nGPCR-x and are expressed at higher levels, as well as variants that provide for constitutively active receptors, are particularly useful in assays of the invention; the variants are also useful in providing cellular, tissue and animal models of diseases/conditions characterized by aberrant nGPCR-x activity.

In a related embodiment, the present invention provides compositions comprising 15 purified polypeptides of the invention. Preferred compositions comprise, in addition to the polypeptide of the invention, a pharmaceutically acceptable (*i.e.*, sterile and non-toxic) liquid, semisolid, or solid diluent that serves as a pharmaceutical vehicle, excipient, or medium. Any diluent known in the art may be used. Exemplary diluents include, but are not limited to, water, saline solutions, polyoxyethylene sorbitan monolaurate, magnesium stearate, methyl- and

propylhydroxybenzoate, talc, alginates, starches, lactose, sucrose, dextrose, sorbitol, mannitol, glycerol, calcium phosphate, mineral oil, and cocoa butter.

5 Variants that display ligand binding properties of native nGPCR-x and are expressed at higher levels, as well as variants that provide for constitutively active receptors, are particularly useful in assays of the invention; the variants are also useful in assays of the invention and in providing cellular, tissue and animal models of diseases/conditions characterized by aberrant nGPCR-x activity.

10 The G protein-coupled receptor functions through a specific heterotrimeric guanine-nucleotide-binding regulatory protein (G-protein) coupled to the intracellular portion of the G protein-coupled receptor molecule. Accordingly, the G protein-coupled receptor has a specific affinity to G protein. G proteins specifically bind to guanine nucleotides. Isolation of G proteins provides a means to isolate guanine nucleotides. G Proteins may be isolated using commercially available anti-G protein antibodies or isolated G protein-coupled receptors. 15 Similarly, G proteins may be detected in a sample isolated using commercially available detectable anti-G protein antibodies or isolated G protein-coupled receptors.

According to the present invention, the isolated n-GPCR-x proteins of the present invention are useful to isolate and purify G proteins from samples such as cell lysates. Example 14 below sets forth an example of isolation of G proteins using isolated n-GPCR-x proteins. Such methodology may be used in place of the use of commercially available anti-G protein 20 antibodies which are used to isolate G proteins. Moreover, G proteins may be detected using n-GPCR-x proteins in place of commercially available detectable anti-G protein antibodies. Since n-GPCR-x proteins specifically bind to G proteins, they can be employed in any specific use where G protein specific affinity is required such as those uses where commercially available anti-G protein antibodies are employed.

25 **Antibodies**

Also comprehended by the present invention are antibodies (e.g., monoclonal and polyclonal antibodies, single chain antibodies, chimeric antibodies, bifunctional/bispecific antibodies, humanized antibodies, human antibodies, and complementary determining region (CDR)-grafted antibodies, including compounds which include CDR sequences which 30 specifically recognize a polypeptide of the invention) specific for nGPCR-x or fragments thereof. Preferred antibodies of the invention are human antibodies that are produced and identified according to methods described in WO93/11236, published June 20, 1993, which is incorporated herein by reference in its entirety. Antibody fragments, including Fab, Fab', F(ab')₂, and F_v, are also provided by the invention. The term "specific for," when used to

describe antibodies of the invention, indicates that the variable regions of the antibodies of the invention recognize and bind nGPCR-x polypeptides exclusively (*i.e.*, are able to distinguish nGPCR-x polypeptides from other known GPCR polypeptides by virtue of measurable differences in binding affinity, despite the possible existence of localized sequence identity, 5 homology, or similarity between nGPCR-x and such polypeptides). It will be understood that specific antibodies may also interact with other proteins (for example, *S. aureus* protein A or other antibodies in ELISA techniques) through interactions with sequences outside the variable region of the antibodies, and, in particular, in the constant region of the molecule. Screening assays to determine binding specificity of an antibody of the invention are well known and 10 routinely practiced in the art. For a comprehensive discussion of such assays, see Harlow *et al.* (Eds.), *Antibodies A Laboratory Manual*; Cold Spring Harbor Laboratory; Cold Spring Harbor, NY (1988), Chapter 6. Antibodies that recognize and bind fragments of the nGPCR-x polypeptides of the invention are also contemplated, provided that the antibodies are specific for nGPCR-x polypeptides. Antibodies of the invention can be produced using any method well 15 known and routinely practiced in the art.

The invention provides an antibody that is specific for the nGPCR-x of the invention. Antibody specificity is described in greater detail below. However, it should be emphasized that antibodies that can be generated from polypeptides that have previously been described in the literature and that are capable of fortuitously cross-reacting with nGPCR-x (*e.g.*, due to the 20 fortuitous existence of a similar epitope in both polypeptides) are considered “cross-reactive” antibodies. Such cross-reactive antibodies are not antibodies that are “specific” for nGPCR-x. The determination of whether an antibody is specific for nGPCR-x or is cross-reactive with another known receptor is made using any of several assays, such as Western blotting assays, that are well known in the art. For identifying cells that express nGPCR-x and also for 25 modulating nGPCR-x-ligand binding activity, antibodies that specifically bind to an extracellular epitope of the nGPCR-x are preferred.

In one preferred variation, the invention provides monoclonal antibodies. Hybridomas that produce such antibodies also are intended as aspects of the invention. In yet another variation, the invention provides a humanized antibody. Humanized antibodies are useful for *in* 30 *vivo* therapeutic indications.

In another variation, the invention provides a cell-free composition comprising polyclonal antibodies, wherein at least one of the antibodies is an antibody of the invention specific for nGPCR-x. Antisera isolated from an animal is an exemplary composition, as is a

composition comprising an antibody fraction of an antisera that has been resuspended in water or in another diluent, excipient, or carrier.

In still another related embodiment, the invention provides an anti-idiotypic antibody specific for an antibody that is specific for nGPCR-x.

5 It is well known that antibodies contain relatively small antigen binding domains that can be isolated chemically or by recombinant techniques. Such domains are useful nGPCR-x binding molecules themselves, and also may be reintroduced into human antibodies, or fused to toxins or other polypeptides. Thus, in still another embodiment, the invention provides a 10 polypeptide comprising a fragment of a nGPCR-x-specific antibody, wherein the fragment and the polypeptide bind to the nGPCR-x. By way of non-limiting example, the invention provides polypeptides that are single chain antibodies and CDR-grafted antibodies.

15 Non-human antibodies may be humanized by any of the methods known in the art. In one method, the non-human CDRs are inserted into a human antibody or consensus antibody framework sequence. Further changes can then be introduced into the antibody framework to modulate affinity or immunogenicity.

20 Antibodies of the invention are useful for, e.g., therapeutic purposes (by modulating activity of nGPCR-x), diagnostic purposes to detect or quantitate nGPCR-x, and purification of nGPCR-x. Kits comprising an antibody of the invention for any of the purposes described herein are also comprehended. In general, a kit of the invention also includes a control antigen for which the antibody is immunospecific.

Compositions

Mutations in the nGPCR-x gene that result in loss of normal function of the nGPCR-x gene product underlie nGPCR-x-related human disease states. The invention comprehends gene 25 therapy to restore nGPCR-x activity to treat those disease states. Delivery of a functional nGPCR-x gene to appropriate cells is effected *ex vivo*, *in situ*, or *in vivo* by use of vectors, and more particularly viral vectors (e.g., adenovirus, adeno-associated virus, or a retrovirus), or *ex vivo* by use of physical DNA transfer methods (e.g., liposomes or chemical treatments). See, for example, Anderson, *Nature*, supplement to vol. 392, no. 6679, pp.25-20 (1998). For additional reviews of gene therapy technology see Friedmann, *Science*, 244: 1275-1281 (1989); Verma, 30 *Scientific American*: 68-84 (1990); and Miller, *Nature*, 357: 455-460 (1992). Alternatively, it is contemplated that in other human disease states, preventing the expression of, or inhibiting the activity of, nGPCR-x will be useful in treating disease states. It is contemplated that antisense therapy or gene therapy could be applied to negatively regulate the expression of nGPCR-x.

Another aspect of the present invention is directed to compositions, including pharmaceutical compositions, comprising any of the nucleic acid molecules or recombinant expression vectors described above and an acceptable carrier or diluent. Preferably, the carrier or diluent is pharmaceutically acceptable. Suitable carriers are described in the most recent 5 edition of *Remington's Pharmaceutical Sciences*, A. Osol, a standard reference text in this field, which is incorporated herein by reference in its entirety. Preferred examples of such carriers or diluents include, but are not limited to, water, saline, Ringer's solution, dextrose solution, and 5% human serum albumin. Liposomes and nonaqueous vehicles such as fixed oils may also be used. The formulations are sterilized by commonly used techniques.

10 Also within the scope of the invention are compositions comprising polypeptides, polynucleotides, or antibodies of the invention that have been formulated with, *e.g.*, a pharmaceutically acceptable carrier.

15 The invention also provides methods of using antibodies of the invention. For example, the invention provides a method for modulating ligand binding of a nGPCR-x comprising the step of contacting the nGPCR-x with an antibody specific for the nGPCR-x, under conditions wherein the antibody binds the receptor.

20 GPCRs that may be expressed in the brain, such as nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70, provide an indication that aberrant nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 signaling activity may correlate with one or more neurological or psychological disorders. The invention also provides a method for treating a neurological or psychiatric disorder comprising the step of administering to a mammal in need of such treatment an amount of an antibody-like 25 polypeptide of the invention that is sufficient to modulate ligand binding to a nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 in neurons of the mammal. nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 may also be expressed in other tissues, including but not limited to, peripheral blood lymphocytes, pancreas, ovary, uterus, testis, salivary gland, thyroid gland, kidney, adrenal gland, liver, bone marrow, prostate, fetal liver, colon, muscle, and fetal brain, and may be found in many other tissues. Within the brain, nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 mRNA transcripts may be found in many tissues, including, but not limited to, frontal lobe, hypothalamus, pons, cerebellum, caudate nucleus, and medulla. Tissues and brain regions where 30 specific nGPCRs of the present invention are expressed are identified in the Examples below.

Kits

The present invention is also directed to kits, including pharmaceutical kits. The kits can comprise any of the nucleic acid molecules described above, any of the polypeptides described above, or any antibody which binds to a polypeptide of the invention as described above, as well

as a negative control. The kit preferably comprises additional components, such as, for example, instructions, solid support, reagents helpful for quantification, and the like.

In another aspect, the invention features methods for detection of a polypeptide in a sample as a diagnostic tool for diseases or disorders, wherein the method comprises the steps of:

5 (a) contacting the sample with a nucleic acid probe which hybridizes under hybridization assay conditions to a nucleic acid target region of a polypeptide having sequences selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120, said probe comprising the nucleic acid sequence encoding the polypeptide, fragments thereof, and the complements of the sequences and fragments; and (b) detecting the presence or amount of the probe:target region hybrid as an
10 indication of the disease.

In preferred embodiments of the invention, the disease is selected from the group consisting of thyroid disorders (e.g. thyreotoxicosis, myxoedema); renal failure; inflammatory conditions (e.g., Crohn's disease); diseases related to cell differentiation and homeostasis; rheumatoid arthritis; autoimmune disorders; movement disorders; CNS disorders (e.g., pain
15 including migraine; stroke; psychotic and neurological disorders, including anxiety, mental disorder, manic depression, anxiety, generalized anxiety disorder, post-traumatic-stress disorder, depression, bipolar disorder, delirium, dementia, severe mental retardation; dyskinesias, such as Huntington's disease or Tourette's Syndrome; attention disorders including attention deficit disorder (ADD) and attention deficit-hyperactivity disorder (ADHD), and degenerative
20 disorders such as Parkinson's, Alzheimer's; movement disorders, including ataxias, supranuclear palsy, etc.); infections, such as viral infections caused by HIV-1 or HIV-2; metabolic and cardiovascular diseases and disorders (e.g., type 2 diabetes, impaired glucose tolerance, dyslipidemia, obesity, anorexia, hypotension, hypertension, thrombosis, myocardial infarction, cardiomyopathies, atherosclerosis, etc.); proliferative diseases and cancers (e.g.,
25 different cancers such as breast, colon, lung, etc., and hyperproliferative disorders such as psoriasis, prostate hyperplasia, etc.); hormonal disorders (e.g., male/female hormonal replacement, polycystic ovarian syndrome, alopecia, etc.); and sexual dysfunction, among others.

As described above and in Example 5 below, the genes encoding nGPCR-42, 46, 48, 49, 30 51, 52, 61, 63, and 70 have been detected in brain tissue indicating that these n-GPCR-x proteins are neuroreceptors. Kits may be designed to detect either expression of polynucleotides encoding these proteins or the proteins themselves in order to identify tissue as being neurological. For example, oligonucleotide hybridization kits can be provided which include a container having an oligonucleotide probe specific for the n-GPCR-x-specific DNA and

optionally, containers with positive and negative controls and/or instructions. Similarly, PCR kits can be provided which include a container having primers specific for the n-GPCR-x-specific sequences, DNA and optionally, containers with size markers, positive and negative controls and/or instructions.

5 Hybridization conditions should be such that hybridization occurs only with the genes in the presence of other nucleic acid molecules. Under stringent hybridization conditions only highly complementary nucleic acid sequences hybridize. Preferably, such conditions prevent hybridization of nucleic acids having 1 or 2 mismatches out of 20 contiguous nucleotides. Such conditions are defined *supra*.

10 The diseases for which detection of genes in a sample could be diagnostic include diseases in which nucleic acid (DNA and/or RNA) is amplified in comparison to normal cells. By "amplification" is meant increased numbers of DNA or RNA in a cell compared with normal cells.

15 The diseases that could be diagnosed by detection of nucleic acid in a sample preferably include central nervous system and metabolic diseases. The test samples suitable for nucleic acid probing methods of the present invention include, for example, cells or nucleic acid extracts of cells, or biological fluids. The samples used in the above-described methods will vary based on the assay format, the detection method and the nature of the tissues, cells or extracts to be assayed. Methods for preparing nucleic acid extracts of cells are well known in the art and can 20 be readily adapted in order to obtain a sample that is compatible with the method utilized.

Alternatively, immunoassay kits can be provided which have containers container having antibodies specific for the n-GPCR-x-protein and optionally, containers with positive and negative controls and/or instructions.

25 Kits may also be provided useful in the identification of GPCR binding partners such as natural ligands or modulators (agonists or antagonists). Substances useful for treatment of disorders or diseases preferably show positive results in one or more *in vitro* assays for an activity corresponding to treatment of the disease or disorder in question. Substances that modulate the activity of the polypeptides preferably include, but are not limited to, antisense oligonucleotides, agonists and antagonists, and inhibitors of protein kinases.

30 **Methods of inducing immune response**

Another aspect of the present invention is directed to methods of inducing an immune response in a mammal against a polypeptide of the invention by administering to the mammal an amount of the polypeptide sufficient to induce an immune response. The amount will be

dependent on the animal species, size of the animal, and the like but can be determined by those skilled in the art.

Methods of identifying ligands

The invention also provides assays to identify compounds that bind nGPCR-x. One such 5 assay comprises the steps of: (a) contacting a composition comprising a nGPCR-x with a compound suspected of binding nGPCR-x; and (b) measuring binding between the compound and nGPCR-x. In one variation, the composition comprises a cell expressing nGPCR-x on its surface. In another variation, isolated nGPCR-x or cell membranes comprising nGPCR-x are employed. The binding may be measured directly, *e.g.*, by using a labeled compound, or may be 10 measured indirectly by several techniques, including measuring intracellular signaling of nGPCR-x induced by the compound (or measuring changes in the level of nGPCR-x signaling). Following steps (a) and (b), compounds identified as binding nGPCR-x can be further tested in other assays including, but not limited to, *in vivo* models, in order to confirm or quantitate binding to nGPCR-x.

15 Specific binding molecules, including natural ligands and synthetic compounds, can be identified or developed using isolated or recombinant nGPCR-x products, nGPCR-x variants, or preferably, cells expressing such products. Binding partners are useful for purifying nGPCR-x products and detection or quantification of nGPCR-x products in fluid and tissue samples using known immunological procedures. Binding molecules are also manifestly useful in modulating 20 (*i.e.*, blocking, inhibiting or stimulating) biological activities of nGPCR-x, especially those activities involved in signal transduction.

The DNA and amino acid sequence information provided by the present invention also makes possible identification of binding partner compounds with which a nGPCR-x polypeptide or polynucleotide will interact. Methods to identify binding partner compounds include solution 25 assays, *in vitro* assays wherein nGPCR-x polypeptides are immobilized, and cell-based assays. Identification of binding partner compounds of nGPCR-x polypeptides provides candidates for therapeutic or prophylactic intervention in pathologies associated with nGPCR-x normal and aberrant biological activity.

The invention includes several assay systems for identifying nGPCR-x binding partners. 30 In solution assays, methods of the invention comprise the steps of (a) contacting a nGPCR-x polypeptide with one or more candidate binding partner compounds and (b) identifying the compounds that bind to the nGPCR-x polypeptide. Identification of the compounds that bind the nGPCR-x polypeptide can be achieved by isolating the nGPCR-x polypeptide/binding partner complex, and separating the binding partner compound from the nGPCR-x polypeptide.

An additional step of characterizing the physical, biological, and/or biochemical properties of the binding partner compound is also comprehended in another embodiment of the invention, wherein compounds identified as binding nGPCR-x can be further tested in other assays including, but not limited to, *in vivo* models, in order to confirm or quantitate binding to 5 nGPCR-x. In one aspect, the nGPCR-x polypeptide/binding partner complex is isolated using an antibody immunospecific for either the nGPCR-x polypeptide or the candidate binding partner compound.

In still other embodiments, either the nGPCR-x polypeptide or the candidate binding partner compound comprises a label or tag that facilitates its isolation, and methods of the 10 invention to identify binding partner compounds include a step of isolating the nGPCR-x polypeptide/binding partner complex through interaction with the label or tag. An exemplary tag of this type is a poly-histidine sequence, generally around six histidine residues, that permits isolation of a compound so labeled using nickel chelation. Other labels and tags, such as the FLAG® tag (Eastman Kodak, Rochester, NY), well known and routinely used in the art, are 15 embraced by the invention.

In one variation of an *in vitro* assay, the invention provides a method comprising the steps of (a) contacting an immobilized nGPCR-x polypeptide with a candidate binding partner compound and (b) detecting binding of the candidate compound to the nGPCR-x polypeptide. In an alternative embodiment, the candidate binding partner compound is immobilized and binding 20 of nGPCR-x is detected. Immobilization is accomplished using any of the methods well known in the art, including covalent bonding to a support, a bead, or a chromatographic resin, as well as non-covalent, high affinity interactions such as antibody binding, or use of streptavidin/biotin binding wherein the immobilized compound includes a biotin moiety. Detection of binding can be accomplished (i) using a radioactive label on the compound that is not immobilized, (ii) using 25 of a fluorescent label on the non-immobilized compound, (iii) using an antibody immunospecific for the non-immobilized compound, (iv) using a label on the non-immobilized compound that excites a fluorescent support to which the immobilized compound is attached, as well as other techniques well known and routinely practiced in the art.

The invention also provides cell-based assays to identify binding partner compounds of a 30 nGPCR-x polypeptide. In one embodiment, the invention provides a method comprising the steps of contacting a nGPCR-x polypeptide expressed on the surface of a cell with a candidate binding partner compound and detecting binding of the candidate binding partner compound to the nGPCR-x polypeptide. In a preferred embodiment, the detection comprises detecting a calcium flux or other physiological event in the cell caused by the binding of the molecule.

Another aspect of the present invention is directed to methods of identifying compounds that bind to either nGPCR-x or nucleic acid molecules encoding nGPCR-x, comprising contacting nGPCR-x, or a nucleic acid molecule encoding the same, with a compound, and determining whether the compound binds nGPCR-x or a nucleic acid molecule encoding the same. Binding can be determined by binding assays which are well known to the skilled artisan, including, but not limited to, gel-shift assays, Western blots, radiolabeled competition assay, phage-based expression cloning, co-fractionation by chromatography, co-precipitation, cross linking, interaction trap/two-hybrid analysis, southwestern analysis, ELISA, and the like, which are described in, for example, *Current Protocols in Molecular Biology*, 1999, John Wiley & Sons, NY, which is incorporated herein by reference in its entirety. The compounds to be screened include (which may include compounds which are suspected to bind nGPCR-x, or a nucleic acid molecule encoding the same), but are not limited to, extracellular, intracellular, biologic or chemical origin. The methods of the invention also embrace ligands, especially neuropeptides, that are attached to a label, such as a radiolabel (e.g., ^{125}I , ^{35}S , ^{32}P , ^{33}P , ^3H), a fluorescence label, a chemiluminescent label, an enzymic label and an immunogenic label. Modulators falling within the scope of the invention include, but are not limited to, non-peptide molecules such as non-peptide mimetics, non-peptide allosteric effectors, and peptides. The nGPCR-x polypeptide or polynucleotide employed in such a test may either be free in solution, attached to a solid support, borne on a cell surface or located intracellularly or associated with a portion of a cell. One skilled in the art can, for example, measure the formation of complexes between nGPCR-x and the compound being tested. Alternatively, one skilled in the art can examine the diminution in complex formation between nGPCR-x and its substrate caused by the compound being tested.

In another embodiment of the invention, high throughput screening for compounds having suitable binding affinity to nGPCR-x is employed. Briefly, large numbers of different test compounds are synthesized on a solid substrate. The peptide test compounds are contacted with nGPCR-x and washed. Bound nGPCR-x is then detected by methods well known in the art. Purified polypeptides of the invention can also be coated directly onto plates for use in the aforementioned drug screening techniques. In addition, non-neutralizing antibodies can be used to capture the protein and immobilize it on the solid support.

Generally, an expressed nGPCR-x can be used for HTS binding assays in conjunction with its defined ligand, in this case the corresponding neuropeptide that activates it. The identified peptide is labeled with a suitable radioisotope, including, but not limited to, ^{125}I , ^3H , ^{35}S or ^{32}P , by methods that are well known to those skilled in the art. Alternatively, the peptides

may be labeled by well-known methods with a suitable fluorescent derivative (Baindur *et al.*, *Drug Dev. Res.*, 1994, 33, 373-398; Rogers, *Drug Discovery Today*, 1997, 2, 156-160).

Radioactive ligand specifically bound to the receptor in membrane preparations made from the cell line expressing the recombinant protein can be detected in HTS assays in one of several standard ways, including filtration of the receptor-ligand complex to separate bound ligand from unbound ligand (Williams, *Med. Res. Rev.*, 1991, 11, 147-184; Sweetnam *et al.*, *J. Natural Products*, 1993, 56, 441-455). Alternative methods include a scintillation proximity assay (SPA) or a FlashPlate format in which such separation is unnecessary (Nakayama, *Cur. Opinion Drug Disc. Dev.*, 1998, 1, 85-91 Bossé *et al.*, *J. Biomolecular Screening*, 1998, 3, 285-292.). Binding of fluorescent ligands can be detected in various ways, including fluorescence energy transfer (FRET), direct spectrophotofluorometric analysis of bound ligand, or fluorescence polarization (Rogers, *Drug Discovery Today*, 1997, 2, 156-160; Hill, *Cur. Opinion Drug Disc. Dev.*, 1998, 1, 92-97).

Other assays may be used to identify specific ligands of a nGPCR-x receptor, including assays that identify ligands of the target protein through measuring direct binding of test ligands to the target protein, as well as assays that identify ligands of target proteins through affinity ultrafiltration with ion spray mass spectroscopy/HPLC methods or other physical and analytical methods. Alternatively, such binding interactions are evaluated indirectly using the yeast two-hybrid system described in Fields *et al.*, *Nature*, 340:245-246 (1989), and Fields *et al.*, *Trends in Genetics*, 10:286-292 (1994), both of which are incorporated herein by reference in its entirety. The two-hybrid system is a genetic assay for detecting interactions between two proteins or polypeptides. It can be used to identify proteins that bind to a known protein of interest, or to delineate domains or residues critical for an interaction. Variations on this methodology have been developed to clone genes that encode DNA binding proteins, to identify peptides that bind to a protein, and to screen for drugs. The two-hybrid system exploits the ability of a pair of interacting proteins to bring a transcription activation domain into close proximity with a DNA binding domain that binds to an upstream activation sequence (UAS) of a reporter gene, and is generally performed in yeast. The assay requires the construction of two hybrid genes encoding (1) a DNA-binding domain that is fused to a first protein and (2) an activation domain fused to a second protein. The DNA-binding domain targets the first hybrid protein to the UAS of the reporter gene; however, because most proteins lack an activation domain, this DNA-binding hybrid protein does not activate transcription of the reporter gene. The second hybrid protein, which contains the activation domain, cannot by itself activate expression of the reporter gene because it does not bind the UAS. However, when both hybrid proteins are present, the

noncovalent interaction of the first and second proteins tethers the activation domain to the UAS, activating transcription of the reporter gene. For example, when the first protein is a GPCR gene product, or fragment thereof, that is known to interact with another protein or nucleic acid, this assay can be used to detect agents that interfere with the binding interaction.

5 Expression of the reporter gene is monitored as different test agents are added to the system. The presence of an inhibitory agent results in lack of a reporter signal.

The yeast two-hybrid assay can also be used to identify proteins that bind to the gene product. In an assay to identify proteins that bind to a nGPCR-x receptor, or fragment thereof, a fusion polynucleotide encoding both a nGPCR-x receptor (or fragment) and a UAS binding domain (*i.e.*, a first protein) may be used. In addition, a large number of hybrid genes each 10 encoding a different second protein fused to an activation domain are produced and screened in the assay. Typically, the second protein is encoded by one or more members of a total cDNA or genomic DNA fusion library, with each second protein-coding region being fused to the activation domain. This system is applicable to a wide variety of proteins, and it is not even 15 necessary to know the identity or function of the second binding protein. The system is highly sensitive and can detect interactions not revealed by other methods; even transient interactions may trigger transcription to produce a stable mRNA that can be repeatedly translated to yield the reporter protein.

Other assays may be used to search for agents that bind to the target protein. One such 20 screening method to identify direct binding of test ligands to a target protein is described in U.S. Patent No. 5,585,277, incorporated herein by reference in its entirety. This method relies on the principle that proteins generally exist as a mixture of folded and unfolded states, and continually alternate between the two states. When a test ligand binds to the folded form of a target protein 25 (*i.e.*, when the test ligand is a ligand of the target protein), the target protein molecule bound by the ligand remains in its folded state. Thus, the folded target protein is present to a greater extent in the presence of a test ligand which binds the target protein, than in the absence of a ligand. Binding of the ligand to the target protein can be determined by any method that distinguishes 30 between the folded and unfolded states of the target protein. The function of the target protein need not be known in order for this assay to be performed. Virtually any agent can be assessed by this method as a test ligand, including, but not limited to, metals, polypeptides, proteins, lipids, polysaccharides, polynucleotides and small organic molecules.

Another method for identifying ligands of a target protein is described in Wieboldt *et al.*, *Anal. Chem.*, 69:1683-1691 (1997), incorporated herein by reference in its entirety. This technique screens combinatorial libraries of 20-30 agents at a time in solution phase for binding

to the target protein. Agents that bind to the target protein are separated from other library components by simple membrane washing. The specifically selected molecules that are retained on the filter are subsequently liberated from the target protein and analyzed by HPLC and pneumatically assisted electrospray (ion spray) ionization mass spectroscopy. This procedure 5 selects library components with the greatest affinity for the target protein, and is particularly useful for small molecule libraries.

Determining whether a test compound binds to nGPCR-51 can also be accomplished by measuring the intrinsic fluorescence of nGPCR-51 and determining whether the intrinsic fluorescence is modulated in the presence of the test compound. Preferably, the intrinsic 10 fluorescence of nGPCR-51 is measured as a function of the tryptophan residue(s) of nGPCR-51. Preferably, fluorescence of nGPCR-51 is measured and compared to the fluorescence intensity of nGPCR-51 in the presence of the test compound, wherein a decrease in fluorescence intensity indicates binding of the test compound to nGPCR-51. Preferred methodology is set forth in "Principles of Fluorescence Spectroscopy" by Joseph R. Lakowicz, New York, Plenum Press, 15 1983 (ISBN 0306412853) and "Spectrophotometry And Spectrofluorometry" by C.L. Bashford and D.A. Harris Oxford, Washington DC, IRL Press, 1987 (ISBN 0947946691), each of which is incorporated herein by reference in its entirety.

Other embodiments of the invention comprise using competitive screening assays in which neutralizing antibodies capable of binding a polypeptide of the invention specifically 20 compete with a test compound for binding to the polypeptide. In this manner, the antibodies can be used to detect the presence of any peptide that shares one or more antigenic determinants with nGPCR-x. Radiolabeled competitive binding studies are described in A.H. Lin *et al.* *Antimicrobial Agents and Chemotherapy*, 1997, vol. 41, no. 10. pp. 2127-2131, the disclosure of which is incorporated herein by reference in its entirety.

25 Another aspect of the present invention relates to methods of identifying a compound that binds to or modulates nGPCR-51. The methods comprise contacting a composition comprising nGPCR-51 and Peptide A with a test compound, or a plurality of test compounds, and determining whether the test compound competes with Peptide A for binding to nGPCR-51. A decrease in the amount of the complex between Peptide A, or a protein homologous thereto, 30 and nGPCR-51 in the presence of a test compound or compounds confirms that the compound or compounds binds to nGPCR-51. In some embodiments, the affinity or displacement of Peptide A is measured, wherein a low affinity indicates that the test compound interacts with nGPCR-51. In these methods, the composition that comprises nGPCR-51 and Peptide A can be cells. Compounds identified as binding to nGPCR-51 are also expected to modulate nGPCR-51

activity. Binding of a test compound to nGPCR-51 can be determined by any of the binding assays described above.

As described above and in Example 5 below, the genes encoding nGPCR-42, 46, 48, 49, 51, 52, 61, 63, and 70 have been detected in brain tissue indicating that these n-GPCR-x proteins are neuroreceptors. Accordingly, natural binding partners of these molecules include 5 neurotransmitters.

Identification of modulating agents

The invention also provides methods for identifying a modulator of binding between a nGPCR-x and a nGPCR-x binding partner, comprising the steps of: (a) contacting a nGPCR-x binding partner and a composition comprising a nGPCR-x in the presence and in the absence of a putative modulator compound; (b) detecting binding between the binding partner and the nGPCR-x; and (c) identifying a putative modulator compound or a modulator compound in view of decreased or increased binding between the binding partner and the nGPCR-x in the presence of the putative modulator, as compared to binding in the absence of the putative modulator.

Following steps (a) and (b), compounds identified as modulating binding between nGPCR-x and an nGPCR-x binding partner can be further tested in other assays including, but not limited to, *in vivo* models, in order to confirm or quantitate modulation of binding to nGPCR-x.

nGPCR-x binding partners that stimulate nGPCR-x activity are useful as agonists in disease states or conditions characterized by insufficient nGPCR-x signaling (e.g., as a result of insufficient activity of a nGPCR-x ligand). nGPCR-x binding partners that block ligand-mediated nGPCR-x signaling are useful as nGPCR-x antagonists to treat disease states or conditions characterized by excessive nGPCR-x signaling. In addition nGPCR-x modulators in general, as well as nGPCR-x polynucleotides and polypeptides, are useful in diagnostic assays for such diseases or conditions.

In another aspect, the invention provides methods for treating a disease or abnormal condition by administering to a patient in need of such treatment a substance that modulates the activity or expression of a polypeptide having sequences selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120.

Agents that modulate (*i.e.*, increase, decrease, or block) nGPCR-x activity or expression may be identified by incubating a putative modulator with a cell containing a nGPCR-x polypeptide or polynucleotide and determining the effect of the putative modulator on nGPCR-x activity or expression. The selectivity of a compound that modulates the activity of nGPCR-x can be evaluated by comparing its effects on nGPCR-x to its effect on other GPCR compounds. Following identification of compounds that modulate nGPCR-x activity or expression, such

compounds can be further tested in other assays including, but not limited to, *in vivo* models, in order to confirm or quantitate their activity. Selective modulators may include, for example, antibodies and other proteins, peptides, or organic molecules that specifically bind to a nGPCR-x polypeptide or a nGPCR-x-encoding nucleic acid. Modulators of nGPCR-x activity will be 5 therapeutically useful in treatment of diseases and physiological conditions in which normal or aberrant nGPCR-x activity is involved. nGPCR-x polynucleotides, polypeptides, and modulators may be used in the treatment of such diseases and conditions as infections, such as viral infections caused by HIV-1 or HIV-2; pain; cancers; metabolic and cardiovascular diseases and disorders (e.g., type 2 diabetes, impaired glucose tolerance, dyslipidemia, obesity, anorexia, 10 hypotension, hypertension, thrombosis, myocardial infarction, cardiomyopathies, atherosclerosis, etc.); Parkinson's disease; and psychotic and neurological disorders, including anxiety, mental disorder, manic depression, schizophrenia, migraine, major depression, attention disorders including ADD and ADHD, delirium, dementia, severe mental retardation and dyskinesias, such as Huntington's disease or Tourette's Syndrome, among others. nGPCR-x 15 polynucleotides and polypeptides, as well as nGPCR-x modulators, may also be used in diagnostic assays for such diseases or conditions.

Methods of the invention to identify modulators include variations on any of the methods described above to identify binding partner compounds, the variations including techniques wherein a binding partner compound has been identified and the binding assay is carried out in 20 the presence and absence of a candidate modulator. A modulator is identified in those instances where binding between the nGPCR-x polypeptide and the binding partner compound changes in the presence of the candidate modulator compared to binding in the absence of the candidate modulator compound. A modulator that increases binding between the nGPCR-x polypeptide and the binding partner compound is described as an enhancer or activator, and a modulator that 25 decreases binding between the nGPCR-x polypeptide and the binding partner compound is described as an inhibitor. Following identification of modulators, such compounds can be further tested in other assays including, but not limited to, *in vivo* models, in order to confirm or quantitate their activity as modulators.

The invention also comprehends high-throughput screening (HTS) assays to identify 30 compounds that interact with or inhibit biological activity (i.e., affect enzymatic activity, binding activity, etc.) of a nGPCR-x polypeptide. HTS assays permit screening of large numbers of compounds in an efficient manner. Cell-based HTS systems are contemplated to investigate nGPCR-x receptor-ligand interaction. HTS assays are designed to identify "hits" or "lead compounds" having the desired property, from which modifications can be designed to improve

the desired property. Chemical modification of the "hit" or "lead compound" is often based on an identifiable structure/activity relationship between the "hit" and the nGPCR-x polypeptide.

Another aspect of the present invention is directed to methods of identifying compounds which modulate (*i.e.*, increase or decrease) activity of nGPCR-x comprising contacting nGPCR-

5 x with a compound, and determining whether the compound modifies activity of nGPCR-x. The activity in the presence of the test compared is measured to the activity in the absence of the test compound. Where the activity of the sample containing the test compound is higher than the activity in the sample lacking the test compound, the compound will have increased activity.

10 Similarly, where the activity of the sample containing the test compound is lower than the activity in the sample lacking the test compound, the compound will have inhibited activity.

Following identification of compounds that modulate an activity of nGPCR-x, such compounds can be further tested in other assays including, but not limited to, *in vivo* models, in order to confirm or quantitate their activity.

15 The present invention is particularly useful for screening compounds by using nGPCR-x in any of a variety of drug screening techniques. The compounds to be screened include (which may include compounds which are suspected to modulate nGPCR-x activity), but are not limited to, extracellular, intracellular, biologic or chemical origin. The nGPCR-x polypeptide employed in such a test may be in any form, preferably, free in solution, attached to a solid support, borne on a cell surface or located intracellularly. One skilled in the art can, for example, measure the 20 formation of complexes between nGPCR-x and the compound being tested. Alternatively, one skilled in the art can examine the diminution in complex formation between nGPCR-x and its substrate caused by the compound being tested.

25 The activity of nGPCR-x polypeptides of the invention can be determined by, for example, examining the ability to bind or be activated by chemically synthesized peptide ligands. Alternatively, the activity of nGPCR-x polypeptides can be assayed by examining their ability to bind calcium ions, hormones, chemokines, neuropeptides, neurotransmitters, nucleotides, lipids, odorants, and photons. Alternatively, the activity of the nGPCR-x polypeptides can be determined by examining the activity of effector molecules including, but not limited to, adenylate cyclase, phospholipases and ion channels. Thus, modulators of 30 nGPCR-x polypeptide activity may alter a GPCR receptor function, such as a binding property of a receptor or an activity such as G protein-mediated signal transduction or membrane localization. In various embodiments of the method, the assay may take the form of an ion flux assay, a yeast growth assay, a non-hydrolyzable GTP assay such as a [³⁵S]-GTP γ S assay, a cAMP assay, an inositol triphosphate assay, a diacylglycerol assay, an Aequorin assay, a

Luciferase assay, a FLIPR assay for intracellular Ca^{2+} concentration, a mitogenesis assay, a MAP Kinase activity assay, an arachidonic acid release assay (e.g., using [^3H]-arachidonic acid), and an assay for extracellular acidification rates, as well as other binding or function-based assays of nGPCR-x activity that are generally known in the art. In several of these 5 embodiments, the invention comprehends the inclusion of any of the G proteins known in the art, such as G_{16} , G_{15} , or chimeric G_{qd5} , G_{qs5} , G_{qo5} , G_{q25} , and the like. nGPCR-x activity can be determined by methodologies that are used to assay for FaRP activity, which is well known to those skilled in the art. Biological activities of nGPCR-x receptors according to the invention include, but are not limited to, the binding of a natural or an unnatural ligand, as well as any one 10 of the functional activities of GPCRs known in the art. Non-limiting examples of GPCR activities include transmembrane signaling of various forms, which may involve G protein association and/or the exertion of an influence over G protein binding of various guanidylate nucleotides; another exemplary activity of GPCRs is the binding of accessory proteins or polypeptides that differ from known G proteins.

15 The modulators of the invention exhibit a variety of chemical structures, which can be generally grouped into non-peptide mimetics of natural GPCR receptor ligands, peptide and non-peptide allosteric effectors of GPCR receptors, and peptides that may function as activators or inhibitors (competitive, uncompetitive and non-competitive) (e.g., antibody products) of GPCR receptors. The invention does not restrict the sources for suitable modulators, which may 20 be obtained from natural sources such as plant, animal or mineral extracts, or non-natural sources such as small molecule libraries, including the products of combinatorial chemical approaches to library construction, and peptide libraries. Examples of peptide modulators of GPCR receptors exhibit the following primary structures: GLGPRPLRFamide, GNSFLRFamide, GGPQGPLRFamide, GPSGPLRFamide, PDVDHVFLRFamide, and pyro- 25 EDVDHVFLRFamide.

Other assays can be used to examine enzymatic activity including, but not limited to, photometric, radiometric, HPLC, electrochemical, and the like, which are described in, for example, *Enzyme Assays: A Practical Approach*, eds. R. Eisenthal and M. J. Danson, 1992, Oxford University Press, which is incorporated herein by reference in its entirety.

30 The use of cDNAs encoding GPCRs in drug discovery programs is well-known; assays capable of testing thousands of unknown compounds per day in high-throughput screens (HTSs) are thoroughly documented. The literature is replete with examples of the use of radiolabelled ligands in HTS binding assays for drug discovery (see Williams, *Medicinal Research Reviews*, 1991, 11, 147-184.; Sweetnam, *et al.*, *J. Natural Products*, 1993, 56, 441-455 for review).

Recombinant receptors are preferred for binding assay HTS because they allow for better specificity (higher relative purity), provide the ability to generate large amounts of receptor material, and can be used in a broad variety of formats (see Hodgson, *Bio/Technology*, 1992, 10, 973-980; each of which is incorporated herein by reference in its entirety).

5 A variety of heterologous systems is available for functional expression of recombinant receptors that are well known to those skilled in the art. Such systems include bacteria (Strosberg, *et al.*, *Trends in Pharmacological Sciences*, 1992, 13, 95-98), yeast (Pausch, *Trends in Biotechnology*, 1997, 15, 487-494), several kinds of insect cells (Vanden Broeck, *Int. Rev. Cytology*, 1996, 164, 189-268), amphibian cells (Jayawickreme *et al.*, *Current Opinion in Biotechnology*, 1997, 8, 629-634) and several mammalian cell lines (CHO, HEK293, COS, etc.; see Gerhardt, *et al.*, *Eur. J. Pharmacology*, 1997, 334, 1-23). These examples do not preclude the use of other possible cell expression systems, including cell lines obtained from nematodes (PCT application WO 98/37177).

10 In preferred embodiments of the invention, methods of screening for compounds that modulate nGPCR-x activity comprise contacting test compounds with nGPCR-x and assaying for the presence of a complex between the compound and nGPCR-x. In such assays, the ligand is typically labeled. After suitable incubation, free ligand is separated from that present in bound form, and the amount of free or uncomplexed label is a measure of the ability of the particular compound to bind to nGPCR-x.

15 It is well known that activation of heterologous receptors expressed in recombinant systems results in a variety of biological responses, which are mediated by G proteins expressed in the host cells. Occupation of a GPCR by an agonist results in exchange of bound GDP for GTP at a binding site on the G_α subunit; one can use a radioactive, non-hydrolyzable derivative of GTP, GTPγ[³⁵S], to measure binding of an agonist to the receptor (Sim *et al.*, *Neuroreport*, 1996, 7, 729-733). One can also use this binding to measure the ability of antagonists to bind to the receptor by decreasing binding of GTPγ[³⁵S] in the presence of a known agonist. One could therefore construct a HTS based on GTPγ[³⁵S] binding, though this is not the preferred method.

20 The G proteins required for functional expression of heterologous GPCRs can be native constituents of the host cell or can be introduced through well-known recombinant technology. 25 The G proteins can be intact or chimeric. Often, a nearly universally competent G protein (e.g., G_{α16}) is used to couple any given receptor to a detectable response pathway. G protein activation results in the stimulation or inhibition of other native proteins, events that can be linked to a measurable response.

Examples of such biological responses include, but are not limited to, the following: the ability to survive in the absence of a limiting nutrient in specifically engineered yeast cells (Pausch, *Trends in Biotechnology*, 1997, 15, 487-494); changes in intracellular Ca^{2+} concentration as measured by fluorescent dyes (Murphy, *et al.*, *Cur. Opinion Drug Disc. Dev.*, 1998, 1, 192-199). Fluorescence changes can also be used to monitor ligand-induced changes in membrane potential or intracellular pH; an automated system suitable for HTS has been described for these purposes (Schroeder, *et al.*, *J. Biomolecular Screening*, 1996, 1, 75-80). Melanophores prepared from *Xenopus laevis* show a ligand-dependent change in pigment organization in response to heterologous GPCR activation; this response is adaptable to HTS formats (Jayawickreme *et al.*, *Cur. Opinion Biotechnology*, 1997, 8, 629-634). Assays are also available for the measurement of common second messengers, including cAMP, phosphoinositides and arachidonic acid, but these are not generally preferred for HTS.

Preferred methods of HTS employing these receptors include permanently transfected CHO cells, in which agonists and antagonists can be identified by the ability to specifically alter the binding of $\text{GTP}\gamma^{35}\text{S}$ in membranes prepared from these cells. In another embodiment of the invention, permanently transfected CHO cells could be used for the preparation of membranes which contain significant amounts of the recombinant receptor proteins; these membrane preparations would then be used in receptor binding assays, employing the radiolabelled ligand specific for the particular receptor. Alternatively, a functional assay, such as fluorescent monitoring of ligand-induced changes in internal Ca^{2+} concentration or membrane potential in permanently transfected CHO cells containing each of these receptors individually or in combination would be preferred for HTS. Equally preferred would be an alternative type of mammalian cell, such as HEK293 or COS cells, in similar formats. More preferred would be permanently transfected insect cell lines, such as *Drosophila* S2 cells. Even more preferred would be recombinant yeast cells expressing the *Drosophila melanogaster* receptors in HTS formats well known to those skilled in the art (e.g., Pausch, *Trends in Biotechnology*, 1997, 15, 487-494).

The invention contemplates a multitude of assays to screen and identify inhibitors of ligand binding to nGPCR-x receptors. In one example, the nGPCR-x receptor is immobilized and interaction with a binding partner is assessed in the presence and absence of a candidate modulator such as an inhibitor compound. In another example, interaction between the nGPCR-x receptor and its binding partner is assessed in a solution assay, both in the presence and absence of a candidate inhibitor compound. In either assay, an inhibitor is identified as a compound that decreases binding between the nGPCR-x receptor and its binding partner.

Following identification of compounds that inhibit ligand binding to nGPCR-x receptors, such compounds can be further tested in other assays including, but not limited to, *in vivo* models, in order to confirm or quantitate their activity. Another contemplated assay involves a variation of the dihybrid assay wherein an inhibitor of protein/protein interactions is identified by detection 5 of a positive signal in a transformed or transfected host cell, as described in PCT publication number WO 95/20652, published August 3, 1995.

Candidate modulators contemplated by the invention include compounds selected from libraries of either potential activators or potential inhibitors. There are a number of different libraries used for the identification of small molecule modulators, including: (1) chemical 10 libraries, (2) natural product libraries, and (3) combinatorial libraries comprised of random peptides, oligonucleotides or organic molecules. Chemical libraries consist of random chemical structures, some of which are analogs of known compounds or analogs of compounds that have been identified as "hits" or "leads" in other drug discovery screens, some of which are derived from natural products, and some of which arise from non-directed synthetic organic chemistry. 15 Natural product libraries are collections of microorganisms, animals, plants, or marine organisms which are used to create mixtures for screening by: (1) fermentation and extraction of broths from soil, plant or marine microorganisms or (2) extraction of plants or marine organisms. Natural product libraries include polyketides, non-ribosomal peptides, and variants (non-naturally occurring) thereof. For a review, see *Science* 282:63-68 (1998). Combinatorial 20 libraries are composed of large numbers of peptides, oligonucleotides, or organic compounds as a mixture. These libraries are relatively easy to prepare by traditional automated synthesis methods, PCR, cloning, or proprietary synthetic methods. Of particular interest are non-peptide combinatorial libraries. Still other libraries of interest include peptide, protein, peptidomimetic, multiparallel synthetic collection, recombinatorial, and polypeptide libraries. For a review of 25 combinatorial chemistry and libraries created therefrom, see Myers, *Curr. Opin. Biotechnol.* 8:701-707 (1997). Identification of modulators through use of the various libraries described herein permits modification of the candidate "hit" (or "lead") to optimize the capacity of the "hit" to modulate activity.

Still other candidate inhibitors contemplated by the invention can be designed and 30 include soluble forms of binding partners, as well as such binding partners as chimeric, or fusion, proteins. A "binding partner" as used herein broadly encompasses non-peptide modulators, as well as such peptide modulators as neuropeptides other than natural ligands, antibodies, antibody fragments, and modified compounds comprising antibody domains that are immunospecific for the expression product of the identified nGPCR-x gene.

The polypeptides of the invention are employed as a research tool for identification, characterization and purification of interacting, regulatory proteins. Appropriate labels are incorporated into the polypeptides of the invention by various methods known in the art and the polypeptides are used to capture interacting molecules. For example, molecules are incubated 5 with the labeled polypeptides, washed to remove unbound polypeptides, and the polypeptide complex is quantified. Data obtained using different concentrations of polypeptide are used to calculate values for the number, affinity, and association of polypeptide with the protein complex.

Labeled polypeptides are also useful as reagents for the purification of molecules with 10 which the polypeptide interacts including, but not limited to, inhibitors. In one embodiment of affinity purification, a polypeptide is covalently coupled to a chromatography column. Cells and their membranes are extracted, and various cellular subcomponents are passed over the column. Molecules bind to the column by virtue of their affinity to the polypeptide. The polypeptide- 15 complex is recovered from the column, dissociated and the recovered molecule is subjected to protein sequencing. This amino acid sequence is then used to identify the captured molecule or to design degenerate oligonucleotides for cloning the corresponding gene from an appropriate cDNA library.

Alternatively, compounds may be identified which exhibit similar properties to the ligand for the nGPCR-x of the invention, but which are smaller and exhibit a longer half time 20 than the endogenous ligand in a human or animal body. When an organic compound is designed, a molecule according to the invention is used as a "lead" compound. The design of mimetics to known pharmaceutically active compounds is a well-known approach in the development of pharmaceuticals based on such "lead" compounds. Mimetic design, synthesis and testing are generally used to avoid randomly screening a large number of molecules for a target property. 25 Furthermore, structural data deriving from the analysis of the deduced amino acid sequences encoded by the DNAs of the present invention are useful to design new drugs, more specific and therefore with a higher pharmacological potency.

Comparison of the protein sequence of the present invention with the sequences present in all the available databases showed a significant homology with the transmembrane portion of 30 G protein coupled receptors. Accordingly, computer modeling can be used to develop a putative tertiary structure of the proteins of the invention based on the available information of the transmembrane domain of other proteins. Thus, novel ligands based on the predicted structure of nGPCR-x can be designed.

In a particular embodiment, the novel molecules identified by the screening methods according to the invention are low molecular weight organic molecules, in which case a composition or pharmaceutical composition can be prepared thereof for oral intake, such as in tablets. The compositions, or pharmaceutical compositions, comprising the nucleic acid molecules, vectors, polypeptides, antibodies and compounds identified by the screening methods described herein, can be prepared for any route of administration including, but not limited to, oral, intravenous, cutaneous, subcutaneous, nasal, intramuscular or intraperitoneal. The nature of the carrier or other ingredients will depend on the specific route of administration and particular embodiment of the invention to be administered. Examples of techniques and protocols that are useful in this context are, *inter alia*, found in Remington's Pharmaceutical Sciences, 16th edition, Osol, A (ed.), 1980, which is incorporated herein by reference in its entirety.

The dosage of these low molecular weight compounds will depend on the disease state or condition to be treated and other clinical factors such as weight and condition of the human or animal and the route of administration of the compound. For treating human or animals, between approximately 0.5 mg/kg of body weight to 500 mg/kg of body weight of the compound can be administered. Therapy is typically administered at lower dosages and is continued until the desired therapeutic outcome is observed.

The present compounds and methods, including nucleic acid molecules, polypeptides, antibodies, compounds identified by the screening methods described herein, have a variety of pharmaceutical applications and may be used, for example, to treat or prevent unregulated cellular growth, such as cancer cell and tumor growth. In a particular embodiment, the present molecules are used in gene therapy. For a review of gene therapy procedures, see e.g. Anderson, *Science*, 1992, 256, 808-813, which is incorporated herein by reference in its entirety.

The present invention also encompasses a method of agonizing (stimulating) or antagonizing a nGPCR-x natural binding partner associated activity in a mammal comprising administering to said mammal an agonist or antagonist to one of the above disclosed polypeptides in an amount sufficient to effect said agonism or antagonism. One embodiment of the present invention, then, is a method of treating diseases in a mammal with an agonist or antagonist of the protein of the present invention comprises administering the agonist or antagonist to a mammal in an amount sufficient to agonize or antagonize nGPCR-x-associated functions.

In an effort to discover novel treatments for diseases, biomedical researchers and chemists have designed, synthesized, and tested molecules that modulate the function of G

protein coupled receptors. Some small organic molecules form a class of compounds that modulate the function of G protein coupled receptors.

Exemplary diseases and conditions amenable to treatment based on the present invention include, but are not limited to, thyroid disorders (e.g. thyrotoxicosis, myxoedema); renal failure; inflammatory conditions (e.g., Crohn's disease); diseases related to cell differentiation and homeostasis; rheumatoid arthritis; autoimmune disorders; movement disorders; CNS disorders (e.g., pain including migraine; stroke; psychotic and neurological disorders, including anxiety, mental disorder, manic depression, anxiety, generalized anxiety disorder, post-traumatic-stress disorder, Schizophrenia, depression, bipolar disorder, delirium, dementia, 10 severe mental retardation; dyskinesias, such as Huntington's disease or Tourette's Syndrome; attention disorders including ADD and ADHD, and degenerative disorders such as Parkinson's, Alzheimer's; movement disorders, including ataxias, supranuclear palsy, etc.); infections, such as viral infections caused by HIV-1 or HIV-2; metabolic and cardiovascular diseases and disorders (e.g., type 2 diabetes, impaired glucose tolerance, dyslipidemia, obesity, anorexia, 15 hypotension, hypertension, thrombosis, myocardial infarction, cardiomyopathies, atherosclerosis, etc.); proliferative diseases and cancers (e.g., different cancers such as breast, colon, lung, etc., and hyperproliferative disorders such as psoriasis, prostate hyperplasia, etc.); hormonal disorders (e.g., male/female hormonal replacement, polycystic ovarian syndrome, alopecia, etc.); sexual dysfunction, among others.

20 Methods of determining the dosages of compounds to be administered to a patient and modes of administering compounds to an organism are disclosed in U.S. Application Serial No. 08/702,282, filed August 23, 1996 and International patent publication number WO 96/22976, published August 1 1996, both of which are incorporated herein by reference in their entirety, including any drawings, figures or tables. Those skilled in the art will appreciate that such 25 descriptions are applicable to the present invention and can be easily adapted to it.

The proper dosage depends on various factors such as the type of disease being treated, the particular composition being used and the size and physiological condition of the patient, including such factors as, for example, weight, age, sex, disease state, etc. Therapeutically effective doses for the compounds described herein can be estimated initially from cell culture 30 and animal models. For example, a dose can be formulated in animal models to achieve a circulating concentration range that initially takes into account the IC₅₀ as determined in cell culture assays. The animal model data can be used to more accurately determine useful doses in humans.

Plasma half-life and biodistribution of the drug and metabolites in the plasma, tumors and major organs can also be determined to facilitate the selection of drugs most appropriate to inhibit a disorder. Such measurements can be carried out. For example, HPLC analysis can be performed on the plasma of animals treated with the drug and the location of radiolabeled 5 compounds can be determined using detection methods such as X-ray, CAT scan and MRI. Compounds that show potent inhibitory activity in the screening assays, but have poor pharmacokinetic characteristics, can be optimized by altering the chemical structure and retesting. In this regard, compounds displaying good pharmacokinetic characteristics can be used as a model.

Toxicity studies can also be carried out by measuring the blood cell composition. For 10 example, toxicity studies can be carried out in a suitable animal model as follows: 1) the compound is administered to mice (an untreated control mouse should also be used); 2) blood samples are periodically obtained via the tail vein from one mouse in each treatment group; and 3) the samples are analyzed for red and white blood cell counts, blood cell composition and the percent of lymphocytes versus polymorphonuclear cells. A comparison of results for each 15 dosing regime with the controls indicates if toxicity is present.

At the termination of each toxicity study, further studies can be carried out by sacrificing the animals (preferably, in accordance with the American Veterinary Medical Association guidelines Report of the American Veterinary Medical Assoc. Panel on Euthanasia, Journal of American Veterinary Medical Assoc., 202:229-249, 1993). Representative animals from each 20 treatment group can then be examined by gross necropsy for immediate evidence of metastasis, unusual illness or toxicity. Gross abnormalities in tissue are noted and tissues are examined histologically. Compounds causing a reduction in body weight or blood components are less preferred, as are compounds having an adverse effect on major organs. In general, the greater the adverse effect the less preferred the compound.

25 For the treatment of many diseases, the expected daily dose of a hydrophobic pharmaceutical agent is between 1 to 500 mg/day, preferably 1 to 250 mg/day, and most preferably 1 to 50 mg/day. Drugs can be delivered less frequently provided plasma levels of the active moiety are sufficient to maintain therapeutic effectiveness. Plasma levels should reflect the potency of the drug. Generally, the more potent the compound the lower the plasma levels 30 necessary to achieve efficacy.

nGPCR-42, 46, 48, 49, 51, 52, 61, 63, and 70 mRNA transcripts may be found in many tissues, including, but not limited to, brain, peripheral blood lymphocytes, pancreas, ovary, uterus, testis, salivary gland, kidney, adrenal gland, liver, bone marrow, prostate, fetal liver, colon, muscle, and fetal brain, and may be found in many other tissues. Within the brain,

nGPCR-42, 46, 48, 49, 51, 52, 61, 63, and 70 mRNA transcripts may be found in many tissues, including, but not limited to, frontal lobe, hypothalamus, pons, cerebellum, caudate nucleus, and medulla. Tissues and brain regions where specific nGPCR mRNA transcripts are expressed are identified in the Examples, below.

5 Sequences selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60 will, as detailed above, enable screening the endogenous neurotransmitters/hormones/ligands which activate, agonize, or antagonize nGPCR-x and for compounds with potential utility in treating disorders including, but not limited to, thyroid disorders (e.g. thyreotoxicosis, myxoedema); renal failure; inflammatory conditions (e.g., Crohn's disease); diseases related to cell

10 differentiation and homeostasis; rheumatoid arthritis; autoimmune disorders; movement disorders; CNS disorders (e.g., pain including migraine; stroke; psychotic and neurological disorders, including anxiety, mental disorder, manic depression, anxiety, generalized anxiety disorder, post-traumatic-stress disorder, Schizophrenia, depression, bipolar disorder, delirium, dementia, severe mental retardation; dyskinesias, such as Huntington's disease or Tourette's

15 Syndrome; attention disorders including ADD and ADHD, and degenerative disorders such as Parkinson's, Alzheimer's; movement disorders, including ataxias, supranuclear palsy, etc.); infections, such as viral infections caused by HIV-1 or HIV-2; metabolic and cardiovascular diseases and disorders (e.g., type 2 diabetes, impaired glucose tolerance, dyslipidemia, obesity, anorexia, hypotension, hypertension, thrombosis, myocardial infarction, cardiomyopathies, atherosclerosis, etc.); proliferative diseases and cancers (e.g., different cancers such as breast, colon, lung, etc., and hyperproliferative disorders such as psoriasis, prostate hyperplasia, etc.); hormonal disorders (e.g., male/female hormonal replacement, polycystic ovarian syndrome, alopecia, etc.); sexual dysfunction, among others.

20 For example, nGPCR-x may be useful in the treatment of respiratory ailments such as asthma, where T cells are implicated by the disease. Contraction of airway smooth muscle is stimulated by thrombin. Cicala *et al* (1999) Br J Pharmacol 126:478-484. Additionally, in bronchiolitis obliterans, it has been noted that activation of thrombin receptors may be deleterious. Hauck *et al.* (1999) Am J Physiol 277:L22-L29. Furthermore, mast cells have also been shown to have thrombin receptors. Cirino *et al* (1996) J Exp Med 183:821-827. nGPCR-x

25 may also be useful in remodeling of airway structures in chronic pulmonary inflammation via stimulation of fibroblast procollagen synthesis. See, e.g., Chambers *et al.* (1998) Biochem J 333:121-127; Trejo *et al.* (1996) J Biol Chem 271:21536-21541.

30 In another example, increased release of sCD40L and expression of CD40L by T cells after activation of thrombin receptors suggests that nGPCR-x may be useful in the treatment of

unstable angina due to the role of T cells and inflammation. See Aukrust *et al.* (1999) Circulation 100:614-620.

A further example is the treatment of inflammatory diseases, such as psoriasis, inflammatory bowel disease, multiple sclerosis, rheumatoid arthritis, and thyroiditis. Due to the 5 tissue expression profile of nGPCR-x, inhibition of thrombin receptors may be beneficial for these diseases. See, e.g., Morris *et al.* (1996) Ann Rheum Dis 55:841-843. In addition to T cells, NK cells and monocytes are also critical cell types which contribute to the pathogenesis of these diseases. See, e.g., Naldini & Carney (1996) Cell Immunol 172:35-42; Hoffman & Cooper (1995) Blood Cells Mol Dis 21:156-167; Colotta *et al.* (1994) Am J Pathol 144:975-985.

10 Expression of nGPCR-x in bone marrow and spleen may suggest that it may play a role in the proliferation of hematopoietic progenitor cells. See DiCuccio *et al.* (1996) Exp Hematol 24:914-918.

15 As another example, nGPCR-x may be useful in the treatment of acute and/or traumatic brain injury. Astrocytes have been demonstrated to express thrombin receptors. Activation of thrombin receptors may be involved in astrogliosis following brain injury. Therefore, inhibition of receptor activity may be beneficial for limiting neuroinflammation. Scar formation mediated by astrocytes may also be limited by inhibiting thrombin receptors. See, e.g., Pindon *et al.* (1998) Eur J Biochem 255:766-774; Ubl & Reiser. (1997) Glia 21:361-369; Grabham & Cunningham (1995) J Neurochem 64:583-591.

20 nGPCR-x receptor activation may mediate neuronal and astrocyte apoptosis and prevention of neurite outgrowth. Inhibition would be beneficial in both chronic and acute brain injury. See, e.g., Donovan *et al.* (1997) J Neurosci 17:5316-5326; Turgeon *et al.* (1998) J Neurosci 18:6882-6891; Smith-Swintosky *et al.* (1997) J Neurochem 69:1890-1896; Gill *et al.* (1998) Brain Res 797:321-327; Suidan *et al.* (1996) Semin Thromb Hemost 22:125-133.

25 The attached Sequence Listing contains the sequences of the polynucleotides and polypeptides of the invention and is incorporated herein by reference in its entirety.

As described above and in Example 5 below, the genes encoding nGPCR-42, 46, 48, 49, 51, 52, 61, 63, and 70 have been detected in brain tissue indicating that these n-GPCR-x proteins are neuroreceptors. The identification of modulators such as agonists and antagonists is 30 therefore useful for the identification of compounds useful to treat neurological diseases and psychiatric disorders. Such neurological diseases and disorders, including but are not limited to, mental disorder, affective disorders, ADHD/ADD, and neural disorders such as Alzheimer's disease, Parkinson's disease, migraine, schizophrenia, and senile dementia as well as depression, anxiety, bipolar disease, epilepsy, neuritis, neurasthenia, neuropathy, neuroses, and the like.

Methods of Screening Human Subjects

Thus in yet another embodiment, the invention provides genetic screening procedures that entail analyzing a person's genome -- in particular their alleles for GPCRs of the invention -- to determine whether the individual possesses a genetic characteristic found in other 5 individuals that are considered to be afflicted with, or at risk for, developing a mental disorder or disease of the brain that is suspected of having a hereditary component. For example, in one embodiment, the invention provides a method for determining a potential for developing a disorder affecting the brain in a human subject comprising the steps of analyzing the coding sequence of one or more GPCR genes from the human subject; and determining development 10 potential for the disorder in said human subject from the analyzing step.

More particularly, the invention provides a method of screening a human subject to diagnose a disorder affecting the brain or genetic predisposition therefor, comprising the steps of: (a) assaying nucleic acid of a human subject to determine a presence or an absence of a mutation altering the amino acid sequence, expression, or biological activity of at least one 15 seven transmembrane receptor that is expressed in the brain, wherein the seven transmembrane receptor comprises an amino acid sequence selected from the group consisting of SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120, or an allelic variant thereof, and wherein the nucleic acid corresponds to the gene encoding the seven transmembrane receptor; and (b) diagnosing the disorder or predisposition from the presence or absence of said mutation, 20 wherein the presence of a mutation altering the amino acid sequence, expression, or biological activity of allele in the nucleic acid correlates with an increased risk of developing the disorder. In preferred variations, the seven transmembrane receptor is nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 comprising an amino acid sequence set forth in SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120, or an allelic variant thereof, and the disease is mental disorder.

25 By "human subject" is meant any human being, human embryo, or human fetus. It will be apparent that methods of the present invention will be of particular interest to individuals that have themselves been diagnosed with a disorder affecting the brain or have relatives that have been diagnosed with a disorder affecting the brain.

By "screening for an increased risk" is meant determination of whether a genetic 30 variation exists in the human subject that correlates with a greater likelihood of developing a disorder affecting the brain than exists for the human population as a whole, or for a relevant racial or ethnic human sub-population to which the individual belongs. Both positive and negative determinations (i.e., determinations that a genetic predisposition marker is present or is absent) are intended to fall within the scope of screening methods of the invention. In preferred

embodiments, the presence of a mutation altering the sequence or expression of at least one nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 seven transmembrane receptor allele in the nucleic acid is correlated with an increased risk of developing mental disorder, whereas the absence of such a mutation is reported as a negative determination.

5 The "assaying" step of the invention may involve any techniques available for analyzing nucleic acid to determine its characteristics, including but not limited to well-known techniques such as single-strand conformation polymorphism analysis (SSCP) (Orita *et al.*, *Proc Natl. Acad. Sci. USA*, 86: 2766-2770 (1989)); heteroduplex analysis (White *et al.*, *Genomics*, 12: 301-306 (1992)); denaturing gradient gel electrophoresis analysis (Fischer *et al.*, *Proc. Natl. Acad. Sci. USA*, 80: 1579-1583 (1983); and Riesner *et al.*, *Electrophoresis*, 10: 377-389 (1989)); DNA sequencing; RNase cleavage (Myers *et al.*, *Science*, 230: 1242-1246 (1985)); chemical cleavage of mismatch techniques (Rowley *et al.*, *Genomics*, 30: 574-582 (1995); and Roberts *et al.*, *Nucl. Acids Res.*, 25: 3377-3378 (1997)); restriction fragment length polymorphism analysis; single nucleotide primer extension analysis (Shumaker *et al.*, *Hum. Mutat.*, 7: 346-354 (1996); and 10 Pastinen *et al.*, *Genome Res.*, 7: 606-614 (1997)); 5' nuclease assays (Pease *et al.*, *Proc. Natl. Acad. Sci. USA*, 91:5022-5026 (1994)); DNA Microchip analysis (Ramsay, G., *Nature Biotechnology*, 16: 40-48 (1999); and Chee *et al.*, U.S. Patent No. 5,837,832); and ligase chain reaction (Whiteley *et al.*, U.S. Patent No. 5,521,065). (See generally, Schafer and Hawkins, *Nature Biotechnology*, 16: 33-39 (1998).) All of the foregoing documents are hereby 15 incorporated herein by reference in their entirety.

Thus, in one preferred embodiment involving screening nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 sequences, for example, the assaying step comprises at least one procedure selected from the group consisting of: (a) determining a nucleotide sequence of at least one codon of at least one nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 allele of the human subject; (b) 25 performing a hybridization assay to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences; (c) performing a polynucleotide migration assay to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences; and (d) performing a restriction endonuclease digestion to determine whether nucleic 30 acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences.

In a highly preferred embodiment, the assaying involves sequencing of nucleic acid to determine nucleotide sequence thereof, using any available sequencing technique. (See, e.g., Sanger *et al.*, *Proc. Natl. Acad. Sci. (USA)*, 74: 5463-5467 (1977) (dideoxy chain termination

method); Mirzabekov, *TIBTECH*, 12: 27-32 (1994) (sequencing by hybridization); Drmanac *et al.*, *Nature Biotechnology*, 16: 54-58 (1998); U.S. Patent No. 5,202,231; and *Science*, 260: 1649-1652 (1993) (sequencing by hybridization); Kieleczawa *et al.*, *Science*, 258: 1787-1791 (1992) (sequencing by primer walking); (Douglas *et al.*, *Biotechniques*, 14: 824-828 (1993) 5 (Direct sequencing of PCR products); and Akane *et al.*, *Biotechniques* 16: 238-241 (1994); Maxam and Gilbert, *Meth. Enzymol.*, 65: 499-560 (1977) (chemical termination sequencing), all incorporated herein by reference in its entirety). The analysis may entail sequencing of the entire nGPCR gene genomic DNA sequence, or portions thereof; or sequencing of the entire seven transmembrane receptor coding sequence or portions thereof. In some circumstances, the 10 analysis may involve a determination of whether an individual possesses a particular allelic variant, in which case sequencing of only a small portion of nucleic acid -- enough to determine the sequence of a particular codon characterizing the allelic variant -- is sufficient. This approach is appropriate, for example, when assaying to determine whether one family member inherited the same allelic variant that has been previously characterized for another family 15 member, or, more generally, whether a person's genome contains an allelic variant that has been previously characterized and correlated with a mental disorder having a heritable component.

In another highly preferred embodiment, the assaying step comprises performing a hybridization assay to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences. In a preferred 20 embodiment, the hybridization involves a determination of whether nucleic acid derived from the human subject will hybridize with one or more oligonucleotides, wherein the oligonucleotides have nucleotide sequences that correspond identically to a portion of the GPCR gene sequence taught herein, such as the nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 coding sequence set forth in SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120, or that 25 correspond identically except for one mismatch. The hybridization conditions are selected to differentiate between perfect sequence complementarity and imperfect matches differing by one or more bases. Such hybridization experiments thereby can provide single nucleotide polymorphism sequence information about the nucleic acid from the human subject, by virtue of knowing the sequences of the oligonucleotides used in the experiments.

30 Several of the techniques outlined above involve an analysis wherein one performs a polynucleotide migration assay, *e.g.*, on a polyacrylamide electrophoresis gel (or in a capillary electrophoresis system), under denaturing or non-denaturing conditions. Nucleic acid derived from the human subject is subjected to gel electrophoresis, usually adjacent to (or co-loaded with) one or more reference nucleic acids, such as reference GPCR-encoding sequences having

a coding sequence identical to all or a portion of SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120 (or identical except for one known polymorphism). The nucleic acid from the human subject and the reference sequence(s) are subjected to similar chemical or enzymatic treatments and then electrophoresed under conditions whereby the polynucleotides will show a differential migration pattern, unless they contain identical sequences. (See generally Ausubel *et al.* (eds.), *Current Protocols in Molecular Biology*, New York: John Wiley & Sons, Inc. (1987-1999); and Sambrook *et al.*, (eds.), *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor, New York: Cold Spring Harbor Laboratory Press (1989), both incorporated herein by reference in their entirety).

10 In the context of assaying, the term "nucleic acid of a human subject" is intended to include nucleic acid obtained directly from the human subject (e.g., DNA or RNA obtained from a biological sample such as a blood, tissue, or other cell or fluid sample); and also nucleic acid derived from nucleic acid obtained directly from the human subject. By way of non-limiting examples, well known procedures exist for creating cDNA that is complementary to RNA derived from a biological sample from a human subject, and for amplifying (e.g., via polymerase chain reaction (PCR)) DNA or RNA derived from a biological sample obtained from a human subject. Any such derived polynucleotide which retains relevant nucleotide sequence information of the human subject's own DNA/RNA is intended to fall within the definition of "nucleic acid of a human subject" for the purposes of the present invention.

15 20 In the context of assaying, the term "mutation" includes addition, deletion, and/or substitution of one or more nucleotides in the GPCR gene sequence (e.g., as compared to the seven transmembrane receptor-encoding sequences set forth of SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120 and other polymorphisms that occur in introns (where introns exist) and that are identifiable via sequencing, restriction fragment length polymorphism, or other techniques. The various activity examples provided herein permit determination of whether a mutation modulates activity of the relevant receptor in the presence or absence of various test substances.

25 30 In a related embodiment, the invention provides methods of screening a person's genotype with respect to GPCRs of the invention, and correlating such genotypes with diagnoses for disease or with predisposition for disease (for genetic counseling). For example, the invention provides a method of screening for an nGPCR-63 hereditary mental disorder genotype in a human patient, comprising the steps of: (a) providing a biological sample comprising nucleic acid from the patient, the nucleic acid including sequences corresponding to said patient's nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 alleles; (b) analyzing the nucleic acid

for the presence of a mutation or mutations; (c) determining an nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 genotype from the analyzing step; and (d) correlating the presence of a mutation in an nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 allele with a hereditary mental disorder genotype. In a preferred embodiment, the biological sample is a cell sample containing human 5 cells that contain genomic DNA of the human subject. The analyzing can be performed analogously to the assaying described in preceding paragraphs. For example, the analyzing comprises sequencing a portion of the nucleic acid (e.g., DNA or RNA), the portion comprising at least one codon of the nGPCR-63 alleles.

10 Although more time consuming and expensive than methods involving nucleic acid analysis, the invention also may be practiced by assaying protein of a human subject to determine the presence or absence of an amino acid sequence variation in GPCR protein from the human subject. Such protein analyses may be performed, e.g., by fragmenting GPCR protein via chemical or enzymatic methods and sequencing the resultant peptides; or by Western analyses using an antibody having specificity for a particular allelic variant of the GPCR.

15 The invention also provides materials that are useful for performing methods of the invention. For example, the present invention provides oligonucleotides useful as probes in the many analyzing techniques described above. In general, such oligonucleotide probes comprise 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50 nucleotides that have a 20 sequence that is identical, or exactly complementary, to a portion of a human GPCR gene sequence taught herein (or allelic variant thereof), or that is identical or exactly complementary except for one nucleotide substitution. In a preferred embodiment, the oligonucleotides have a sequence that corresponds in the foregoing manner to a human GPCR coding sequence taught herein, and in particular, the coding sequences set forth in SEQ ID Numbers 61, 62, 68, 91, 94, 25 96, 97, 99, 100, and 111-120. In one variation, an oligonucleotide probe of the invention is purified and isolated. In another variation, the oligonucleotide probe is labeled, e.g., with a radioisotope, chromophore, or fluorophore. In yet another variation, the probe is covalently attached to a solid support. (See generally Ausubel *et al.* and Sambrook *et al.*, *supra*.)

30 In a related embodiment, the invention provides kits comprising reagents that are useful for practicing methods of the invention. For example, the invention provides a kit for screening a human subject to diagnose a mental disorder or a genetic predisposition therefor, comprising, in association: (a) an oligonucleotide useful as a probe for identifying polymorphisms in a human nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 seven transmembrane receptor gene, the oligonucleotide comprising 6-50 nucleotides that have a sequence that is identical or exactly

complementary to a portion of a human nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 gene sequence or nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 coding sequence, except for one sequence difference selected from the group consisting of a nucleotide addition, a nucleotide deletion, or nucleotide substitution; and (b) a media packaged with the oligonucleotide

5 containing information identifying polymorphisms identifiable with the probe that correlate with mental disorder or a genetic predisposition therefor. Exemplary information-containing media include printed paper package inserts or packaging labels; and magnetic and optical storage media that are readable by computers or machines used by practitioners who perform genetic screening and counseling services. The practitioner uses the information provided in the media

10 to correlate the results of the analysis with the oligonucleotide with a diagnosis. In a preferred variation, the oligonucleotide is labeled.

In still another embodiment, the invention provides methods of identifying those allelic variants of GPCRs of the invention that correlate with mental disorders. For example, the invention provides a method of identifying a seven transmembrane allelic variant that correlates with a mental disorder, comprising steps of: (a) providing a biological sample comprising nucleic acid from a human patient diagnosed with a mental disorder, or from the patient's genetic progenitors or progeny; (b) analyzing the nucleic acid for the presence of a mutation or mutations in at least one seven transmembrane receptor that is expressed in the brain, wherein the at least one seven transmembrane receptor comprises an amino acid sequence selected from the group consisting of SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120 or an allelic variant thereof, and wherein the nucleic acid includes sequence corresponding to the gene or genes encoding the at least one seven transmembrane receptor; (c) determining a genotype for the patient for the at least one seven transmembrane receptor from said analyzing step; and (d) identifying an allelic variant that correlates with the mental disorder from the determining step.

25 To expedite this process, it may be desirable to perform linkage studies in the patients (and possibly their families) to correlate chromosomal markers with disease states. The chromosomal localization data provided herein facilitates identifying an involved GPCR with a chromosomal marker.

The foregoing method can be performed to correlate GPCRs of the invention to a number of disorders having hereditary components that are causative or that predispose persons to the disorder. For example, in one preferred variation, the disorder is a mental disorder, and the at least one seven transmembrane receptor comprises nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 having an amino acid sequence set forth in SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120 or an allelic variant thereof.

Also contemplated as part of the invention are polynucleotides that comprise the allelic variant sequences identified by such methods, and polypeptides encoded by the allelic variant sequences, and oligonucleotide and oligopeptide fragments thereof that embody the mutations that have been identified. Such materials are useful in *in vitro* cell-free and cell-based assays for identifying lead compounds and therapeutics for treatment of the disorders. For example, the variants are used in activity assays, binding assays, and assays to screen for activity modulators described herein. In one preferred embodiment, the invention provides a purified and isolated polynucleotide comprising a nucleotide sequence encoding a nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 receptor allelic variant identified according to the methods described above; and an oligonucleotide that comprises the sequences that differentiate the allelic variant from the nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 sequences set forth in SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120. The invention also provides a vector comprising the polynucleotide (preferably an expression vector); and a host cell transformed or transfected with the polynucleotide or vector. The invention also provides an isolated cell line that is expressing the allelic variant GPCR polypeptide; purified cell membranes from such cells; purified polypeptide; and synthetic peptides that embody the allelic variation amino acid sequence. In one particular embodiment, the invention provides a purified polynucleotide comprising a nucleotide sequence encoding a nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 seven transmembrane receptor protein of a human that is affected with a mental disorder; wherein said polynucleotide hybridizes to the complement of SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120 under the following hybridization conditions: (a) hybridization for 16 hours at 42C in a hybridization solution comprising 50% formamide, 1% SDS, 1 M NaCl, 10% dextran sulfate and (b) washing 2 times for 30 minutes at 60C in a wash solution comprising 0.1x SSC and 1% SDS; and wherein the polynucleotide encodes a nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 amino acid sequence that differs from SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120 by at least one residue.

An exemplary assay for using the allelic variants is a method for identifying a modulator of nGPCR-x biological activity, comprising the steps of: (a) contacting a cell expressing the allelic variant in the presence and in the absence of a putative modulator compound; (b) measuring nGPCR-x biological activity in the cell; and (c) identifying a putative modulator compound in view of decreased or increased nGPCR-x biological activity in the presence versus absence of the putative modulator.

Additional features of the invention will be apparent from the following Examples. Examples 1-6, 12, 14, and 15 are actual, while the remaining Examples are prophetic.

Additional features and variations of the invention will be apparent to those skilled in the art from the entirety of this application, including the detailed description, and all such features are intended as aspects of the invention. Likewise, features of the invention described herein can be re-combined into additional embodiments that also are intended as aspects of the invention.

5 irrespective of whether the combination of features is specifically mentioned above as an aspect or embodiment of the invention. Also, only such limitations which are described herein as critical to the invention should be viewed as such; variations of the invention lacking limitations which have not been described herein as critical are intended as aspects of the invention.

EXAMPLES

10 **Example 1: Identification of nGPCR-x**

A. Database search

The Celera database was searched using known GPCR receptors as query sequences to find patterns suggestive of novel G protein-coupled receptors. Positive hits were further analyzed with the GCG program BLAST to determine which ones were the most likely 15 candidates to encode G protein-coupled receptors, using the standard (default) alignment produced by BLAST as a guide.

Briefly, the BLAST algorithm, which stands for Basic Local Alignment Search Tool is suitable for determining sequence similarity (Altschul *et al.*, *J. Mol. Biol.*, 1990, 215, 403-410, which is incorporated herein by reference in its entirety). Software for performing BLAST 20 analyses is publicly available through the National Center for Biotechnology Information through the world wide web of the Internet (ncbi.nlm.nih.gov/). This algorithm involves first identifying high scoring sequence pair (HSPs) by identifying short words of length W in the query sequence that either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood 25 word score threshold (Altschul *et al.*, *supra*). These initial neighbourhood word hits act as seeds for initiating searches to find HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Extension for the word hits in each direction are halted when: 1) the cumulative alignment score falls off by the quantity X from its maximum achieved value; 2) the cumulative score goes to 30 zero or below, due to the accumulation of one or more negative-scoring residue alignments; or 3) the end of either sequence is reached. The Blast algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The Blast program uses as defaults a word length (W) of 11, the BLOSUM62 scoring matrix (see Henikoff *et al.*, *Proc. Natl. Acad. Sci. USA*, 1992,

89, 10915-10919, which is incorporated herein by reference in its entirety) alignments (B) of 50, expectation (E) of 10, M=5, N=4, and a comparison of both strands.

The BLAST algorithm (Karlin *et al.*, Proc. Natl. Acad. Sci. USA, 1993, 90, 5873-5787, which is incorporated herein by reference in its entirety) and Gapped BLAST perform a 5 statistical analysis of the similarity between two sequences. One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a nucleic acid is considered similar to a GPCR gene or cDNA if the smallest sum probability in comparison of the test nucleic acid to a GPCR nucleic acid is less 10 than about 1, preferably less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

Homology searches are performed with the program BLAST version 2.08. A collection of about 200 to about 350 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.01 15 were collected from each BLAST search. The amino acid sequences have been edited to remove regions in the sequence that produce non-significant alignments with proteins that are not related to GPCRs.

Multiple query sequences may have a significant alignment to the same genomic region, although each alignment may not cover exactly the same DNA region. A procedure is used to 20 determine the region of maximum common overlap between the alignments from several query sequences. This region is called the consensus DNA region. The procedure for determining this consensus involves the automatic parsing of the BLAST output files using the program MSPcrunch to produce a tabular report. From this tabular report the start and end of each 25 alignment in the genomic DNA is extracted. This information is used by a PERL script to derive the maximum common overlap. These regions are reported in the form of a unique sequence identifier, a start and the end position in the sequence. The sequences defined by these regions were extracted from the original genomic sequence file using the program fetchdb.

The consensus regions are assembled into a non-redundant set by using the program phrap. After assembly with phrap a set of contigs and singletons were defined as candidate DNA 30 regions coding for nGPCRs. These sequences were then submitted for further sequence analysis.

Further sequence analysis involves the removal of sequences previously isolated and removal of sequences that are related to olfactory GPCRs.

nGPCR-70

Homology searches were performed with the program BLAST version 2.08. A collection of 286 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.1 were collected from each BLAST search. The amino acid sequences were edited to remove regions in the sequence that produce non-significant alignments with proteins that are not related to GPCRs.

The consensus regions were assembled into a non-redundant set by using the program "GelStart". After assembly with GelStart, a set of contigs and singletons were defined as candidate DNA regions coding for nGPCR-70. Further sequence analysis involved the removal of sequences previously isolated and removal of sequences that are related to olfactory GPCRs. The transmembrane regions for the sequences that remained were determined using a FORTRAN computer program called "tmrest.all" (Parodi *et al.*, Comput. Appl. Biosci. 5:527-535(1994)). Transmembrane regions within the amino acid sequences were visually inspected with an outliner (MaxThink) and the Celera sequences with promising transmembrane regions were kept. In a last step, the blast results were compared against SWPLUS and the transmembrane regions of the sequences and promising Celera sequences for full length cloning were selected. The sequence of nGPCR-70s is shown above in Table 5.

nGPCR-63

Homology searches were performed with the program BLAST version 2.08. A collection of 286 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.1 were collected from each BLAST search. The amino acid sequences were edited to remove regions in the sequence that produce non-significant alignments with proteins that are not related to GPCRs.

The consensus regions were assembled into a non-redundant set by using the program "GelStart". After assembly with "GelStart" a set of contigs and singletons were defined as candidate DNA regions coding for nGPCR-63. Further sequence analysis involved the removal of sequences previously isolated and removal of sequences that are related to olfactory GPCRs. The transmembrane regions for the sequences that remained were determined using a FORTRAN computer program called "tmrest.all" (Parodi *et al.*, Comput. Appl. Biosci. 5:527-535(1994)). Transmembrane regions within the amino acid sequences were visually inspected with an outliner (MaxThink) and the Celera sequences with promising transmembrane regions were kept. In a last step, the blast results were compared against SWPLUS and the

transmembrane regions of the sequences and promising Celera sequences for full length cloning were selected. The sequence of nGPCR-63 is shown above in Table 5.

nGPCR-42

Homology searches were performed with the program BLAST version 2.08. A collection of 340 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.01 were collected from each BLAST search. The amino acid sequences have been edited to remove regions in the sequence that produce non-significant alignments with proteins that are not related to GPCRs.

The consensus regions were assembled into a non-redundant set by using the program phrap. After assembly with phrap a set of contigs and singletons were defined as candidate DNA regions coding for nGPCR-42. These sequences were then submitted for further sequence analysis. Further sequence analysis involves the removal of sequences previously isolated and removal of sequences that are related to olfactory GPCRs. The transmembrane regions for the sequences that remained were determined using a FORTRAN computer program called "tmrest.all" (Parodi *et al.*, Comput. Appl. Biosci. 5:527-535(1994)). Only sequences that contained transmembrane regions in a pattern found in GPCRs were retained. These nGPCR-42s are shown above in Table 5.

nGPCR-46

Homology searches are performed with the program BLAST version 2.08. A collection of 340 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.01 were collected from each BLAST search. The amino acid sequences have been edited to remove regions in the sequence that produce non-significant alignments with proteins that are not related to GPCRs.

The consensus regions are assembled into a non-redundant set by using the program phrap. After assembly with phrap a set of contigs and singletons were defined as candidate DNA regions coding for nGPCR-46. These sequences were then submitted for further sequence analysis. Further sequence analysis involves the removal of sequences previously isolated and removal of sequences that are related to olfactory GPCRs. The transmembrane regions for the sequences that remained were determined using a FORTRAN computer program called "tmrest.all" (Parodi *et al.*, Comput. Appl. Biosci. 5:527-535(1994)). Only sequences that contained transmembrane regions in a pattern found in GPCRs were retained. These nGPCR-46s are shown above in Table 5.

nGPCR-48

Homology searches are performed with the program BLAST version 2.08. A collection of 340 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.01 were collected from each BLAST search. The amino acid sequences have been edited to remove regions in the 5 sequence that produce non-significant alignments with proteins that are not related to GPCRs.

The consensus regions are assembled into a non-redundant set by using the program phrap. After assembly with phrap a set of contigs and singletons were defined as candidate DNA regions coding for nGPCR-48. These sequences were then submitted for further sequence analysis. Further sequence analysis involves the removal of sequences previously isolated and 10 removal of sequences that are related to olfactory GPCRs. The transmembrane regions for the sequences that remained were determined using a FORTRAN computer program called "tmrest.all" (Parodi *et al.*, Comput. Appl. Biosci. 5:527-535(1994)). Only sequences that contained transmembrane regions in a pattern found in GPCRs were retained. These nGPCR-48s are shown above in Table 5.

15 *nGPCR-49*

Homology searches were performed with the program BLAST version 2.08. A collection of 286 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.1 were collected from each BLAST search. The amino acid sequences were edited to remove regions 20 in the sequence that produce non-significant alignments with proteins that are not related to GPCRs.

The consensus regions were assembled into a non-redundant set by using the program "GelStart". After assembly with GelStart a set of contigs and singletons were defined as candidate DNA regions coding for nGPCR-49. Further sequence analysis involved the removal 25 of sequences previously isolated and removal of sequences that are related to olfactory GPCRs. The transmembrane regions for the sequences that remained were determined using a FORTRAN computer program called "tmrest.all" (Parodi *et al.*, Comput. Appl. Biosci. 5:527-535(1994)). Transmembrane regions within the amino acid sequences were visually inspected with an outliner (MaxThink) and the Celera sequences with promising transmembrane regions 30 were kept. In a last step, the blast results were compared against SWPLUS and the transmembrane regions of the sequences and promising Celera sequences for full length cloning were selected. The sequence of nGPCR-49 is shown above in Table 5.

nGPCR-61

Homology searches were performed with the program BLAST version 2.08. A collection of 286 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.1 were collected from each BLAST search. The amino acid sequences were edited to remove regions 5 in the sequence that produce non-significant alignments with proteins that are not related to GPCRs.

The consensus regions were assembled into a non-redundant set by using the program phrap. After assembly with phrap a set of contigs and singletons were defined as candidate DNA regions coding for nGPCR-61. Further sequence analysis involved the removal of sequences 10 previously isolated and removal of sequences that are related to olfactory GPCRs. The transmembrane regions for the sequences that remained were determined using a FORTRAN computer program called "tmrest.all" (Parodi *et al.*, Comput. Appl. Biosci. 5:527-535(1994)). Transmembrane regions within the amino acid sequences were visually inspected with an outliner (MaxThink) and the Celera sequences with promising transmembrane regions were 15 kept. In a last step, the blast results were compared against SWPLUS and the transmembrane regions of the sequences and promising Celera sequences for full length cloning were selected. The sequence of nGPCR-61 is shown above in Table 5.

nGPCR-51

Homology searches were performed with the program BLAST version 2.08. A 20 collection of 340 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.01 were collected from each BLAST search. The amino acid sequences were edited to remove regions in the sequence that produce non-significant alignments with proteins that are not related to GPCRs.

The consensus regions were assembled into a non-redundant set by using the program phrap. After assembly with phrap a set of contigs and singletons were defined as candidate DNA regions coding for nGPCR-51. These sequences were then submitted for further sequence 25 analysis. Further sequence analysis involved the removal of sequences previously isolated and removal of sequences that are related to olfactory GPCRs. The transmembrane regions for the sequences that remained were determined using a FORTRAN computer program called "tmrest.all" (Parodi *et al.*, Comput. Appl. Biosci. 5:527-535(1994)). Only sequences that contained transmembrane regions in a pattern found in GPCRs were retained. The sequence of nGPCR-51s is shown above in Table 5.

nGPCR-52

Homology searches are performed with the program BLAST version 2.08. A collection of 340 query amino acid sequences derived from GPCRs was used to search the genomic DNA sequence using TBLASTN and alignments with an E-value lower than 0.01 were collected from each BLAST search. The amino acid sequences have been edited to remove regions in the 5 sequence that produce non-significant alignments with proteins that are not related to GPCRs.

The consensus regions are assembled into a non-redundant set by using the program phrap. After assembly with phrap a set of contigs and singletons were defined as candidate DNA regions coding for nGPCR-52. These sequences were then submitted for further sequence analysis. Further sequence analysis involves the removal of sequences previously isolated and 10 removal of sequences that are related to olfactory GPCRs. The transmembrane regions for the sequences that remained were determined using a FORTRAN computer program called "tmrest.all" (Parodi *et al.*, Comput. Appl. Biosci. 5:527-535(1994)). Only sequences that contained transmembrane regions in a pattern found in GPCRs were retained. These nGPCR-52s are shown above in Table 5.

15 nGPRCR-x cDNAs were sequenced directly using an ABI377 fluorescence-based sequencer (Perkin-Elmer/Applied Biosystems Division, PE/ABD, Foster City, CA) and the ABI PRISM™ Ready Dye-Deoxy Terminator kit with Taq FS™ polymerase. Each ABI cycle sequencing reaction contained about 0.5 µg of plasmid DNA. Cycle-sequencing was performed using an initial denaturation at 98C for 1 minute, followed by 50 cycles using the following 20 parameters: 98C for 30 seconds, annealing at 50C for 30 seconds, and extension at 60C for 4 minutes. Temperature cycles and times were controlled by a Perkin-Elmer 9600 thermocycler. Extension products were purified using Centriflex™ gel filtration cartridges (Advanced Genetic Technologies Corp., Gaithersburg, MD). Each reaction product was loaded by pipette onto the column, which is then centrifuged in a swinging bucket centrifuge (Sorvall model RT6000B 25 tabletop centrifuge) at 1500 x g for 4 minutes at room temperature. Column-purified samples were dried under vacuum for about 40 minutes and then dissolved in 5 µl of a DNA loading solution (83% deionized formamide, 8.3 mM EDTA, and 1.6 mg/ml Blue Dextran). The samples were then heated to 90C for three minutes and loaded into the gel sample wells for sequence analysis using the ABI377 sequencer. Sequence analysis was performed by importing 30 ABI377 files into the Sequencer program (Gene Codes, Ann Arbor, MI). Generally, sequence reads of 700 bp were obtained. Potential sequencing errors were minimized by obtaining sequence information from both DNA strands and by re-sequencing difficult areas using primers annealing at different locations until all sequencing ambiguities were removed.

The following Table 5 contains the sequences of the polynucleotides and polypeptides of the invention. Start and stop codons within the polynucleotide sequence are identified by boldface type. The transmembrane domains within the polypeptide sequence are identified by underlining.

5

Table 5

The following DNA sequence nGPCR-Seq61 <SEQ ID NO. 1> was identified in *H. sapiens*:

CCTATGCTACAATCGGCAGATGGGAACTAGGAGCCATGATCTCTCAGATTGCAGGTCTCATTG
GAACCACTTTATTGGATTTCTTTAGTAGTACTAACATCATACTACTCTTTGTAAGCCA
TCTGAGAAAAATAAGAACCTGTACGTCCATTATGGAGAAAAGATTGACTTACAGTTCTGTGAA
AAGACATCTTGGTCATCCAGATTCTACTAAATAGTTGCTTCCTCCTTATAGTATTTTAAAC
CCATTTTATGTTCTACACCAAGAGATAACTGTCAGCAATTGAATTATTAAATAGAAACAA
AAAACATTCTCACCTGCTTGGCCAGAAGTAGCACAGACCCCCATTATTTCTTATT
AGACAAAACATTCAAGAAGACACTATATAATCTCTTACAAAGTCTAATTCAAGCATATGCA
ATCATATGGTGTGACTTTGAATGGAAAACCCACAATATTAAGAAAAGCATTGATGTGACTTT
ATTAGGGACACTAAACTACATCATTAACATGTACAG

The following amino acid sequence <SEQ ID NO. 61> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 1:

YATIGRWE LGAMISQIAG LIGTTFIGFS FLVVLTSYYS FVSHLRKIRT CTSIMEKDLT
YSSVKRHLLV IQILLIVCFL PYSIFKPIFY VLHQRDNCQQ LNYLIETKNI LTCLASARSS
TDPIIFLLLD KTFKKTLYNL FTKNSNAHMQ SYG*LLNGKP HNIKKSIHVT LLGTLNYIIN MSQ

The following DNA sequence nGPCR-Seq63 <SEQ ID NO. 2> was identified in *H. sapiens*:

CAGTGAGCCG AGATGGTGCC ATTGCACTCT AGCCTGGGGC AACAGAGCCG ACTCCATCT
CAAAAAAAA AAGGCCATTC TGAGGATCAA GGCACCACTA GCAACAGGG AGCCCATGGG
TCTCAGACCC TCTCCCCACA TCTCTGGTCCCTGCCCCA CCTGGCGTAC AGGGACCAGC
CCCACGGAAG GCTCTTGAGG CCAGGTAACC ATGGGGAGGG GAGGAATGGG GACACCTTCC
TCCCTGAGTGT CTTAGGGAAG AGAAGCTTAG GTCAGGTGGC TGAGGGTGG AATGAGAGAG
GGGTCCTCTC CTGGAGGGTC TCACCATTC CTTGGTCACC CACCAACTC TCATCTCCCC
TGATGTGGGG AGGAGCAGGG GGCATGGATT CCTGAGCCCC AGACTCAACT GTTGTGGTT
ACAGGGGCAT CAGGAGAGAG AGCGAGCAGA ACACACTCT GCAGCATCCC CTGGCCCC
GCCCATGAT GGAGCCAGA GAAGCTGGAC AGCACGTGGG GGCGCCAAC GGCGCCAGG
AGGATGTGGC CTTCAACCTC ATCATCCTGT CCCTCACCGA GGGCTCGGC CTGGTGGC
TGCTGGGAA TGGGCAGTC CTCTGGCTGC TCAGCTCAA TGTCTACAGA AACCCCTTCG
CCATCTACCT CCTGGACGTG GCCTGCGGG ATCTCATCTT CCTGGCTGC CACATGGTGG
CCATCGTCCC CGACTTGCTG CAAGGCCGGC TGGACTTCCC GGGCTCGTG CAGACCAGCC
TGGCAACGCT GCGCTTCTTC TGCTACATCG TGGCCTGAG TCTCTGGCG GCGTCAGCG
TGGAGCAGTG CCTGGCCGCC CTCTTCCAG CCTGGTACTC GTGCCGCCGC CCACGCCACC
TGACCACCTG TGTGTGCGCC CTCACCTGGG CCCTCTGCC GCTGCTGCAC CTGCTGCTCA
GCGGCCCTG CACCCAGTTC TTCGGGGAGC CCAGGCCCA CTGCTGCCGG ACGCTGTGGC
TGGTGGCAGC GGTGCTGCTG GCTCTGCTGT GTTGCACCAT GTGTGGGGCC AGCCTATGC
TGCTGCTGCG GGTGGAGCGA GGCCCCAGC GGCCCCCACC CCGGGGCTTC CCTGGCTCA
TCCTCCTCAC CGTCCTCTC TTCCCTTCT GCGGCCCTGCC CTTGGCATC TACTGGCTGT
CCCGAACCT GCTCTGGTAC ATCCCCACT ACTTCTACCA CTTCAGCTTC CTCATGGCCG
CCGTGCACTG CGCGGCCAAG CCCGTCGTCT ACTTCTGCC GGGCAGTGCC CAGGGCCGA
GGCTGCCCT CCGGCTGGTC CTCCAGCGAG CGCTGGGAGA CGAGGCTGAG CTGGGGGCCG
TCAGGGAGAC CTCCCGCCGG GGCCTGGTGG ACATAGCAGC CTGAGCCCTG GGGCCCCGA
CCCCAGCTGC AGCCCCCGTG AGGAAGAGG GTGACGTGGG GAAGGTGGTG GGGTCAGAGG
CTGGGGCCAG CCGGACCTGG AGGAGGCCTT GGTGGGTGAC CCGGTCTG GCTGTCAAAG
TTGTGACCTT TGGTCTGGAG CATGAGGCTC CCCTGGGAGG CAGCTGGAAA GG

The following amino acid sequence <SEQ ID NO. 62> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 2:

VSRDGA IAL*PGATEP DSISKKRPF *GSRHH*QQG APWVSDPLPT SPGPCPHLAY
 RDQPHGRLR PGNHGEGRNG DTFLSVLGK RSLGQVAEGG NERGVSSWRV SPFPWSPTQL
 SSPLMWGGAG GMDS*APPST VVYRGRRE SEQNTLLQHP LAPRPMMEPR EAGQHVGAA
GAQEDVAFNL IILSLTEGLG LGGLLNGAV LWLSSNVYR NPFAIYLLDV ACADLIFLGC
HMVAIPDLL QGRLDFFPGFV QTSLATLRF CYIVGLSLLA AVSVEQCLAA LPPAWYSCR
PRHLLTCVCA LTWALCLLH LLLSGACTQF FGEPSRHLCLR TLWLVAAVLL ALLCCTMCGA
SLMLLLRVER GPQRPPPGRF PGLLILTVLL FLFCGLPFGI YWLSRNLLWY IPHYFYHFSF
LMAAVHCAAK PVVYFCLGSA QGRRLPLRLV LQRALGDEAE LGAVRETSRR GLVDIAA*AL
GPPTPAAAPV RQEGDVGVKVV GSEAGASRTW RRPWWVTRSC AVKVVTLGE HEAPLGGSWK

The following DNA sequence nGPCR-Seq64 <SEQ ID NO. 3> was identified in *H. sapiens*:

AAAATTGCTC TTCCCTCTGA GCTTGTACAC AATGATTGAG TTCAAGATGA AGAAGATGGA
 GCAGGGCACC AGGTAGACGG TGAAGCAGTG GATCCAGATG AGGACGTGAT GCACAGAGGT
 GCTGATGTAG TCTTCAGTCC AGATGTTGGG CCACCAGTAA TAGGGGATGC TGGTCAGGAA
 GCAGGTGATG TAAACACTTA CAATGACTTT CCGGGTGCAG GCTGGGTATG AGACCGTGTG
 GTACTTGAGC GGGTGGCAGA CAGCGATATA CCTGTCAATG GTIAACGGTA CAGTAATCCA
 TATGGAGGTG TGGATGGATG AGAATTCCAG CACTTCTATG ATCTTGTCCGG GGACCTGAGG
 CATCTGCATG TTCAAGATGA AATCTTCCAA CAGGAAGTCC ACAAACACTA TGAAAAAGAG
 GACCAAGATG TCGGCAGCAG CGAGTGCCAA GAGATAGTTG TAGGAGGACT TCTGCTTCT
 TGCCACCAGC TGGGAGAGGA TGATCACTGT CAAGATATT GCTGTGGAGA GAAGAAAAAC
 TGGTTTAGCT CTGAAGCAAA GATGACTTCG TTGGCTCTA TGGGGGCCCT AGGCATATGT
 TTATTITGCA CTCCCATGGA AGTAAAATG ATTGAATCAA TGCTTTGAG GGACAAACCTC
 AGCATTACAA ATAGCACCTC ATACAATTAG TGGATACTAT TTAAAGTTA TGCTTATATT
 CTAACACAAAC CATGAGAGGT GGTGCCTCCA TTCTCCTCAT CTTAGAAGTG AAACGGGC
 TCTGAGAGCC TCACACAGCC ATGAGAGGTG GTGCCGCCAT TCTCCTCACA CAACCATGAG
 TGGTGTGCC GCCATTCTCC TCACACAAACA ATGAGAGGTG GTGCCACCAT TCTCCTCACA
 CAACCGTAG AGGCGATGCT GCCATTATCC

The following amino acid sequence <SEQ ID NO. 63> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 3:

G* WQHRLSRLCE KNGGTTSHCC VRRMAATPLM VV*GEWRHHL SWLCEALRAP
 VSLLR*GEWR HHL SWL C*NI SITLK*YPLI V*GAICNAEV VPQKH*FNHF HFHGSAK*TY
A*GPHRSQRS HLCFRAKPVF LLSTANILTV IILSQLVARR QKSSSYNYLLA LAAADILVLF
FIVFVDFLLE DFILNMQMPQ VPDKIIEVLE FSSIHTSIWI TVPLTIDRYI AVCHPLKYHT
VSYPARTRKV IVSVYITCFL TSIPYYWWPN IWTEDYISTS VHHVLIWIHC FTVYLVPCSI
FFILNSIIVY KLRRKSNFDN GSIA SHGCVR

The following DNA sequence nGPCR-Seq65 <SEQ ID NO. 4> was identified in *H. sapiens*:

GGCCCGCTCCCACGCTGTAGTGTACTTCATTTCATAATAAATCACTTCATCCCTCCT
 TGCTTGTGTTGCGTTTGTCCAATTCTTGTCAAGACGCCAAGAACCTGGACAGCCT
 CCACCATTAAGAAATACAAGAGCAGTTCTGTACATGTACATATGGGGTGGGAGATCT
 CGCTCAGCCTTCCAGGACACAGCGGTTGATGGAATGCTTTCTGAAACTCGTGGCAGA
 ATGAGTACGGAAGAGGGCTGATAGCCGATTCAGCCAGTGGAGCCAGAACGCGTGTCTCAT
 AGAGAAAGTGTGGACATGTGACCCGTGGCAGGCTCCGATGATCACGAGCAGTGTAG
 GGAGGCCAGCACAGGACAAATGACACACCATGACGTCAAGCGACGTGGACGCTCATTTG
 TCCCGGGAGACGCCGTGGCTGGGAGGCTGGCTGTGCTGCCATTGCAAGGCCTCTCGC
 TGCCCTGTAGCGGGTCTTGTGCTAGGATTATCAAGTCCACCTGCAGAGGTGA

The following amino acid sequence <SEQ ID NO. 64> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 4:

HLCRWT**SY AEDPLQGSEN ACNGSRQPSL PSTASPGTNE RPRRLTWC HLSCAGLPTL
 LVIIGAACHG SHVQHFLYET RFWLHWLNSA ISPLPYSFCH EFQKSIPSNR CVLERLSEIH
 PPPYVHVTET ALVFLMVEAV QVLGVLNKEL DKTHKQSKEG MK*FIENEST LHSVAG

The following DNA sequence nGPCR-Seq66 <SEQ ID NO. 5> was identified in *H. sapiens*:

TACTTTACCCAAAACATTATGTCATTATTAGAGTTCTAGAAAAAATACCTAAGGAG
TTAATGGTTCATATCTTTGCTCTATTATGAAGAAGAAAATGTTTACAAATATTTC
ATTATTGGAGCATTTTGTTAGTGAATTATCAAACACTAGGATTGATTCTATTCT
GTTACTTTGTTATAATCTTATCCTTCTCTTAATTCTGTATTGGATGCCAAC
TTAGAATACATTACCAAAGTTACCTTTCATTTAGTCTCTCAATACAAGATGATTAAAA
CATTTATGGTTACCTTTTAAATTCTTCTGCTATGCAAATTATAAAAGGGCAAAGTCTT
TGTGCTCTAAATAACCTGCTTCTCATGTTTACATGTTCTACGATTATTGTTT
ATAATGTAATTTCGTTACCTAATTGTGCACATAGTGAATAATAGATTATAATGAAGAA
AACTTGAGTAAATCTATTGTTAAAAGGTTTCTAGGCAATAATAATCATTGGATT
TTCTGTGTTATTAAAAGATATGTTATTGAGCAACTCGTGTGC

The following amino acid sequence <SEQ ID NO. 65> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 5:

YFYPKLLCSL LEFPRKIPKE LMVSYLLLYY EEEKCFNTIF IIGAFFVVSE IIKTRIDFYS
VYFCYNYLPF LLISVFWMPN LEYITKVTFS FSLSIQDDLK HLWLPFLIFL LCKFIKGQSL
CALIIPAFSC FTCSTYFVF IM*FSFT*LC T**IIDYNEE NLD*NLLLKR FFRQ**IIGF F*CILKRYVY
F*ATRV

The following DNA sequence nGPCR-Seq67 <SEQ ID NO. 6> was identified in *H. sapiens*:

GGCTACTTGT TATGGAAAAG TTATGATTGG TGTAGAACGT TACAGGTGTC TTCTGGCACT
TGAAGAAAGC CTTGAGCGTG GCGGCTGCCA TGGTGCAGAA CCGCTTGCTG ATGAAGCAGT
AGAGGAAGAA GTTGATGGCT GTGTTAGCAA GGGCTAGCAT GTTGGCAATG TCGGACATGA
TGTGTACCAAG CCAGCGGTTTC TGGATGGCG CCCCCATAGAG GTGGTAAAGA ATCATGATGA
TGCGGGGGGC CCAAAGTGTG GCAAAGATGG AGGTAATGGT GAACAAGATG GCGGTGGTCT
TCCCCGTGGA GTAGCCACGG AGACGAAAAT TGCTCTTCTC CCTGAGCTG TACACAATGA
TTGAGTCAA GATGAAGAAC ATGGAGCAGG GCACCAAGTA GACGGTGAAG CAGTGGATCC
AGATGAGGAC GTGATGCACA GAGGTGCTGA TGTAACTTC AGTCCAGATG TTGGGCCACC
AGTAATAGGG GATGCTGGTC AGGAAGCAGG TGATGTAAC ACTTACAATG ACTTCCGGG
TGCGGGCTGG GTATGAGACC GTGTGGTACT TGAGCGGGTG GCAGACAGCG ATATACTGT
CAATGGTTAA CGGTACAGTA ATCCATATGG AGGTGTGGAT GGATGAGAAAT TCCAGCACTT
CTATGATCTT GTCGGGGACCG TGAGGCATCT GCATGTTCAA GATGAAATCT TCCAACAGGA
AGTCCACAAA CACTATGAAA AAGAGGACCA AGATGTCGGC AGCAGCGAGT GCCAAGAGAT
AGTTGTAGGA GGACTTCTGT CTTCTGCCA CCAGCTGGGA GAGGATGATC ACTGTCAAGA
TATTGCTGT GGAGAGAAGA AAAC

The following amino acid sequence <SEQ ID NO. 66> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 6:

FLL STANILTVII LSQLVARRQK SSYNYLLALA AADILVLFPI VFVDFLLEDIF ILNMQMPQV
DKIIEVLEFS SIHTSIWITV PLTIDRYIAV CHPLKYHTVS YPARTRKVIV SVYTICFLTS
IPYYWWPNIW TEDYISTSVH HVLIWIHCFT VYLVPCSMFF ILNSIIVYKL RRKSNFLRLG
YSTGKTTAIL FITTSIFATL WAPRIIMILY HLYGAPIQNR WLVHIMSDIA NMLALLNTAI
NFEFLYCFISK RERTMAAATI KAFFKCOkop VOFTYTNHNES ITSS

The following DNA sequence nGPCR-Seq68 <SEQ ID NO. 7> was identified in *H. sapiens*:

GACACCTGCC AACATGTTCA TTATCAACCT CGCGGTCAAG GACTTCCTCA TGCCCTTCAC
CCAGGCCCT GTCTTCTTCA CCAGTAGCCT CTATAAGCAG TGGCTCTTG GGGAGACAGG
TAGATGCTGG GGCTCCCTT TGCTGGAGGG AGGAGGAGGG TTTGACCTG GGGATGCCCT
CAATGGAGGG TGGCCCAAAG GAGGTGATTG GCTGCTTCTG GGCAGAGAGT GGGTAGCTGC
CCTCAGTCTGTGAGTAAGC AAGAAGGGAA GATGCAGTGT TGGTCTTAAG GCCTCTGCCA
GCCTTGGCCA GATGTGGCAG GTGGAGGGGG TGGAGTGCAG TCAGTCTGC TCTTCTGTG
AGGTGAAGGC CAGAGCAGAG TCTACCCCTGT CCCCCAGACCC TCCTCCCCAG GACTCAGAGC
AGGGGCTGTG CCCACAGGGT GCGAGTTCTA TGCCTCTGT GGAGCTCTCT TTGGCATTTC
CTCCATGATC ACCCTGACGG CCATGCCCT GGACCGCTAC CTGGTAATCA CACGCCCGCT
GGCCACCTT GGTGTGGCGT CCAAGAGGCG TGCGGCATTG TGCCTGCTGG GCGTTGGCT
CTATGCCCTG GCCTGGAGTC TGCCACCCCTT CTTCGGCTGG AGTAAGTGGG CTGCTGGAAAC

TGGAAGGGG GCAGATGGC TGGGAGGGC ACATTCAAGG GTAAGTAGGT GACTTGGGT
CAGCCAGCTG CGGGGACAG GGTGCCAGG AGCTACCTGA GCCTCAGGTG AGATGGACAT
TCAGGGGAC ATGACTGGCA GCAAGGGAAA CTGACACTGC CCCA

The following amino acid sequence <SEQ ID NO. 67> is a predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 7:

TPANMF IINLA VSDL MSFTQAPVFF TSSLYKQWLF GETGRCWGSL LLEGGGGF*P
GDALNGGWPK GGDLLLLGRE WVAALSPVSK QEGKMQCWS* GLCQPWPDV A GGGGVRSVL
LFL*GEGQSR VYPVPRPSSP GLRAGAVPTG CEFYAFCGAL FGISSMITLT A IALDRYLVI
TRPLATFGVA SKRRAAFVLL GVWLYALAWS LPPFFGWSKW AAGTGRGADG L GGAHSRVSR
WTWVSQLAGA GCPGAT*ASG EMDIQGDMTG SKGN*HCPHL

The following DNA sequence nGPCR-Seq70 <SEQ ID NO. 8> was identified in *H. sapiens*:

TTCTCCCCITGACGGGTGACTAACTCTGCCTGCGTCTTCTTTGTCAACCAGCATAGGCA
CTGAGTGGCTGTGCACCCCTTGCACCCACCGGTGCCGGACTGAGCCTGCAACCT
GTCTCACGCCCTCTGGCTTGTGCCATGACGTCCACCTGCACCAACAGCACGCCGAGAGT
AACAGCAGCCACACGTGCATGCCCTCTCCAAAATGCCATCAGCCTGGCCACGGCATC
ATCCGCTCAACCGTGCTGGTTATCTCCTCGCCCTTCTCGTCGGCAACATAGTGCTG
GCGCTAGTGTGCAAGCCGAGCTGCTGAGGTGACCAACCGTTATCTTAACT
CTCCTCGTACCCGACCTGCTGCAGAATTGCTCGTGGCCCCCTGGGTGGTGGCCACCTCT
GTGCTCTCTCTGGCCCTCAACAGCCACTTCTGCACGGCCCTGGTTAGCCTCACCCAC
CTGTCGCTTCGCCAGCGTCAACACCATTGCTGGTGTCAGTGGATCGCTACTTGTCC
ATCATCCACCCCTCTTCTACCCGTTCAAGATGACCCAGCGCCGGTACCTGCTCCTC
TATGGCACCTGGATTGTGG

The following amino acid sequence <SEQ ID NO. 68> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 8:

FSP*RVNTSA CVFLLSPA*A LSAVCAPLCH PPVPALSLQP VSRPLAVAMT STCTNSTRES
NSSHTCMPLS KMPISLAHGI IRSTVLFVFL AASFVGNIVL ALVLRKPQL LQVTNRFIFN
LLVTDLLQIS LVAPWVWATS VPLFWPLNSH FCTALVSLTH LFAFASVNTI VVSVDRYLS
IIHPLFYP SK MTQRRGYLLL YGTWIV

The following DNA sequence nGPCR-Seq71 <SEQ ID NO. 9> was identified in *H. sapiens*:

AGGACAACGA TGGTCACTGATTGGTGA CTCGACAGTCTCCGGCGCTGGCTCCGGTC
GGCGTCTCCGGCTACCGCGGCCCTCCTTGGTCCCCGCCGGCGGGTCGGCGATG
AAGCGCACCAGCAGCAGTAGCACAAGATAATGATGCCAGCGGCAGCACGAAGCCAGC
AGCACCTCTCGAGTGGTAGAGGGCCAGCCAGAACTGCCTGTCGGGCCAGCAACTTG
TCCGGAAACCGCACCAGGCACAGCTCTGCCATCACCTGACCGTGGAGAAAATG
GCACTGGCAGCGAGGCCAGCGGCCAAAGCCAGATCCACACACAGCGCTTGGCC
GAGAACAGCAGCTGCCCCAGGCTCCGGCGCAGCAGTCGCCCGGGCCGTCTCGG
GTCCCGGTGGCTCTCAGAGCCAGGGCACCGAATGGTAGCGCGTCACACTCATGGCAGT
GAGGAAGAACACGCTGGCGTACATGTTATGGACGTCACCATGGACACGATCTTACACA
TGGCCTTGCC

The following amino acid sequence <SEQ ID NO. 69> is a predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 9:

QGHV*D RVHGDVP*TC TPACSSSLP* V*RATIRWPR L*RATGTRGH GRGDCCGRSL
GDSCCFSAKA LCVWIWALAA LASLPSAIFS TTVKVMGEEL CLVRFPDKLL GRDRQFWLGL
YHSQKVLLGF VLPLGIILC YLLLVRFIAD RRAAGTKGGA AVAGGRPTGA SARRLSKVTK
SVTIVV

The following DNA sequence nGPCR-Seq69 <SEQ ID NO. 10> was identified in *H. sapiens*:

CTGGAAAGGG TGCTGGTGGC GCGCCGCGAC GCCGCGCGC GACTGCCTGC CTGGTACTCA
ACCTCTCTG CGCGGACCTG CTCTCATCA GCGCTATCCC TCTGGTGTG GCCGTGCGCT
GGACTGAGGC CTGGCTGCTG GGCCCCGTTG CCTGCCACCT GCTCTCTAC GTGATGACCC

TGAGCGGCAG CGTCACCAC TCACAGCTGG CGCGGTCAG CCTGGAGCGC ATGGTGTGCA TCGTCACCT GCAGCGCGC GTGCGGGTC CTGGCGGCG GGCGCGGCA GTGCTGCTGG CGCTCATCTG GGGCTATTG CGCGTCGCCG CTCTGCCTCT CTGCGTCTTC TTCCGAGTCG TCCCGCAACG GCTCCCCGC GCCGACCAGG TGAGCGCCC TCTGTGTGTG CGGGCAGGT GTCCTGCGCA GGCTGGGAAG CGGGGCCCCG ACGGAAGCTG GGATGAGGAT GATCAAGAAC ACAAATAGCC ATTATTGCA CTTAATCGTT GTGCCAAATC TTGTGCCAT GGCTGTGAAG TTAAATCTCT TAAATCTCAC TACAACGCTG TGACACGCC CTCCTAAATG ATGTAAGTGG AGTCCCCAA ATTCTTGCAC AATGCAATGA CTGTTGCGAG GTTAATTAAC GAGTAGTTA GGAGCGAGAC GGAACCTTGG GGGTCAGGG TGCCAAACA CTTGTATTG AATCATGATT CCTCGCCAGG TGCTACAATA CTGTTATTAT CACACCCATT TCACAGATGA GAACCAAGAGG CACACCGAAG TGTATAATAA CTGCCCAGA GTATTTATC CGTAATTCA GGAGGAGATG GGCTCCTTCC AGAAGTTAC CCGTAATTCA AGGAGGAGTT GGGCTCTGT CCAGGGTTGG GTTATGGTCC TGCTTTGAAA GCGCGCGAC AGGCATGTGA GACCCGGGGA CCCCCAGATGC AATGCTGTCT TTAGGGACT TGTGACAGAA TTCCCTTCG GGGCTCTCAG TTTTTTCAGC TGCAAAACGG AAGGATTACA CTAGACCTTC GAGGTGTCCT GGGCGCTGA AATGTGCAGA TTACAGAGGC TGGACCGACG AGCT

The following amino acid sequence <SEQ ID NO. 70> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 10:

GKGAG GAPRRRGATA CLVNLFCAD LLFISIPLV LAVRWTEAWL LGPVACHLLF
YVMTLSGSVT ILTLAAVSLE RMVCIVHLQR GVRGPGRRAR AVLLALIWGY SAVAALPLCV
FFRVPQRLP GADQVSAPLC VPGRCPAQAG KRGPDGSWDE DDQEQQ*PFI ALNRAKSCA
HGCEV*SLKS HYNAVHTPS* MM*VESPKFL QNAMTVARL I NE*FRSETEL WGCRVAKHFV
LNHDSSPGAT ILLLSHPFHR *EPEAHRSV* *LAQSILSVI RGGDGLLPEV YP*FKEELGS
CPGLGYGPAL KARGQACETR GPQMQCCL*G TCDRIPFRGL QFFQLQNGRI TLDLRGVLGA
*NVQITEAGP TS

The following DNA sequence nGPCR-Seq2011 <SEQ ID NO. 11> was identified in *H. sapiens*:

TTAATCCCTGGAAAGTCCACGAACAATGAATCCATTCTCATGCATCTTGTGGAACACCTCT
GCCGAACCTTTAAACAAATCCTGGATAAAAGAGTTTGCCTATCAAACGTGCCAGTGTGGTA
GATACAGTCATCCTCCCTCCATGATTGGGATTATCTGTTCAACAGGGCTGGTTGGCAAC
ATCCTCATTGTATTCACTATAAAAGTAAGGAATGGCTCTTTTTTTTTTTCTTC
CATACTTAAAGAAAATCACAGTCAAAGCTCCCTAAATGAGTCCTTCCCTGTAGCATT
GCTTAATGAAATGCAATTGGAAATATTGCTTAAGATAATTAAATGAAGATTCTACAGA
TATTTCGTCATGCAATTAGTAACATCTCAGTTGCAAATCTCAACATGCTAACAGCTAGG
CCAATGCTTACTGCTGGGTCACTGAGTTTAAAGGAAATGACTCTCACTCTCAGTCTTAG
CTGCATATTAGAATCATCTGGGAGCTTAAACACTCTGATATGCAGTTCACCCAGA
CCCATTAACTCAGAACATCTCAAGGGTAGGGCCCGGGTAAGATTAAAATG

The following amino acid sequence <SEQ ID NO. 71> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 11:

LIPGSPTTMN PFHASCWNITS AELLNKSWNK EFAYQTASVV DTVILPSMIG IICSTGLVGN
ILIVFTIIR* GMAPFFFFSF HTLGNYSQSS LNESFPL*HF A**NAILEIF A**DN**RFYR
YFRHALGNIS VANLNMLRPR PMLTAGSVSF *GNDHSQS* LHRIIWGAL KTPDMQFHPR
PINSESLRVG PG*DLKL

The following DNA sequence nGPCR-Seq2012 <SEQ ID NO. 12> was identified in *H. sapiens*:

GGGTGACAGGAACCTGGAAGGTGATAGTTTCCAGGTGAAGAAAGGGACAGTAGGTTCC
CTGCTTACTGCTATCTCCCTATGGCCCTCGGTCAAGGCCGTGAACCTGTGATGAGAGTCCT
GCCCTGCTCTTGGATTCCCTCAAATATCCTCCCTCATTTGTTCCCTGAATTATCTAGA
TCAATACAAACCAATCCCTCTGCTTAAACTTTAATTGCTACAGGGAAAGTGCA
GACTCCTTAGCTGGCGACAAGCCTTTGCACCTGGCACACATAGAGCTTCTAGGGCTA
TCTCCTGCCATTCTCTAGCCTGTCTCCCTCTGCTGGTCTCCAGAACTGCCATGAAT
TCCCTCACATGCCCTTCT

The following amino acid sequence <SEQ ID NO. 72> is the predicted amino acid sequence derived from

the DNA sequence of SEQ ID NO. 12:

VTGTLEGDS FPGEETVGF PALLSPYGL RSGLNCDESL CPCSLGFLKY PPFIVP*II*
INTNQSLCL KLLIAYRGKC RLLSWRTSLL HLAHIELSRA ISCHSLACLL LLWVSRTAHE
FPHMPF

The following DNA sequence nGPCR-Seq2013 <SEQ ID NO. 13> was identified in *H. sapiens*:

CCTATGCTTCCTATAGCTCATGGACCAAGCCATTCTCCAACATCATAATTCTTCCCTTA
TCATCCCTTCCTTGCATTATTCTCCAGATGAAAAAAGTCAAATGCCTCCCTCCCTATT
TCAGCTGAGAAGGTGAGATGCCGTGCTGGATTTCAGGCTCTGCCTGCTTGGCAGAC
CACTGTGACCGTGCACGACTAAGTCCACTCTTCCCAGCCAGCATCCCCTGCTCTCCCCAG
GCCAACTTCTTGTGTTCTTCATTCCTATCTCCTGCCGGAACTACTCAGACCCCTCT
GTCTTCTGCCCTTGCCTTGCCTTGCCTCCAAAA
ATGATTTAAGCATTGTGAGTCTTCTCCACAGAACTCCAGACTCTTGAAGATGCC
GTAGTTGTAAATTACTTCAATATTAAGAAGTGCATGCTTCCCTGCCGGTCA
TGCTCTTGGTATTGATTTCTT

The following amino acid sequence <SEQ ID NO. 73> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 13:

YAFL*L MDQAISSPT* FFLIIPFLAL FSR*KKFKCL LPIFS*EGEM PVPFGPSCL LGRPL*PCTT
KSTLPSQHPL LSPGQLLCVL FIPISLPELL RPLCLSASCP IFQALVCWLS ASKNDFKHL*
VFLSTELQTL *KCRSS*ITS NI*EVLIMLP CGFMLFGIDF

The following DNA sequence nGPCR-Seq2014 <SEQ ID NO. 14> was identified in *H. sapiens*:

CAAATGAGGTGGATGCACTTAGAAGGAAGGGTAGAACAAATATCGTTGATGATATAACAA
TGTGTCAGATACAGAGATACACAGACTAGGGCTGGGAATCTCAGATTAAATTGTCA
ACAGTGTGTTTGATTTTTGTTGTATATTGCCTCCCCAATTAAATTACAGGCTT
AATCATCTTACAAGACATTATTAAAGAGAAAGCGAACCTACTGAAGTTTATGCTTC
CCTGATTGTGATGAGCTGGTGTAGCTAGTTCAATGTTCTGGAAAATACTGAA
GTACTTCCACCTGGTGGCACTTAGTGAACATTGCAGAACCGAGTAATAGGTATTCCGTT
GGGTTCTCGAACAAATCTGAGTTAGCTAGGAACTCTCAACTAACAAATTATGAGAAC
CTTCTGCTACACATGTGAATTACATAATTATTCTCTTGCAAGTGAAGGATGGATGTAT
GCAAAGGAGAAATGAACTCTACTACAGTAATGGAAAGAAGTGTAGAACATGAAACTCCAA
AATGCCTATCTGCTACTGAGTGGAACTCACTAGAAATATCAAATGCAAGAGAACAT
GTGACTAGGCTCTAGTCAAACAAATTGTCAAAACTTGCTGAATATATAACACATT
AGGGAGCTGTG

The following amino acid sequence <SEQ ID NO. 74> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 14:

K*GGCT* KEG*NKYSLM IYNVSDTEIH RLGLWESQL ICGQCVLIFL FCIFASPIINF TGLIIFTRHY
FKEKANLLKF YASLIVMSWL ILALVSMFWK ILKYFHLVAL SEHCRTE**V IPLGFSNKSE
L*LGTLN*QF MRTFCYTCEF TYLFLCS*RM GCMQRRMNST TVMERSDRR* NSKMPICYLS
GTH*NISNAE RNM*LGSSQT IVIKYLLNI* HILGSC

The following DNA sequence nGPCR-Seq2015 <SEQ ID NO. 15> was identified in *H. sapiens*:

AAAAAAATACATTCCTGTATGGAAGACTAAATATTAAAGAAAAAACAGTTATCCATATG
TTGTTCATAGCTTAAGTATCCTAGTCTAAATCCAAAGGGACTTTTGGGACTCAA
AAAAAAAATTCTAAAGTTGATCCAGAAGAGTAAAGAGACTAATATAGTCAAATAATAT
TTGAAAAGGTTAAATAATTGGGAAACATGCTTAGCAAAATATAAACAAACAAACT
TCAAAACAGACAAAAACCTATGAAAACCAAAACCTATTAAAAACTAAATTCTAAATG
AGAACTAACATACAAATCAAGAACATGCTAGATAGCTAAGAACAAATTAGCATATA
AACTAATTCAATTATTATAAAG

The following amino acid sequence <SEQ ID NO. 75> is the predicted amino acid sequence derived from

the DNA sequence of SEQ ID NO. 15:

FI IN*ISLYAII CFLAIYADS* FVC*FSFRI* FLNRFLVFHR VFVCLKFCLF IFLLSMFSHN
YLTFSNIIL T ILVSLLFWIN FRFFFESPK KSLWILD*DT *SYETTYG*L FFSLIFSLPY RNVFF

The following DNA sequence nGPCR-Seq2016 <SEQ ID NO. 16> was identified in *H. sapiens*:

CTAGGCTGCTCTGTGTGGTTATAATGAATCTCATCAGCTACAATCATTCCAAAAAGG
 ACAGAAAAGTGAATCTGTCTATAAAAGATGGTTTCAAAGCCAACATACAGATATCATAGAA
 AAAAAAAAACAGTGGCAGATAAGACCTTCATCTTTCTTATCCCATGGGCTTT
 CTGCCTCCAGATTCCTGCACATCAAGGGGTTCTGTCAAATCAGTGGGCTGGCCCT
 CTGCCAACCTGGGCTAACGCTGTGGTGCATGGCTACGAAGACAAGAATGACCCCTGGT
 TTTATAGAGCCCTGGGTCCCTGCTCACAGGCCTTTCAGTTGATATTCCTTCATCTC
 TTCAGAACCCATGTGGCAGGTGCAACAAGGAGTTCAACATGCTCCGGCAAAGTGAGT
 AATGAGGGACACATGCACGACCAGGAGCAGGCAGCCCTGAAGAAGTGGTTGTGAGGGTGT
 GGAGGAAGGGCTTGCTTCCACACTCTGCACTCTGGGCTCTAAGGCACAAACACCTG
 GCTCAGGCATCATCTCAGCCTCATGCTACTTTGTCCAGGGACAGAACAGAGGTGCA
 GCACTCAAGGGAATCCTGCACCACAAACAGGCAGGGACCATCATGACTGCTGAAATAGGC
 AGCAGGAGTCCGAACTATGGCATAAAATGTCACAGCAACACCACAGGGGGAACCCCTG
 GGGGAAGTAGGGCATGGTGA

The following amino acid sequence <SEQ ID NO. 76> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 16:

RLLLCG YNESHQLQSF PKKDRK*ICL IKMVFKANYR YHRKKKKNSG R*DLSFSYP
 MGFSASRFPC TSRGFCANQW AWPSAHPGLS CGCHGYEDKN DPWFYRALGP
 LLTGLFS*YF FHILLQNPWMQ VQQGVHQCSG QSE**GTHAR PGRAALKW
 VRVWRKGLLP HSALLGPKAL ITPGSGIILS LMLLLSRDR RTGASTQGNPA
 PQQAGDHHD C *NRQQESELW HKNVTATPQG GTPGGK*GHG

The following DNA sequence nGPCR-Seq2017 <SEQ ID NO. 17> was identified in *H. sapiens*:

ATTATCTCATTATGCTCTATCAGTTCTTATAGAATATGATTACTACTTAGTGATAAA
 GCTTCCTTGCTAAGATTCAGCCTACGAACCATGATCCAACCCCTACTCTAAACATA
 TAAACATGCTTACAAGTATCCTATATATGGAAAAGTCCTGGAAATTATTGGTAATT
 ACCTGGTATTACATGTGTATTCTGATCTACTATTGCAAATTGACA

The following amino acid sequence <SEQ ID NO. 77> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 17:

SIC NSRSGNNTCN TRLITQIIPR FPYIGYL*S MFICFRSRVW IMVRLKS*Q RKLYH*VVII
 FYKETDRA*W FPYIGYLAS MFICF

The following DNA sequence nGPCR-Seq2018 <SEQ ID NO. 18> was identified in *H. sapiens*:

TCTTGATTACACAAAAACCATAACTAACTAAAGAAAAACTTTAATTATCTGTATT
 GTTGTATAAAATGTTTATATATTAGGAATCTAAAAATTGTTTTGCTTTATCCCT
 GTCATAGGAAGAAGACATCATGTCCTGTTACTGTAACCCTAAATAAAGGCTAAATAACA
 GACAGTACATGATGATACTTTAACTAGGGCAAACAAAAGTAATATTAAACATGAGGT
 TTGGTCCTTGCTATCTATAACCTCATGCTAATTCTCCTACAATGTAATGTCATTCCCTC
 CTCTCTACCCATTGTAAGGGCTCAGTTCTGCTCTGCTGACTTATTAAAGGGT
 CACAATAAGGCCAGGTAATTCAATTAAAAATTCCATTAGAATAATTACATCTAAAA
 ATTCAACAAGAAAGACAATTCAATATAAAATAAAATTACTAATATTGGAATTCAAGC
 ATTAGTCATGGAAAAAGAGATAATTGTAAGCAGAATATTAAATGGCAACTTTCTTAT
 TCTATCACATTGTTCTATTGTTATGACCAAAGAAATTACTCTATATCCACTACAA
 TTCATAAAACAGGCATGGAAGAAGTCTTTCTGGTGCATGCTAAGAAGATGA
 ACCTCAGAAGTATGTCATTTCATACTATGTTCTGAAACAGACAGCACACATTATTT
 GAATGGACACCAATCTCAAACATATAGA

The following amino acid sequence <SEQ ID NO. 78> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 18:

LIYTKHNH*L K*KTFNYLYC CIKCFIY*ES KNLF A FILC HRKK T S CLCL L*PLIKAN NR
QYMMILLTRA NKS NILT MMRF GLCY LY LMSN FP Y NVN V I P P L Y P F V R V S V F C S C M T Y F K G S
Q*GQV I H I L K I P F R I I T S K N S Q E R Q F Q Y K I I N Y * Y W N F K H * S W Q K R D N L * Q N I L M A T F L F
Y H L L C S I C Y D Q R N Y S I S T T I H K T G M E E V F F F L V L M S K K M N L R S M S F N T M F * T D S T H Y F *
M D T K S Q T Y I

The following DNA sequence nGPCR-Seq2019 <SEQ ID NO. 19> was identified in *H. sapiens*:

The following amino acid sequence <SEQ ID NO. 79> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 19:

FILTI YY*QRNLRKQ KFRI*MFL*T CVSFFTUVLE LSYPWQLILL *LYSLFIVLT LKLRT*SNVN
LDELKTENVC KYV*KYVYKN MYF*SYF*KS FILYITHHTHT HTHTM*RSLL TTQYKIJFLR
NIVFKYCFIP YKSNLWLFG FHQAMSL*TN FANKG*TQ*G MKYLLTNKKP SNSMYVIGKI
KSS*VNSIHE LTSI*SALLS LKIS*NSLKI IR

The following DNA sequence nGPCR-Seq2020 <SEQ ID NO. 20> was identified in *H. sapiens*:

TTCTAAATCTTGTACATTCTTACTCTGAGGGGTTTAGTCATCTGCTAGTAA
CTAGTTAGTCCTTCCGTCACTGGGAAAAAGCTATAAATGGTTTCAACTCTATGCAG
CTCCCTGGGTTGCTCATTGATCTGAGCCAGGTTGAAGTGGGCCATTGAGAATGATC
TGTCTCATGATGTAGGGGGTGAECTCCAGCCAATTCTAATATTGCTGGAATTAAAC
TTGATTCCCTCACTGAAAGAGGGCATTTACTCTACCGTGTATTAAATACCTTAT
AACCTGATTTTTGATTTGTTCTGCTCAGTGAGTACTAAAAATCCCTGCCGAT
ATAGTAATTGTTCCCTGTTAATATGGCTTTCTTGTAACTCTGGACTCAAGTCT
TTGTAGCTGTTTCTAAGTCAGCTAAAAGTGTGGTCAATTGGATTGGCAGCCTT
TCAAGCTTACCCACCAAGAATCCTACAGG

The following amino acid sequence <SEQ ID NO. 80> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 20:

SK SLLHSSLL*G VLVIC**TS* SFPSAG*KAI NGFQLYAAPL GCSFDLSQV* SWAIENDLSS
*CRG*LQPII NIAGIN*L*F L*LKEGILLY RVLLNTL*LD FFDFVFLLSB YLKYPCRYSN
LFPV*YGFLL CLLWTOVFVA CESKSAOKLL VIGFGSLFKL YPPESYR

The following DNA sequence nGPCR-Seq2001 <SEQ ID NO. 21> was identified in *H. sapiens*:

CCGGGCCAGTCTCACATCTGTAAGTGGGGAGATAAAACCTATCTAAGGGGGATATTA
TGAGGAGCAGGAGAGAAGGCATGTCACTCTCTAATTAACTGGCATTCACTAAAGAGTTA
TGTGATTATAAACATAATAAAAATATAAGGTGCTTAGTTATAATACAT
GAGGTCTGAGGGATGGTGATGACTGCACTTCAGTGGACTACCTTATGAGGGACTGACTGTC
AAGAAGCTCTGTGCATGTGCTTGTGTGCATCTGTGATTATATATACATATA
CGGTATATAAAATAGGCACTTTATTATATAAGAAAATGATGTGCACAGTAAAAATGCATAA
ATATTATGAGCATGTTGTATAAACAGAGTGAACAAAGAGACATTGGGATGCAAGACCA

GTGGTATAATCTGCCAAACACAAGCCCCGTTTACTGTTGCTAATGCACAAAGAGAGGG
CTGGCAGACATGGCCTGCATGCTCTGTACACCTGCCCATCTGCACACACTCCCCCATGG
TCTTGCTGTTGCTGTGGGCTCATCCGCCTCTCACA

The following amino acid sequence <SEQ ID NO. 81> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 21:

CERRDEPNTA TTARPWGSVV QMGRCTEHAG HVCQPSLCAL ATVKRGLCLG RLYHWSCIPN
VSLFTRLYKH AQ*YLCIFTV TSFSYIJKVP IYIPYMYIYN TQMHTHKHTC TELLDSQSLI
LCIFTV TSFSYIJKVP IYIP R*STEVQSSP SSDLMII*LE HLIFLYFIMY LIIT*LFSEC QLIRRMTCLL
SCSS*YPP*I GFISPLTRCG DWAG

The following DNA sequence nGPCR-Seq2002 <SEQ ID NO. 22> was identified in *H. sapiens*:

AGGCAGAGATTATGTGAGACTTGCATGATAGGGCTGTGCATTCTGCGCATTTCTTA
GCTCTGTGAGCAATATTAAGTAGTCATTACATAAAATCTATTGAACCTATACAAAGA
AGTGAACACACTGTGATGAGTGGCTCTACCAAGTAAGGCCTGGGGTTGGTCCCTTAT
GGTCCCCAGCACACATCTAAAATAGTCCTTGTGGATAAAACTCTCTTGAATTATIC
TATTTGAGTGTGCCATCTGATTCTTAGGACTCTGACTTAAAAGGTAGAGCATTG
CTCTCAT

The following amino acid sequence <SEQ ID NO. 82> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 22:

QRQLCETLHD RGLCILAHFL ALCEQYLSSH YIKSIEPIQR SEHTVMSWLL
PSKAWGLVLY GSPAHLK*F LCG*TLFELF YFECAI*FLL GL*L*KGRAL LS

The following DNA sequence nGPCR-Seq2003 <SEQ ID NO. 23> was identified in *H. sapiens*:

CAAATGGATTAATTACATATGGATGAACATGATCTGCTCTCCAGGTCCAAAGGAT
ACATAAGAAGAAAATTAGTGTGATGTTACTGGATGATGTCTTTAAGACAACACAATACA
ATATCTGAGTATGTACCCCTACGACATAGAGAAGGGATTTCAAAATATTAACTTAA
TAGATTCACTAAAAGAAATCACCTCCAACCCTGTTCTGTCTGGTCAAATTAGGGT
CATATAATTGTTTCATTGTATTACAAAAGGTAGAAATGTACACTGTATAATGAATAAA
TAATATAGATTACTAGATAAGCAGATAAAATAACAAAAGCACAAAAATACAAAAGCAA
ATGACCACTCAACTCACTTCCACCCACTCAAGGAAACACAGTTCTTATCATAGTTA
CTATTAGAAGTCTTCCTGCTTCATTCTACAAGCATGCAGAAATATATATGTGGAC
ACATTGTGCCAGAATTCTTTCTGACACTCACTTTCTCCTACTCCACAATA
TGTCAAGACAATTCTACAAGATATAGCAAATGGAGTAACATAGAATAGAGCAAACAT
GAAAACCTCAAACCTATTAGGGATGATGTTT

The following amino acid sequence <SEQ ID NO. 83> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 23:

KWINLHMDE HDLLLSRSQR IHKKKNLVML LDDVF*DNTI QYLSMYPYDI EKGFSKYFNL
NRFTKRNHLP TTVPLWSIR VILFLSYK R*ECTLYK*I NNIDY*ISR* INTKAQKYKS
K*PLNSLPPT SRKTQFLYHS YY*KSFLSSF SYKHAEIYMW THLCQNSFFS DTHFFPPTPQ
YVRTIFYKI* QME*HRIEQN MKTSNSLVDD V

The following DNA sequence nGPCR-Seq2004 <SEQ ID NO. 24> was identified in *H. sapiens*:

GGGAAGGCTCTCTAAAGAAACCACGCCACACACAAATTAGTAAATTGACAAAACAGGA
AACAAACAAAAAATTGTGTTAAAAGTAACACTAAATCAAATGAAAAGATAAAAGGCAA
ACACAAAATTGACAAGGATGCTGCAACTGTGTAACGTGATAAAGGGTATTTCCTGAA
TATATTCTCTATCTATAATTACTCTTAAATATATTAAAACCTCTGTAAGTAAG
TATGAAATAGACAAATGATAAGAACAGTTATCAGAAAATGAAATACAAATGATTCAAACA
TGAAAAGATGCTCGAGCTCAGTCTATAATAATAGACCCCTAAAAGTATAATGAAATATCT
TTTTTAAACCCCTATCTGATTAGCAAAGACCAAGTTGGAAAAACAGAGGGCTTTT
CAAAGATCAAGCTGTGAGCCAAACTAAAGGATCTGTGCTTCAAGAATTACACTGTT

AGAGTTGGACTTTGAAAAAAATGTACATGTCTATGAAATAATTGGCCTTTAAAAA
 AGAGTTGCAGCAATGCTGAAAGTAGTGGCCAAAGCATGATCAAGAGATAGGATATTCA
 TCTAACTCAAACCATCTCCCTACAAGCTATTATCAATTACAAGTGAAAATACTAAC
 TGCACAGTGGTGA

The following amino acid sequence <SEQ ID NO. 84> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 24:

SPLC S*YFSLCN** IACREMV*D* MNILSLDHAF GHYFQHCKL FFKRPIYFID MYIFFKKSKL
 *TV*FLKAQN PLVWLTSIF EKALCFSNLL VFANQIGV*K KIFHYTFKGL LL*TELEHLF
 MFESFVFHFL ITVLIICLFH TYLQKFLKYI KESNYRIEKY IQENSPLSVT QVASILVNFW
FAFYLFILI* CYF*HKIFCC FLCRUY*FV CGRGFLEEPS

The following DNA sequence nGPCR-Seq2005 <SEQ ID NO. 25> was identified in *H. sapiens*:

ACAAAAGTCACTCCTCTGCTCAAAATCTTGACGGTTCTGCTGTACTTGGAAATACAA
 CCCCAGTCCTTCTAGGGCACTAGCCTGTATTGGCCTTGATCACTGCTCCTGCCAC
 CTCATACCACTTGTCACTTGGCATTATAGTCCAACAAACATCCACTCATCCGCTTCTCG
 CAGGCCCTGGAATATTCTAACAGCACTTCCCCTCTCAGGGTGTGTAATGCTGTTCTC
 TACCTGCTCTCATCCCTCACTCTTATCAACTAACAGTCTCACATACACATGTTGATCT
 CAAGCTGTGTGTGTTACTTGTGTTATTGAAGCACTATGCTGAGCCATAGTGGAAATC
 TGAGGTAAAAGGAAAATAAGTAATATTGACTCTGAGTTATGTAATATTGATTGTTT
 CCCATGAAGGAATTGGCATTAAATTGTTATTAAAATATTGATTAAAATTGTTA
 TCTTGATTACTGAGTATTGGTGTCCCCCTAAATTCTCACCAAAAGGACAAGTGCCTCG
 CTCACCTCACCCCTTCCAGTACTGGTCATTGGTGTGCCCCACCAAACGTAAAGTAAAC

The following amino acid sequence <SEQ ID NO. 85> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 25:

QKSLLCSKSL HGFLLYLEYN PKSFPRALAL YWP*SLLLPT SYHLSFGHYS PTTSTHPLSR
 RPWNILSTFP SQGVVNAVSS TCSHPPTLYQ LSSHIIHMFDL KLCVFTCLLF EALC*AIVES
 EVKGKISNID SEFM*YFDFP P*RNFAFLIFI LKILY*NFIY LDY*VFWCPL KILTKGQVPR
 SPHPFSSTGH WSVPNNCK*

The following DNA sequence nGPCR-Seq2006 <SEQ ID NO. 26> was identified in *H. sapiens*:

CCGAGGGCTTCCCTGTGAGAATCAGTTCAAATATGAGGTGTATTATTTCGGT
 TGCAATTAACTGAAAAAAATTTCAAAGCTGTATAATTAAAGGCCGGCATTTATTAGG
 GTTTAAAATAGTCGCTATGAAATTGCTCTGCCATTGTAATTACAACGTACATCTTAA
 TATCCCCCTGCTTGCCAGCCCCGAGATAACAGCGCTGTGCTGCATTGGCTTGCA
 GAGAAGGTAAAATCCTTATCTTACTCCATCTGCACACAACACAGCCTCTTCCAAT
 TCAACCTTAATCTCTTAGTAATAACTGATATTTCACCAATAACAAAATGGAGCTG
 GTGCCAGATTCTCACTGATAACACCTCTCGTTACACACCATTCTGTCAAATCGGCC
 CGATTTTCTTAATCTCTTAACTTATCAAAAAAAATCTTACATTGGCCATAGTA
 AGTTCTTCTACCAAATAATAATTATAAAATCCTTGCCTACAGCTGACTCAT
 TGAAAAGATACTCAGCCTCCTCATGAAATGAAATTGTTCTGCTCTCAATT
 TTCCCTCCAAGTTGGGGTCCACACCAAGGAATTGTTCCAGGCAGGCCGTGAGCCAT
 CTTGTATCTGCAAGGGGTCACTTGGTCCAGGATGTGCATGCTTCTC

The following amino acid sequence <SEQ ID NO. 86> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 26:

EKH AHPGPR*PPA RYKDGS GPAW EQFPWCGPPN LGGKLKSRTI SIHEENEEAE YLFNESAVGK
 GIYL*YYYLV EKNLLWPNVK SIFFDKLKRL RKIGPILTEN GV*TRGVISE NLAPAPFCYGY
 EKYQYYLLRR LGLNWKRGWL LCADGDKIRI LPSLQSQMWH TQRCYLGAGK QGDIKMYVVI
 TNGRSKFHSD YFKP**NAGL *LYSFEIFFS VNCNRGK*NQ PHI*N*FSQG KPS

The following DNA sequence nGPCR-Seq2007 <SEQ ID NO. 27> was identified in *H. sapiens*:

AGTATAAAATCTGGGAGAAAAATCCATCAAATTATAAAATAGAACTAATAATGCTTTCAT

GGGTAAGCAATGAAACAAACCAAAAGGTAAACCGCGTTAAAGATTGATATTCTTAA
 GAAGAGAATGGAAAATAAATTAAATACATTATATTGATTATCATGAAGTTTCT
 AGTTCTACCTGAAGTTCTGGAGGAACCTAACAGCACTCATTGACATTAGGAAGA
 AGCCGAATGTAAATAAATCCTAGTGGATAAGCATAATATAATGAGTAATATGATCTAA
 ATGGAAAAAAGATTATGTAAAGTTTACTCTAACATTTCCACCCACTACACTGT
 GAAGCAGATTAACCTTTGTCAAAGGTGTAAACCTTCT

The following amino acid sequence <SEQ ID NO. 87> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 27:

EGY TFDKRLICFT V*WVENVksi NNFT*SFHl RSYYSLYYAY PPRIYLHSAS S*CQMSAVKV
 PPGNFR*ETR KTS**SI*NV YFNLSILFL KKYQSLKRWF TFGLFHCLPM KALLVLFYNL
MDFSPRFYT

The following DNA sequence nGPCR-Seq2008 <SEQ ID NO. 28> was identified in *H. sapiens*:

CTTTGAAGGATGAAGAGAAGTTACCAAGTGAACGTGGAGGAAAAGCAGTCCAGGCAGAA
 GGAACAGCTGCTGAAAAGGTCAAGGTGCCAAACCAGAAAGGCATGTGAGGGAAATTATGG
 GCAGTTCTGGAGAACCAAGCAGAGCAGAAGGAGACAGGCTAGAGAATGGCAGGGAGATAGCCCTA
 GAAAGCTCTATGTGTGCCAGGTGCAAAAGGCTTGTGCCAGCTAAGGAGTCTGCACCTT
 CCCCTGTAAGCAATTAAAGTTTAAGCAGAGGAGGGATTGGTTGTATTGATCTAGATA
 ATTCAAGGAAACAATGAAGGGAGGATATTGAGGAATCCAAAGAGCAGGGCAGAGACTC
 TCATCACAGTTCAAGGCCCTGACCGAAGGCCATAGGGAGATAGCAGTAAAGCAGGGAAACCT
 ACTGTCCCTTCTCACCTGGAAAACATATCACCTTCCAGAGTTCTGTCACCCCCAGTGAA
 GGCAACCACTGTCCTCTGTTACCTAACACGTGGTACAAACAGCTACCATAGCATG

The following amino acid sequence <SEQ ID NO. 88> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 28:

MLW*LF VPRVR*QKRQ WLPSLGVGT LEGDSFPGE GTVGFALL SPYGLRSGLN
 CDESLCPCSL GFLKYPPFTV P*I*INTNQ SLLCLLLIA YRGKCRLLSW RTSLHLAHI
 ELSRAISCHS LACLLLWFS RTAHEFPHMP FWFGLTFSA AVPSAWTAFP PRSLGELLFI LQ

The following DNA sequence nGPCR-Seq2009 <SEQ ID NO. 29> was identified in *H. sapiens*:

TATAAGGTAGTTGTAGCTTAAGACTAACGACCTCTACACCTGGAAATTGGTATACTTT
 TTCAATAAAGCATCAGATAGTAAAGTTTGTGGGCCATATGTCCTGCTGCAACTACTCA
 GCTCTGCCATTGTGCAAAAGCAGCCACAGATAATATGTAATGAATGAACCTGGCTGTT
 TCACAAAAACAGGCGGCTGTTAGGATTGGCTGCTGACTCTAACITGGATTCTCACTT
 CTGAAAGCATGGCTAGGGAGACCAGCAGCATCAGCATTACCTGTGAACCTGTTAGAGATA
 CAGAATGTTGAGCCCCCTTCATACTGATTGCTCTAGAGTCTAAACCATGTTCTCAAAC
 TTCAGCATACATAGGACTCACCTGGGGATCTGTGAAAGTACAGACTCTGAGATAGCAGG
 GCTGAACACCAACTAGCCTGGGGAGGCAATGTTATGACCTGGTCAGTGTCTGGGG
 AGTCCGCAATCTATTAGGAGAGACTGCTATATAACAAATAATTCAATACAGACTAGTA
 AATGCTATAACGGAGGTATCTCAGCAGAGGAGCACCA

The following amino acid sequence <SEQ ID NO. 89> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 29:

YKVVV*L*D* DLYTWKLVYF FNKASDSKVL WAICLCCNYS ALPLCKSSHR *YVNE*TWLF
 SQKQAAVRIW LADS*LGFSL LKAWSRRPAA SALPVNLLEI QNVEPLLILI ALESKPCFSN
 FSIHRTHLGI L*KYRL*DSR AEHHTSLGEA ML*PGQCSLG SPQSIRRDCY INK*FQYRLV
 NAITEVSSAE EH

The following DNA sequence nGPCR-Seq2010 <SEQ ID NO. 30> was identified in *H. sapiens*:

ATTTTCATGTTGATGGCAATCATCCAAGAGAGAGCAGGCCATGTCACGCAATAATAGAAC
 AAGGAGAAATGCACACTTCTTCTTCCATGTCTCTGCTGCATTACATTGCTAATACC
 ACITTAGCCAGATAAAATTAAACACAGCGCAGCACATAGTCAAAAAAATAGAACGACAGAAC

TTACAGAACAACTGACATGGTTAGAAAAGCAATTAGACCATCAGTGAAATCAGTCTAC
 CACAAAAGGTAAATTCACTTGTCAACAGCTTGAAAGACTTATCTTCCAAGACAGGA
 AATGGTTCTCCACTGAAGGGTGAAGACATTCAATTTCAGTCATTGGGGAGAGTTGG
 ATCTCCAACGAGTAACCTTCATGCAAGGACAAGAATTAGTAGTGAAATAGAGGTTATT
 GTTTTTTACCATAAAATAATTAAATCTTGAGGCAGTTCTCATAGCAGTTATT
 GGCAGITGTGTCATTACAGGAAAACGAGAAACTCTAAAGATGTTTGGGAAAAAAA
 GTATTGAAAGCTTGCAGTGTTAACCTCCACAATAGATATACTCTTC

The following amino acid sequence <SEQ ID NO. 90> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 30:

R VYLLWKLTLA SFQNTFFSQK HLRVSQFSCK *TQLP**LL* GNCLQDY*LF YGKKTNNLYF
 TTKFLSLHES YSLEIQLFPK *LKIEMSSPF SGEFPVLED KSFQQRCEQM NYLLW*TDFT
DGLNCSKPC QLFCNFWSI FLTMCCAVLI YLAKVVLANV MQAETWKKKS VHFSLFYYCV
 TWPALSWMIA INMK

The following DNA sequence nGPCR-Seq42 <SEQ ID NO. 31> was identified in *H. sapiens*:

CGCACAGCGC GCAGGTCCCT ACCAGAGCTC TGGTGGCCAC CTCTGTCCCG CCATGCTGCT
 CACCGACAGT GGCCAGGGCC CACAGCACCA AGAGGCTTGG GCCACAAAGT
 AAAGGGTCGC GGAGCCTCGC CGGGCGCCAT GTGGAGCTGC AGCTGGTTCA ACGGCACAGG
 GCTGGTGGAG GAGCTGCCCTG CCTGCCAGGA CCTGCAGCTG GGGCTGTAC TGTGTCGCT
 GCTGGGCTG GTGGTGGGCG TGCCAGTGGG CCTGTGCTAC AACGCCCTGC TGGTGTGCG
 CAACCTACAC AGCAAGGCCA GCATGACCAT GCCGGACGTG TACTTGTCA ACATGGCAGT
 GGCAGGCCCTG GTGCTCAGCG CCCTGGCCCC TGTGCACCTG CTGGGCCCG CGAGCTCCCG
 GTGGCGCTG TGGAGTGTGG GCGCGAAGT CCACGTGGCA CTGCAGATCC CCTTCATGT
 GTCCTCACTG GTGGCCATGT ACTCCACCGC CCTGCTGAGC CTCGACCACT ACATCGAGCG
 TGCACGCG CGGACCTACATGGCAGCGT GTACAACACG CGGCACGTGT GCGGCTTCGT
 GTGGGGTGGC GCGCTGCTGA CCAGCTTCTC CTCGCTGCTC TTCTACATCT GCAGCCATGT
 GTCCACCCGC GCGCTAGAGT GCGCCAAGAT GCAGAACGCA GAAGCTGCC ACGCCACGCT
 GGTGTTCATC GGCTACGTGG TGCCAGCACT GGCCACCCCTC TACCGCTGCTGG TGCTACTCTC
 CCGCGTCCGC AGGGAGGACA CGCCCCTGGA CGGGACACG GGCGGCGTGG AGCCCTCGGC
 ACACAGGCTG CTGGTGGCCA CGTGTGAC GCAGTTGGG CTCTGGACGC CACACTATCT
 GATCCTGCTG GGGCACACGG GCATCATCTC GCGAGGGAAAG CCCGTGGACG CACACTACCT
 GGGGCTACTG CACTTGTGA AGGATTCTC CAAACTCTG GCCTTCTCCA GCAGCTTGT
 GACACCACTT CTCTACCGCT ACATGAACCA GAGCTTCCCC AGCAAGCTCC AACGGCTGAT
 GAAAAAGCTG CCCTGCGGGG ACCGGCACTG CTCCCCGGAC CACATGGGGG TGCAAGCAGGT
 GCTGGCGTAG GCGGCCAGC CCTCTGGGG AGACGTGACT CTGGTGGACG CAGAGCACTT
 AGTTACCCCTG GACGCTCCCC ACATCTTCC AGAAGGGAGAC GAGCTGCTGG AAGAGAACGA
 GGAGGGGTGT TTTTCTGAA GTTTCCTTT TCCCACAAAT GCCACTCTG GGCCAAGGCT
 GTGGTCCCCG TGGCTGGCAT CTGGCTTGTAG TCTCCCCGAG GCCTGTGCGT CTCCCAAACA
 CGCAGCTCAA GGTCCACATC CGCAAAAGCC TCCTCGCCT CAGCCTCCTC AGCATTCACT
 TTGTCATGA AGTGTGAAA GCTTAGAGCC AGTATTATA CTTGTGGIT AAAATACTG
 ATTCCCCCTT GTTGTTTA CAAAAACAGA TGTTTCTAG AAAATGACA AATAGAAAA
 TGAACAAAAC CCTACGAAAG AATGGCAACA GCCAGGGTGG CGGGGCCCTG CCAGTGGCG
 GCGTGTGCTA GCAAGGCCCTG CGGGGTGTGC CGCAGTCACC ACAGGGTTCT GAGAACATT
 CACAGAAGTG CCTGAGACGC GGAGACATGG CTGGTGTAA ATGGAGCTAT TCAATAGCAG
 TGACCGCTC TCCTCAGCCA

The following amino acid sequence <SEQ ID NO. 91> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 31:

MWSC SWFNGTGLVE ELPACQDLQL GLSLLSLLGL VVGVPGVLCY NALLVLANLH
 SKASMTMPDV YFVNMAVAGL VLSALAPVHL LGPPSSRWAL WSVGGEVHVA LQIPFNVSSL
 VAMYSTALLS LDHYIERALP RTYMASVYNT RHVCGFWWGG ALLTSFSSLL FYICSHVSTR
 ALECAKMQNA EAADATLVI GYVVPALATL YALVLLSRVR REDPLDRDT GRLEPSAHL
 LVATVCTQFG LWTPHYLILL GHTGIISRGK PVDAHYLGLL HFVKDFSKLL AFSSSFVTPL
 LYRYMNQSFP SKLQRLMKKL PCGDRHCSPL HMGVQQVLA

The following DNA sequence nGPCR-Seq44 <SEQ ID NO. 32> was identified in *H. sapiens*:

CCACAAGACAGTGCCATTAGTGGCAGCCCTAGGATAAAGATGATACTGTAGGCCAGGGA
 GAGGTAGACTTGCTTGTACTTCTCTGAGAACTGGCAGAGACCTTGTCTGTGATGTATT
 CATGTCACCTCTCCATGTCCCGGGAGGCTCCCTCAGGAGCAGAGCTCCACGACGGCT
 CCCGCTTCTGCTTCCCTGGAAGGAAGCAAAATGGACAGCATAACATCATGACCTCAGCA
 TGGCATCAGCCTCTGCCCAACCCGGCATTCAACTTGTCCCCAAAGTCTAGTCAGAGG
 CCTTGTAAAAGAGACCAACCCAGGAAATAGAGTATTCAACAAAGGGCCAATAATCT
 TCCAAAGCTGGCGTCTAGACTTACCAAGGTCAAAGGGTGGCCTACATGGACTTGTAT
 GCTGTCTACTCTAGGCTGGGTGTGATAGAAATGGTGGGGATGACACTGTCAATTAAA
 CAGGGTACTCAGAACCTCCAGCTGTCTCCTGCTAGAATCCATTGCATAGAGTC
 AGAAAATCTCC

The following amino acid sequence <SEQ ID NO. 92> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 32:

TRLWGTKLNA GLGQRLMPC* GHDVMLSILL PSRGSRSGSR RGALLLEGAS
RDMEKVDMNT SSEQGLCQFS EKYKQVYLSL AYSIIFILGL PLNGTVLW

The following DNA sequence nGPCR-Seq45 <SEQ ID NO. 33> was identified in *H. sapiens*:

CATCTTAAGCTCGGGCACACAAAGTCCCAGAGGCACACAGATTCAAGTGGCTGGGGATAG
 CATATTCACACTCTGGGCCCCAACTCAAAATCCCCAACACCCCTCAGTCACCTGTGGC
 TCCAGTCCTGTTTGGTTTCCACAGATCACAAAGGCATCAAGGAAGAGGCTCACGGTAA
 GCCTGGCCTACTCGGAGAGCCACCAAGATCCCGGTGTCCAGCAGGACTTCCGGCTCTCC
 GCACCCCTTCCCTCTAATGGTCTCCTTCTCATCATGTGGAGCCCCATCATCATTACCA
 TCCTCTTAATCCTGATCCAGAACCTCAAGCAAGACCTGGTCACTGGCCGTCCCTTCT
 TCTGGTGGTGGCTCACATTGCTAATTGCCCTAAACCCATCCTACAAACATGACACT
 GT

The following amino acid sequence <SEQ ID NO. 93> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 33:

S*ARAHK VPEAHRFSGL GIAYSNSWAP NSKSPTTSH LWLQSCFGFP QITKASRKRL
TVSLAYSESH QIRVSQQDFR LFRTLFLMV SFIMWSPII ITILLILQFN FKQDLVWPS
LFFWVVGFTF ANSALNPILY NMTL

The following DNA sequence nGPCR-Seq46 <SEQ ID NO. 34> was identified in *H. sapiens*:

GCTGGGAAGTGCTCCTGGTCAGCCGGCGGTAGATGGCGGAGGGCACGGTGAGCAGCAAG
 GCCAGTGTCCAGGCTGCCCCACAGGCCACCTGCACCCCGCACGCCGCTGAACCGTATAC
 CACCAGGCAGGCCAGAGGCCAGGAAGCAGAGGTGGCACTGAGAGCTGCCAGGAGCAGG
 ACGCTGGCATACATGGTCAGCAGGATGATGGAGGGCAGCGCCGACAGCCCACTGCACCA
 TACGGTCAGTGGCCTCCACGGCAATGGCACTGCCAGGATGGCAGAGACAAACAGCAC
 AGCAAATCCGCCACGGCCAGGTGGAGCAACCAAGGTGGCACCCACCCCTCCGGCGGGCCACC
 TTCCCAAGCCACCCAGGCCACCATGGCATTGCCGGCACCCCCACCAAGGAAGATGGCGCA
 TACAGTGGAGCGGGGCCACGCGCAGCGGGTCATGGCCAGGCAGGCCATCCAGGCAG
 TCCACAGGGCGGTCCGAGAGGTGCTGTAATCCCCATACTCGTAGCTGACAGAATCG

The following amino acid sequence <SEQ ID NO. 94> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 34:

DSVSYE YGDYSDLSDR PVDCLDGACL AIDPLRVAPL PLYAAIFLVG VPGNAMVAWV
AGKVARRRVG ATWLLHLAVA DLLCCLSLPI LAVPIARGGH *PYGAVGCRALPSIILLTMY
ASVLLAALS ADLCFLALGPAAWWYTVQRAC GVQVACGAAW TLALLTVPS AIYRRLHQEH FP

The following DNA sequence nGPCR-Seq47 <SEQ ID NO. 35> was identified in *H. sapiens*:

GGCTCCACTGGCCTTCGGCAAGGGGCTCTGCAGGCTGACGGCGTTGTGCTCACACCG
 ACACCTACGGGGGTCTACCTCATGGCCTGTGAGCGTGGACCATTACCCAGCTGTGGT
 CTGTGCCACTGGGCCGTGCCCTCGCACGGCTGGCCGCCAGGCTGGTCTGCGTGGC

CATCTGGACCTGGTCTGCAGACGATGCCCTGCTCTGATGCCATGACCAAGCC
 GCTGGTGGCAAGCTGGCCTGCATGGAGTACAGCAGCATGGAGTCAGTCCTCGGGCTGC
 CCCTCATGGCCTGGCTTGCATTGGCTCTGTGGCCAGTGGGATCATCCTGT
 CCTGCTATATGAAGATCACCTGGAAGCTGTGCAGCACAGCTGGAGAACCCAGTGACCAAG
 CGGGAAAGGACACCACCGCGGGCAGCCGGAGGACCCAGTGACCAAGCAGGAAAGGAC
 GCCACCGCGGGCAGCCAGGACCCAGTGACCAAGCAGGAAAGGACACCCACCG

The following amino acid sequence <SEQ ID NO. 95> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 35:

AP LAFRQGALQA DGVCALHRHLR GVYLMACVS VDHYPAVVCA HWGPCLRTAG
 RARLVCVAIW TLVLLQTMPL LLMPMTKPLV GKLACMEYSS MESVLAAPH GPGGLCHWLL
WASGDHPVLLYEDHLEAVQH SWENPVTSGK GHHRRGSPGG PSDQQERTPP AGQPRRTQ*P
 AGKDTT

The following DNA sequence nGPCR-Seq48 <SEQ ID NO. 36> was identified in *H. sapiens*:

GAGGCAGGCT GCAGGGAAAGT AAGGAGGGAGG CATGGCACCT TCTCATCGGG CATCACAGGT
 GGGGTTTTCGC CCCACCCCTG AACGCCCTCT GTGGCGCCCT CCACCCACCT GTAGGCCAG
 AAGGATGTCG GTCTGCTACC GTCCCCCAGG GAACGAGACA CTGCTGAGCT GGAAGACITC
 GCGGCCACA GGCACAGCCT TCCTGCTGCT GGCGCGCCTG CTGGGGCTGC CTGGCAACGG
 CTTCGTTGTG TGGAGCTTGG CGGGCTGGCG GCCTGCACGG GGGCGACCGC TGGCGGCCAC
 GCTTGTGCTG CACCTGGCGC TGGCCGACGG CGCGGTGCTG CTGCTCACGC CGCTTTGT
 GGCCTCCTG ACCCGGCAGG CCTGCCGCT GGGCCAGGCG GGCTGCAAGG CGGTGTACTA
 CGTGTGCGCG CTCAGCATGT ACGCAGCGT GCTGCTCACCG GGCCTGCTCA GCCTGCAAGCG
 CTGCCTCGCA GTCACCCGCC CCTTCCTGGC GCCTGGCTG CGCAGCCCGG CCCTGGCCCG
 CCGCCTGCTG CTGGCGGTCT GGCTGGCCGC CCTGTTGCTC GCCGTCCCGG CCGCCGTCTA
 CCGCCACCTG TGGAGGGACC GCGTATGCCA GCTGTGCCAC CGCTGCCGG TCCACGCCGC
 CGCCCCACCTG AGCCTGGAGA CTCTGACCGC TTCTGTGCTT CCTTTCGGGC TGATGCTCGG
 CTGCTACAGC GTGACGCTGG CACGGCTGCG GGGCGCCCG TGCGGCTCCG GGCGCACGG
 GGCGCGGGTG GGCGCGCTGG TGAGCGCCAT CGTGTGCTGCC TTCTGGCTTGC TCTGGGGCCC
 CTACCAACGCA GTCAACCTTC TGCAAGCGGT CGCAGCGCTG GCTCCACCGG AAGGGGCCTT
 GGCAGAGCTG GGCGGAGCCG GCCAGGCGC GCGAGCGGGGA ACTACGGCCT TGGCCCTTCTT
 CAGTTCTAGC GTCAACCCCG TGCTCTACGT CTTCACCGCT GGAGATCTGC TGCCCCGGGC
 AGGTCCCCGT TTCTTCACGC GGCTCTTCGA AGGCTCTGGG GAGGCCGAG GGGCGGCCG
 CTCTAGGAA GGGACCATGG AGCTCCGAAC TACCCCTCAG CTGAAAGTGG TGGGGCAGGG
 CGCGCGCAAT GGAGACCCGG GGGGTGGGAT GGAGAAGGAC GGTCCGGAAAT GGACCTTTC
ACAGCAGACC CTACAACCTG CTGCCCTTCC CTGCTCCCTT CCACCCCCCACCCACCTCC
AGAGGTCAGT GTTCTGGGAC ATTTGGGAC CCTCTTTGA CTAGAGTTG GATCTGGCTG
GGTAGGATTA CTATACTT GGGCAGGCC CAGGCTCCTC CAAACTGAGG GATTATGAGG
GTGGTGTGATGG TCCCTGTAA GGACTATTGT GTGCTITGCAA GTTGGCATGT ACCCATGTGC
CAGCATTGCT

The following amino acid sequence <SEQ ID NO. 96> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 36:

MS VCYRPPGNET LLSWKTTSRAT GTAFLLLAAL LGLPGNGFVV WSLAGWRPAR GRPLAATLVL
HLALADGAVL LLTPLFVAFL TRQAWPLGQA GCKAVYYVCA LSMYASVLLT GLLSLQRCLA
VTRPFLAPRL RSPALARRLL LAVWLAALLL AVPAAVYRHL WRDRVCQLCH PSPVHAAAH
SLELTAFVLF PGFLMLGCVS VTLARLRGAR WGSGRHGRARV GRLVSAIVLA FGLLWAPYHA
VNLLQAVAAL APPEGALAKL GGAGQAAAG TTALAFFSS VNPVLYVFTA GDLLPRAGPR
FLTRLFEGSG EARGGGRSRE GTMELRTTPQ LKVVGQGRNGDPGGGMEKD GPEWDL

The following DNA sequence nGPCR-Seq49 <SEQ ID NO. 37> was identified in *H. sapiens*:

CCAGCCGTCCAGGCGACGCCAGCAGCACAGGAACCAAGGTGACGCTGCCACGTGGTAG
 CGCGGGCAGACCAAGGGGCACGCCGTGTCGCAGCAGGCCAGGCCAGGGCTGGCGGGCAGC
 GGCACGGCCGCAGGGTCGGGTCACACCAGGGCGCAGAGGACGCCAGGCCAGGCCTGCTG
 GGCCGGCAGCCCTGGTAGCAGGCCAGGGAAAGAGGTCGGAGAGGCAGCGCTCACGCTGAAG
 GCCGCCAGCAGCCAGAGCCCCACCGCGAACCACAGGAAGGTGAGCAGCAAGTAGAGTGTG

TCCTGGCGCCAGGGCAGCCTGAGCCACGGAGAAGCCCACACGGCAGGAGAGGAACAGG
AAGTCGGCGGCCAGGTGCAGCAGGTAGATGGAGAAGGGGCCCTCTGATGCGGAAG
CCGAGGTTCCAGAGCACCAGCCGTTACCTACCGTCCCCGAGGCCACGATCAGCGTC
AGGTAGAAGACCAACTGTCGAAGGTTCTCCAGAGGCCAACAGCCAAACATCCTGGCC
GGCTCAGAGGGTGCTGGCGAGGGACCTGCAAGATGGAGACAGACATGGTCTCAGGAGTC
TTTGTGCCCGCTGGGGCACAGGGAGTGGCAT

The following amino acid sequence <SEQ ID NO. 97> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 37:

MPLPVPP AGAQKTPEDH VCLHLAGPSP APSEPARMFG LFGLWRTFDS VVFYLTLLVG
LGGPVGNGLV L WNLGFRIKK GPFSIYLLHL AAADFLFLSC RVGFSVAQAA LGAQDTLYFV
LTFLWFAVGL WLLAAFSV ER CLSDLFPACY QGCRPRHASA VLCALVWTPT LPAVPLPA NA
CGLLRNSACP LVCPRYHVAS VTWFLVLARV AWTA

The following DNA sequence nGPCR-Seq50 <SEQ ID NO. 38> was identified in *H. sapiens*:

ATGTCTACAGAAACCCCTCGCCATCTACCTCTGGTACGTGGCCTGCAGCAGGATCTCA
TCITCCCTGGCTGCCACATGGTGGCCATCGTCCCCGACTGCTGCAAGGCCGGCTGGACT
TCCCGGGCTTCGTGCAGACCAGCCTGGCAACCGCTGCGCTTCTCTGCTACATCGTGGGCC
TGAGTCTCCTGGCGGCCGTCAAGCGTGGAGCAGTGCCTGGCCCTCTTCCCAGCCTGGT
ACTCGTGCAGCCGCCACCTGACCACTGTGTGCGCCCTCACCTGGGCCCTT
GCCTGCTGCTGACCTGCTGCTCAGCAGCGCTGCACCCAGTTCTCGGGGAGGCCAGCC
GCCACTTGTGCCGGACGCTGTGGCTGGCAGCGGTGCTGCTGGCTCTGCTGTGCA
CCATGTGTGGGCCAGCCTTATGCTGCTGCTGCCGGTGGAGCGAGGCCCCAGCGGCC
CACCCGGGGCTCCCTGGCTCATCCTCCTCACCGTCCCTTCCTGCGGC
CTGCCCTCGGCATCTACTGGCTG

The following amino acid sequence <SEQ ID NO. 98> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 38:

VYRNP FAIYLLVRLG QQDLIFLGCH MVAIVPDLLQ GRLDFPGFVQ TSLATLRFFC
YIVGLSLLAA VSVEQCLAA FPAWYSCRRP RHLLTCVCAL TWALCLLHL LLSSACTQFF
GEPSRHLCRT LWLVAAVLLA LLCCTMCGAS LM**LLRVERG PQRPPRGFP GLILPHRPL**
PLLRPALRHL LA

The following DNA sequence nGPCR-Seq51 <SEQ ID NO. 39> was identified in *H. sapiens*:

AAGGTTCTAGGAACACATTGCCCTATAATTCTATGGTTGTTATACCTGCTATCTGA
ATGTTCCCTCTGCAGCTGCAATCCCAGTGTACCAAAACAGAGAAGTGTGAAGTTGACAA
AGATGGTGCTGGTGTGGTAGTCTTATCCTGAGTGTGCTGCCCTTATCATGTGATAC
AACTGGTGAACCTACAGATGGAACAGCCACACTGCCCTCTATGTGGTTATTACCTCT
CCATCTGCTCAGCTATGCCAGCAGCATTAAACCTTCTACATCCTGCTGAGTG
GAAATTCCAGAAACGTCTGCCCTAAATCCAAAGAAGAGCAGTGAAGGAAATCAACA
ATATGGGAAACACTCTGAAATCACACTTTAGGAAAGTACATGGATCACCAGTGTCTAG
ACATGATTGTCTATCTTACTGGTATTATTAGAAAGGGCAGGTGTACCGATATGTATGC
CCATTCTCTGTGTACTTGTGACTCTAGCAGCATGGAAGAGAAGTGTAAACCATGCAAA
TACAATGAGCTTAATATGTAACCTTAGCAAGATGTAAATGTGTATCTATATTGIGGGT
AGGGAATGGGATAGTCTGAGATACCCAGGCTCATGATGGTGTATATTATTCAGCATAT
TATAAACTAGTCACTAATG

The following amino acid sequence <SEQ ID NO. 99> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 39:

GSRNTLPH NFYGC~~L~~YPVY LNVSSAAAIP VYQNREVMKL TKMVLVLVVV FILSAAPYHV
IQLVNLQMEQ PTLA~~F~~YV~~G~~YY LSICLSYASS SINPFLYILL SGNFQKRLPQ IQRRATEKEI
NNMGN~~T~~LKSH F*ESTWITMS LDMIVYLTGI IRKGRC~~T~~DMF MPILLVYL*L LAAWKR~~S~~VTM
QIQ*A*YANF SKM*NVDLYC G*GMG*SEIP RLHDGVYYFS IL*TSH*

The following DNA sequence nGPCR-Seq52 <SEQ ID NO. 40> was identified in *H. sapiens*:

CAATTCTATTGCCCTCTGGCTGTGACTCTTGGTAGGTGTGACTGTGATGCTT
 TTCAGCATGGTCAGGACGGTGGAGAGCTGCTGGTATTGGAGCCAAATTTGACTCTT
 CACAGTTGCTGTGATGTGGCATTTGTTACTCTTCTGCTCCACTGTGCTTCATCTGC
 ATCGACAGGTACATTGTGGTTACTGATCCCCGGTCTATGCTACCAAGTTACCGGTGTCT
 GTGTCGGAATTGCAATCAGCGTCTGGATTCAGCCCTCACGTACAGCGGTGTGTT
 TTCTACACAGGTGTCAATGATGATGGGCTGGAGGAATTAGTAAGTGCTCTCAACTGCGTA
 GGTGGCTGTCAAATTATTGTAAGTCAAGGCTGGGTGTGATAGATTCTGTTATTCTTC
 ATACCTACCCCTGTTATGATAATTCTTACAGTAAGATTCTTATAGCTAACAAACAA
 GCTATAAAATTGAAACTACTAGTAGCAAAGTAGAATCATCCTCAGAGAGTTAAAATCAG
 AG

The following amino acid sequence <SEQ ID NO. 100> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 40:

QFSIASLACA DFLVGVTVML FSMVRTVES WYFGAKFCTL HSCCDVAFCY SSVLHLCFIC
 IDRYIVVTDP LVYATKFTVS VSGICISVSW ILPLTY SGAV FYTGVNDDGL EELVSALNCV
 GGCQII VSQG WVLIDFLLFF IPTLVMILY SKIFLIAKQQ AIKIETTSSK VESSSESYKIR

The following DNA sequence nGPCR-Seq2021 <SEQ ID NO. 41> was identified in *H. sapiens*:

ATTATTCACTCTTAAAGACATTAAACATTGAGCACCTGCTGTATGCAAAGCACTGG
 GCACCCACACTAAAGGATGAAAACCAAGGATAAGTAAGACAGCATGTAGATTCTAGCTGCC
 TGGTTGAGAGGTACAGTCAGAAAGTCTTATAGCCATTCACTGACACATAAGATAGGGAT
 AGGAGAGGAAATGGGGTGGACACAGGGAGGGATAAGTATAGGGTCAGGGCTACCAACCT
 TCTTTCCCCATGACCCATGGGGCAAATATGCTCTCTCCGGTGGGTGATGT
 TCACTGCCCCCTTCTCCATTCAATTGAACTCTAGATTGAGCCCGAAGCTAGACTTGC
 AGATCACAAATTAAAGAAAGTGGATGTTCTGCAACAGGTACACAGGAGCTATATTGA
 GTCATCTTCTGTTCAACCAATCATTATTCACTTCAACAAAGTATTCCCAAG
 CACCAATTCCATCCTCAACAAAAGACTCTAAATCACAGGCCAGATGAAAGACGTCACTA
 TTCACTGGAAAAATTCACTGATTATGATCCTGGTCGGAGGTACTCAGCGTCC

The following amino acid sequence <SEQ ID NO. 101> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 41:

YSFL *TLNIH*APA VCKALGHTK DENQG*VRQH VDSSCLVERL QSESPYSHSV
 TYR*G*ERKW GEHREGISIG SGLPPFFSP* PHGGKYVLSL PGWVMFTAPF
SPIGTSRLS PKLDLQITIL RKLDVLQVQV QELSLSHLFV HPIIYSFIHS
 TSISQAPIPS STKDS*ITGQ MKDVTIHEK FS*L*SWSEC TQR

The following DNA sequence nGPCR-Seq2022 <SEQ ID NO. 42> was identified in *H. sapiens*:

ATGGAATTAGAAAAATGAGAAAGTAATCAAAGATTGAAAAAAATCTGAGATAGAATTAA
 AAGGAATAAAAATTGAAATCAAATATGTAATTCTTTAAAGTTGACACATTGTAATT
 TTATATAATTGTTGGAGTATGATTGGTACATTAATTGTTATATGTTGATGCTGATCA
 AATCAGGGTATGACTAGGCCATTGGTATGGTATAAGAAATACCTGAAGCTGGTA
 ATTAAAGAAAAGAAGTTATTGACTCATGGTCTATAGGCTGTGAACACATCTGCTT
 GGCTTCTGGTGGAGCCTCGGAAAGCTTCAATAGTGGCAGAAGGGGAAGGGGAAGCTGG
 AGTATCACATGGTGTGAAAGGGAGCAAGAGAGAGAGAGAGACAGGAGGTCTAGACTT
 TTAAACAAACAGATCTGTGTGAACTATTCACTGACTAAGAACTCATCACCAAG
 CGGAAGGTGCCAAGTCATTGAGGGATCCACCTCATATAACACTACCTTCCACCAAG
 TCTCACTACACATTGGGAATGACATTCACTGATAAGATTGGAGAGGACAAATGTCCA
 AACTATATCAGGGTAGTTAGCTATTCACTCACCTCATGTCTTATCATTTC

The following amino acid sequence <SEQ ID NO. 102> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 42:

NDKDMR** MS*LP*YSLD ICPLQILC*N VIPNVVSETW WKVVYYEGGS LMNDLPSAW
 **MSS*SHEI VHTRSGLKV *DLLSLSL AFHTM*YSS FPFPFCHY*K LSEAPTRSQA

DVVHSL*NHE SNKLLFFKLP SFRYFFIPMQ KWPSTYPDLI SIQH_{TY}KLMY
QIILH_{KY}IKL QC_{VNL}KRITY LIYK_{FF}IPFN SISV_{FFT}IFD YFLIFLIP

The following DNA sequence nGPCR-Seq72 <SEQ ID NO. 43> was identified in *H. sapiens*:

ATGTAAGGGCCTTGGAAAGAAA^{CTGTAA}CTCCACTGTGGGCCATTAAAGAAGTCT
TGAATACATAGATTAAATCCCTGGTTGAGATAATAAA^{CTCA}ATATTACCATGCT
TTTGTCA^{TTAA}ATACAAATTGTGGACATTTCACCAGGGCAACAGATGAGTTACAACCA
TCC^{TTT}TATCAGTTGCACAGAATCTTATTGAATGGGTATATCCTACTTATGTCA^{GT}TT
AGTAGTGAATATTAGGTGTTCTAATTTCATATTAGAAACAATGTTAAAAGAACTT
GCTCTATTGTGTTCTATGTCTACACTAGTATTCAATACCTTCTAGGAGTGGAAAT
TCCTGGCTAAACAAACTGCCTCCAAGAAGATTACTAATATACATTCTACTAATGCT
GTTT^{AGA}ATGCCTTCTCATATTCTATGAACACTGAGTGTCTCTTTAACACT
TTTTCTCAATCTACTAGACTCCTCTAAATATACTTTCAAAACGCCTGCTTAC
CCTCAATATATTGTCTTAAAAGATTATTGTAGGTACTGTTATGTGTTAGGTTGGT
GGAAAATTAAATTGTGATTTCGCAAAACTGCAATTACTTTGCACCAACCTAATATTACAT
GTAGA

The following amino acid sequence <SEQ ID NO. 103> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 43:

M*RAFWKKTV NSTVGPLKKS *IHRLNPLV* DNKTLNIYHA FVI*IQIVDI FTRATDELQP
SFYQLHRILL NGYILLMSVL SSEY*VVSNF SILETMLKEL ALICVSYVYT SIQYLSRSGN
SWLKQTAFQE DLLIYIPTNA VLECLFLHIP MNTECSLFNT FFPQSTRLLL KYILFKTPAY
PSIYCLKDYL*VLFMC*VG GKLIVLQKL QLLLHQPNII HV

The following DNA sequence nGPCR-Seq2024 <SEQ ID NO. 44> was identified in *H. sapiens*:

TCTCTTTGGTCCCTTTCTCATTACCTACCTCTTCTCCTTGTGGTCCTTCACA
TCTTCTGGCCTCTCTTGTCTGCATTACTTGTGGTCTTCGTCCTTAACACATCT
ACCCATCAGCTACTCCAAATGTATATATCTATTCCCGGACCTTCTCTGAACCTCAGA
TTGTATTCCAAC^{ACTAC}CTCAATTAGTACCTTCTGGATATTATTAAAATTGAATC
ACGACATGCCCTAAATTGAACTTCTATCTCTGAAGCAAAGCCCTATCCTCTATTGTT
TCCCCTCCAAATAATTGCCACTCATTCTCCAGATGCTTGGCAACATTTCAGTC
ATCTTGAATTCTCTTGTATCCCACATACATTCAACAGCATATGCTGCCCTG
TCTACATTAAACATAACCAGCATCCAACATTCTACAATTGCTACTAGCCTGTATAG
CCCTATCCTCTCCTCTGGATTACTGTATTAGCCTCTGAAC^{ACTGG}ATCCGCTTCAACC
CTTCCAACCTCCTCTCCAAAGCCA^{ACTCTG}GAACAGCAGCTAAAGTTGTTCTTCA
AAAA

The following amino acid sequence <SEQ ID NO. 104> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 44:

LFWFPPFLI TYLFSFAGPS HLPGLFFVCI YLCGLPSLTH LPISYSQMYI SIPGPFL*TP
DLYFQLPTQ* YLLGYLLKFE SRHA*N*TSY L*SKALSFYC FSHPNKLPLI LPDAWATFLQ
SSLISFL*Y PTYIHQHMLP CLHFKNHQHP TFLQLLLALY SPILSPLDYC ISL*TGIPLQ
PFPTSSSQSQ LCEQQQLKLFQ QK

The following DNA sequence nGPCR-Seq2025 <SEQ ID NO. 45> was identified in *H. sapiens*:

CAACCCTGGTAAAGCATCGCACCTGGTTATGTACCTCAGGGTTATTGACGC^{ACTGG}
CTAAAATGTTGAAGGACATCCTGTTCCAGGTGGGACTGGAACAGAGCCTGGACTGTT
TAGCCAATGGCTCCTTACCTCAGGATGTTGCATTCCAGCACATTCTGGTGTGAG
AACTACAAACAAGAAAGTGGGAAGA^{ACTG}TCTGCACCATTATGTAAC^{CT}CTAGGAAA
GCAA^{ACTAATG}TATTGTGACAGAAGGAAGATGAGTGATTGCTTAAACACAAGGGAGGAAT
GCTACACAGGGAAAGGTCTGGAGAGTTGGATTACAAAGAGAGACAAGGACACTTTGGGAG
AGATGGATATGTTATTATCTTGCTTATGGTGA^{AGT}TTCATAGGCCATAATCCAAA
AGCATCTCATTGTACACTATGTATGATTATTATATTAAATTACCTCAATAAAG
TTGTTTTAAAAAGTACCACTTAATCCTGAAATAGACCAGGAAGGCAATTAAATTAA

TATTTCTAGTTACTTTGAGAAACTAAAGCTTAGTA

The following amino acid sequence <SEQ ID NO. 105> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 45:

NPGKASHL GLCTSGLFDA LG*NVEGHPV SRWGLEQSLD CFSQWLLTSG CCIPSTFWLV
LRTTNKKVGR TVLHHLCKL GKQTNVL*QK EDE*LLKHKG GMLHREGLES WITKRDKDTF
GRDGYVYYLA YGDSFIGP*I PKASHCLTM YDLFILIPQ *SCFLKKLPL NPVNRPGRQL
INIF*FTFEK LKLS

The following DNA sequence nGPCR-Seq2026 <SEQ ID NO. 46> was identified in *H. sapiens*:

AGTTTATGGACCAGCCTCCCTGTGAAATTGACTTTCCCTCTTGCTGAATTGGTCAG
GTTAACAAATGGTTACCCCTGGATTACAGGAAGGGCATGTGCTAAAAGCCTTTGGAGAC
CCACATGGCCCTCAGATGAGCAATTGTCAGATTCCCTTTCTTTCCATGGGAA
TAAGCTTCCCTCTCCAAGTACATGTTAGGCTTTTATTTCTTGCTACTCCAA
GGACCTGGTGATATTTCTTACCATGCATTAACAGAAACTGTGAGTCTTCTGGAA
AAAAAAAAGGCAGGAGGAAACATACTAGTTAAAAGTTCTGGGTACACTACCAAGATGT
ACCTATTTATGATATACAAAATGGCATAAGTATTGAATGCTGCTATAGGCAATTCTCA
AGAACTTGTAAGAATTGACTTACATGAGCTACTCATAGCAGTTGATGATATAATGT
TGTTATTATCACCACTTACAGATAAGGAAATAGAGACAGACATAACTGAATGACATGCTC
AACGCCACTCCACTAGCAAGTGGCAGAACCCAAGC

The following amino acid sequence <SEQ ID NO. 106> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 46:

LGFCHL LVEWR*ACHS VCLSLFPYL* SGDNNNMYII ELL*SSSCKS ILTKFLENAY SKHSITYAIC
ISINRYILVV YPETF*LVCS LLPFFFPEKT HRFCLMHGKE KYHQVLGSSK KIKKPKTCTL
ERGKLPMEK KKRNLNNCS SEGHVGLQRG F*HMPFL*SR GNHC*PDQFS KEGKVKFIRE
GWSIN

The following DNA sequence nGPCR-Seq2027 <SEQ ID NO. 47> was identified in *H. sapiens*:

CGTGGCAGGTTGCACCAACTTTAAACCCACAAGCAATTATGAGAGGTTCTTCCCTTC
ACTCACAAAAAACTGCATCTTACGTTTAAGAAATCTTTGCTGCATTTGCTGGGATT
AACTTGTCTCTCATTTCTATATTGCATTCTAAAGCTTTAAGCTTAAGCTGAATATTTCCA
TATGGGCCATTGTATTGCAAAACACTACCCCTCAATGATTGCTGGCTTCT
GCCTTCACTAGTATTATTGCTCTTAAACACTAAAGCTTAATAGTTTATTGTTCTT
TGTCAGCCTCTCTGAAATGATGACACTTTCAACAGTACCACTTCTACTCATT
TCAGTTCAATGCTCATATAGGGCATGTGCACTGACTGTCCAGCTGTGCACTAAGCA
GGTGAATCTCCCTGCCCTGGATCGAGTAAAACAGAACTTAAAGGCTCTGGTA
GGAAAATCATGGGTACTGGTAACAGAGAGATGGATTCAAACTGGCTTCTACTTA
CTCACTCAA

The following amino acid sequence <SEQ ID NO. 107> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 47:

VAGCTNF *THKQL*EVH FPSLTTKLHL ITF*EIFAIL LGLTCLSLFY ICISKLLSLN NFHMGHLYLQ
NKHYPFDFA WLLPSLVFIA SLKH*VNSFI CSFVSLL*K* *HFSNSTTSF YSFQFNAHIG
HCAY*LSKLC TKQVNLPCPW IE**QNLKKA PG*ENHGYW* QRDMDSNPGF STYSL

The following DNA sequence nGPCR-Seq2028 <SEQ ID NO. 48> was identified in *H. sapiens*:

ACACCAGGAACACACTTGGATGCCATCGGCCCCAGGTTCTCTGCTGGCCTGTCTGG
CCATGTGTTGGTTGAAAGTGTCCAGATTGCTGGTCTCCTGGATCTCACGCAA
GAAACGAGCACATGATGGTGACCTGAACTCTCCTCGCTGGCGTACTGCATTTTGCAA
AGCCTGTAACTGTCTGCTGCATTCTGGCTCCAAGGCTCAGTCTCTGATTTTCTG
ACCTGCTCTCCCTAGGCTGCCTTCTTTTCTTATACAGTAAAGTCTGTATGT
ACTGCAGCATATTCCATTATTGGGAATTGAATATATTCTATGCCAGGATTAAAA

TTAGGATTGTTAATATTGTGGTTGTGCTCTGGTTACAGAATTCTAAGGTTGGGTGTT
 CTATCCCAGCACCATACTTCTAAAGCTTGTCTGTGAGTGGGTGATTTGCACAAT
 GCAAACITCTCAGGTACTTCTGAGCTCCAGGTCCCCATATCCAACATCTACTCAAC
 ATCTTATTCTGAGATGTTACCTCAAACATGTGAGAATCAACATGTCAACATGTCAATAC
 CTCTGCAGGATTCCTAACTCCACAGATGGTATCCGTCCAATTGTGCAAGCCAAGAGCA
 AGGCGCCAGCTGGGACGTTCCCATAACCAATCTATCATGTCCCATTAAATTAC
 AGTAGT

The following amino acid sequence <SEQ ID NO. 108> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 48:

TRNTLGC PSAPGFFCWS CLAMCLGLKV SRLPGSPGSS RKRNEHMMVT WNSPRWRHCl
 FAKPVTVLSA FWAPRLSPLI FPDLSPF*AA FLFFLITVKF CMYCSIFHLL GIEYISSMPG
 FKIRIVNIVV CALVTEFLRF GCSIPAPYFL KALLSAVGDF AQCKLLRYFL LSSRSPYPTS
 TQHLILRCSP QTCEHQVNM SIPLAGFPNS TDGIRPIVQA KSKAPAGTFP IPNLSSCPIK FYQ*

The following DNA sequence nGPCR-Seq2029 <SEQ ID NO. 49> was identified in *H. sapiens*:

CACTTGAGAGGTGATCTCATCTATAAGAGAGTATTTAAGGAGCTAATTACACAGGT
 AGATGACAAAACCTTACACCAACTCTCAAGCACAGGAAGCACCTTACACGAAACACTTT
 GGTGAATGCTCATTTACTCAATTAAACTAAATTCAACTGAAAATACCTCATTTAA
 AGAGGATATTCAAGTTGACTCACTGGGGAAAAACAATTGGATAAACTGGTTGTTGCCCTT
 ACCGAGATAAAATCACTGATAAGTGAATGCATTGTAATGTGTCAGACAGATGGCT
 TTAATAGTCACACATATTAAGTATGGGATATGTGGCGAGGAATAGAGGTTTATATACA
 GAAGTAAGAAGTGTAGAGGGGGAAACCATGAAAGATAGAAAAAGATGAGAAGAAAGAGG
 ACAGAGATGATACACTCAAGAAAAGCTGCCTATTCAAGAGTAAATATTGTAAGCCCC
 AGTATAGTTGAATGCTTACAAGAGGAGAAGCTTTAGCAACTCCTTGTGTTCTTAATGTA
 TATATTATATCTGAACAGAATGCTTCTTGTGAGCTAGACAGTAGGAAGTAATCTAT
 CTTCAACCTGTTGGTGTAAATAAACTTT

The following amino acid sequence <SEQ ID NO. 109> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 49:

KFYLHTKQ G*R*ITSYCL AHRKEAFCS D IYTLRNKG V AKSFSSCKHS T ILGLTIYST
 LK*AAFLECI ISVLFLLIFF YLSWFPPSTV LTSVYKNL YS SPHIVYLICV T IKAICLDTL
 QKCIQLISDF ISVRANNQFI QLFFPSESTE YPL*NEVFSV EFSLKLSRNE HSPKCFV*GA
 SCA*RVGVRF CHLPV*ISSL NILSL*MRSP LK

The following DNA sequence nGPCR-Seq2030 <SEQ ID NO. 50> was identified in *H. sapiens*:

GTGATACGGTTGGCTGTGTCCTTACCAAATCTAATCTGAAATTACAGCTCCATAATT
 TCCATGTGTTGTGGAAGGGGCCTGGTGGAGATAACCGAATCATGGGGCGGCTCCCC
 ATACTGTTCTCTGGTAGTGAATAAGTCTCATAAGATCTGATCGTTTATAAGGGAAAC
 CCCTTTGCTGGTCTTCTCTGCTGGCTGCCATGTAAGATGCTTCTTGTCTC
 TTGACTTGTGTTCTGCCATGACTGTGGGCCTCCAGCCATGTGAAATGTGAGGAAAT
 TAAACCTCTTCCCTTATAAAATTGCCAGTCTGGTATGCTTTATCAGCAGTGTGAAA
 TGGACTAATACACCAGAACAGTTCAAGACTAGCTGGCAACACAGTAAGACCCGTCTGTA
 TAAAGAAAGAATAAGAAAAAGAAAAATATCAGTGGCATAATATTAGACAGAATACTATAA
 AACTGATTAAGGCACATTAATATAGCCAGAATAATTATCTGATTATATCATTGAAAG
 ACCAGATAATCAATATAACTGATTCTATCCCGAGTAGCTCAGACTTGCAAGCATGCACT
 ACCATACTTGGATAATT

The following amino acid sequence <SEQ ID NO. 110> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 50:

*YGS A VSPSKSNLE LQLP*FPCVV EGAWWEITES WGRLPPYCSL GSE*VS*DLI VL*GETPFAW
 FLFSLCWLPK KMLLCSSFVF CHDCGASPAM WKCEEIKPLS FINCPVLGMS LSAV*NGLIH
 QKFKTSLGNT VRPCLYKERI RKRKISVA*Y *TEYYKTD*G TLI*PE*LF* LYHFERPDNQ
 YN*FLSRVAQ TCKHALPYLD N

The following DNA sequence nGPCR-Seq63 <SEQ ID NO. 51> was identified in *H. sapiens*:

CAGTGAGCCG AGATGGTGCC ATTGCACTCT AGCCTGGGGC AACAGAGCCA GACTCCATCT
 CCAAAAAAAA AAGGCCATTG TGAGGATCAA GGCACCACTA GCAACAGGGG GCCCCATGGG
 TCTCAGACCC TCTCCCCACA TCTCCTGGTC CCTGCCCGCA CCTGGCGTAC AGGGACCAGC
 CCCACCGAAG GCTCTTGAGG CCAGGTAACC ATGGGGAGGG GAGGAATGGG GACACCTTCC
 TCCTGAGTGT CTTAGGAAAG AGAACGTTAG GTCAGGTGGC TGAGGGTGGA AATGAGAGAG
 GGGTCTCCTC CTGGAGGGTC TCACCATTCC CTTGGTCACC CACCCAACCTC TCATCTCCCC
 TGATGTGGGG AGGAGCAGGG GGCATGGATT CCTGAGCCCC AGACTCAACT GTTGTGGTTT
 ACAGGGGCAT CAGGAGAGAG AGCGAGCAGA ACACACTCCT GCAGCATCCC CTGGCCCCC
 GCCCCATGAT GGAGCCAGA GAAGCTGGAC AGCACGTGGG GGGCGCAAC GGCGCCAGG
 AGGATGTGGC CTTCAACCTC ATCATCCTGT CCCTCACCGA GGGGCTCGGC CTCGGTGGGC
 TGCTGGGAA TGGGGCAGTC CTCTGGCTGC TCAGCTCCAA TGCTACAGA AACCCCTCG
 CCATCTACCT CCTGGACGTG GCCTGCGCGG ATCTCATCTT CCTTGGCTGC CACATGGTGG
 CCATCGTCCC CGACTTGTG CAAGGCCGGC TGGACTTCCC GGGCTCGTG CAGACCAGCC
 TGGCAACGCT GCGCTTCTTC TGCTACATCG TGGGCTGAG TCTCTGGCG GCCGTAGCG
 TGGAGCAGTG CCTGGCCGCC CTCTCCCAG CCTGGTACTC GTGCCGCCGC CCACGCCACC
 TGACCACCTG TGTGTGCGCC CTCACCTGGG CCCTCTGCCT GCTGCTGCAC CTGCTGCTCA
 GCGGCGCTG CACCCAGTTT TTGGGGAGC CCAGCCGCCA CCTTGTGCCGG ACGCTGTGGC
 TGGTGGCAGC GGTGCTGTG GCTCTGCTGT GTTGCACCAT GTGTGGGCC AGCCTATGC
 TGCTGCTGCG GGTGGAGCGA GGCCCCCAGC GGCCCCCACC CGGGGGCTTC CCTGGGCTCA
 TCCTCCTCAC CGTCCTCCTC TTCTCTTCT GCGGCTGCC CCTCGGCATC TACTGGCTGT
 CCCGGAACCT GCTCTGGTAC ATCCCCCACT ACTTCTACCA CCTCAGCTTC CTCATGGCG
 CCGTGCACTG CGCGGCCAAG CCCGTCGTCT ACTTCTGCCT GGGCAGTGCC CAGGCCGCA
 GGCTGCCCCCT CGGGCTGGTC CTCCAGCGAG CGCTGGGAGA CGAGGCTGAG CTGGGGCCG
 TCAGGGAGAC CTCCGCCGG GGCCTGGTGG ACATAGCAGC CTGAGCCCTG GGGCCCCCGA
 CCCCAGCTGC AGCCCCCGTG AGGCAAGAGG GTGACGTGGG GAAGGTGGTGGTCAAGG
 CTGGGGCCAG CGGACCTGG AGGAGGCCCTT GGTGGGTGAC CCGGTATGT GCTGTCAAAG
 TTGTGACCTT TGGTCTGGAG CATGAGGCTC CCCTGGAGG CAGCTGGAAA GG

The following amino acid sequence <SEQ ID NO. 111> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 51:

MMEPREAGQHVGAAANGAQEDVAFNLIILS**T**TEGLGLGGLLNGA**V**LWLLS
 SNVYRNPFAIYLLDVACADL**I**FLGCHMVAIVPDLLQGR**L**DFPGFVQTSLA
 TLRFCYIVGLSLLAAVS**Q**EQCLAALFP**A**WYS**C**RPRH**L**TCVCALT**W**ALC
 LLLHLLSGACT**Q**FP**G**EP**S**RH**L**CRTL**W**LVA**A**V**L**ALL**C**TM**C**G**A**SL**M****L**
 RVERGPQRPPPRGFPGL**L**LLT**V**LL**F**CG**L**PF**G**Y**W**LSRN**L**W**Y**IP**H**Y**F**
 HFSFLMAAV**H**CA**A**K**P**V**V**Y**F**CL**G**SA**Q**GR**R**PL**R**LV**L**Q**R**AL**G**DE**A**EL**G**AV**R**
 TSRRGLVDIAA

The following DNA sequence nGPCR-Seq70 <SEQ ID NO. 52> was identified in *H. sapiens*:

ATGACGTCCACCTGCACCAACAGCACGCCGAGAGTAAACAGCACGCCACCGTGCATGCC
 CTCTCCAAAATGCCCATCAGCCTGGCCCACGCCATCATCCGCTCAACCGTGTGGTTATC
 TTCTCGCCGCCCTTCCGCGAACATAGTGTCTGGCTAGTGTGAGCGCAAGCCG
 CAGCTGCTGCAGGTGACCAACCGTTTATCTTAAACCTCCTCGTCAACCAGCTGCTGCAG
 ATTTCTGCTGTCGGCCCTGGGTGGCCACCTCTGTCCTCTCTGCCCCCTCAAC
 AGCCACTCTGCACGCCCTGGTTAGCCTCACCCACCTGTCGCTCTCGCCAGCGTCAAC
 ACCATTGTCGTGGTGTCACTGGATCGTACTTGTCCATCATCCACCCCTCTCCTACCCG
 TCCAAGATGACCCAGCGCCGCCGTACCTGCTCCTATGGCACCTGGATTGTGGCCATC
 CTGAGAGCACTCCACTCTACGGCTGGGCCAGGCTGCCCTTGATGAGCGCAATGCT
 CTCTGCTCCATGATCTGGGGGCCAGCCCAAGCTACACTATTCTCAGCGTGGTGTCCCTC
 ATCGTCATTCCACTGATTGTCAATTGCTGCTACTCCGTGGTGTCTGTGAGCGCCGG
 AGGCAGCATGCTCTGCTGACAATGTCAAGAGACACAGCTTGAAGTGCAGGTCAAGGAC
 TGTGTGGAGAATGAGGATGAAGAGGGAGCAGAGAAGAAGGAGGAGTCCAGGATGAGAGT
 GAGTTTCGCCGCCAGCATGAAGGTGAGGTCAAGGCCAAGGAGGGCAGAATGGAAGCCAAG
 GACGGCAGCCTGAAGGCCAAGGAAGGAAGCAGCAGGGGACCAAGTGAAGAGTAGTGTAGAGGCC
 AGGGGCAGCGAGGAGGTCAAGAGAGAGCAGCACGGTGGCCAGCGACGGCAGCATGGAGGGT
 AAGGAAGGCAGCACCAAGTTGAGGAGAACAGCATGAAGGCAGACAAGGTCGCACAGAG

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GTCAACCACTGCAGCATTGACTTGGGTGAAGATGACATGGAGTTGGTGAAGACGACATC
AATTCAGTGAGGATGACGTCGAGGCAGTGAAACATCCCGAGAGCCTCCCACCCAGTCGT
CGTAACAGCAACAGCAACCCCTCTGCCAGGTGCTACCAAGTGCTAAAGTG
ATCTTCATCATCATTTCTCTATGTGCTATCCCTGGGGCCCTACTGCTTTAGCAGTC
CTGGCCGTGTGGATGTCGAAACCCAGGTACCCAGTGGGTGATCACCATAATCATC
TGGCTTTCTCCTGCAGTGCTGCATCCACCCCTATGTCTATGGCTACATGCACAAGACC
ATTAAGAAGGAAATCCAGGACATGCTGAAGAAGTCTCTGCAAGGAAAAGCCCCGAAA
GAAGATAGCCACCCAGACCTGCCCGAACAGAGGGTGGACTGAAGGCAAGATTGTCCCT
TCCTACGATTCTGCTACTTTCTTGA

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The following amino acid sequence <SEQ ID NO. 112> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 52:

```

MTSTCTNSTRESNSSHTCMPLSKMPISLAHGIIRSTVLVIFLAASFVGNIVLALVLQRKP
QLLQVTNRFIFNLLVTDLLQISLVAPWVVATSVPLFWPLNSHFCTAVLSLTHLFAFASVN
TIVVVSVDRYLSIIHPLSYPSKMTQRRGYLLYGTWIVAILQSTPPLYGWGQAAFDERNA
LCSMIWGASPSYTILSVVSFIVIPLIVMIACYSVVFCAARRQHALLYNVKRHSLEVRVKD
CVENEDEEGAEKKEEFQDESEFRRQHEGEVKAKEGRMEAKDGLSKAKEGSTGTSESSVEA
RGSEEVRESSTVASDGSMEGKEGSTKVEENSMKADKGRTEVNQCSIDLGEDDMEFGEDDI
NFSEDDVEAVNIPESLPPSRRNSNSNPPLPRCYQCKAAKVIIIIFSYVLSGPYCFLAV
LAVWVDVETQVPQWVTIIIWLFLQCCIHPYVYGYMHKTIKKEIQDMLKKFFCKEKPPK
EDSHPDLPGTEGGTEGKIVPSYDSATFP

```

The following DNA sequence nGPCR-Seq63 <SEQ ID NO. 53> was identified in *H. sapiens*:

```

ATGAT GGAGCCCAGA GAAGCTGGAC AGCACGTGGG GGCGCCCAAC AGCGCCCAGG
AGGATGTGGC CTTCAACCTC ATCATCCTGT CCCTCACCGA GGGGCTCGGC CTCGGTGGC
TGCTGGGAA TGGGGCAGTC CTCTGGCTGC TCAGCTCAA TGCTACAGA AACCCCTTCG
CCATCTACCT CCTGGACGTG GCCTGCCGCG ATCTCATTT CCTGGCTGC CACATGGTGG
CCATCGTCCC CGACTTGCTG CAAGGCCGG TGGACTTCCC GGGCTCGTG CAGACCAGCC
TGGCAACGCT GCGCTTCTTC TGCTACATCG TGGGCTGAG TCTCCTGGCG GCGTCAGCG
TGGAGCAGTG CCTGGCCGCC CTCTCCCAG CCTGGTACTC GTGCCGCCGC CCACGCCACC
TGACCACCTG TGTGTGCC CTCACCTGGG CCCTCTGCCT GCTGCTGCAC CTGCTGCTCA
GCGGCGCCTG CACCCAGTC TTCGGGAGC CCAGCCGCA TTGTGCCGG ACGCTGTGGC
TGGTGGCAGC GGTGCTGCTG GCTCTGCTGT GTTGACCCAT GTGTGGGCC AGCCTTATGC
TGCTGCTGCG GGTGGAGCGA GGCCCCCAGC GGCCCCCACC CGGGGGCTTC CCTGGGCTCA
TCCTCCTCAC CGTCCTCCTTTCT GCGGCCCTGCC CTTCGGCATC TACTGGCTGT
CCCGGAAACCT GCTCTGGTAC ATCCCCACT ACTTCTACCA CTTCAGCTTC CTCATGGCCG
CCGTGCACTG CGCGGCCAAG CCCGTCGTCT ACTTCTGCCT GGGCAGTGCC CAGGGCCGCA
GGCTGCCCCCT CCGGCTGGTC CTCCAGCGAG CGCTGGGAGA CGAGGCTGAG CTGGGGCCG
TCAGGGAGAC CTCCCGCCGG GGCTGGTGG ACATAGCAGC CTGA

```

The following amino acid sequence <SEQ ID NO. 113> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 53:

```

MMEPREAGQHVGAANSAQEDVAFNLILSTEGLGLGGLLNGAVLWLLLSSNVYR
NPFAIYLLDVACADLIFLGCHMVAIVPDLLQGRDFPGFVQTSLATLRFCYIVGL
SLAAVSVEQCLAALFPAWYSCRPRHLTCVCALTWALCLLLHLLSGACTQFF
GEPSRHLCRTLWLVAAVLLLCTMGASLMLLRVERGPQRPPRGFPLLL
TVLLFLFCGLPFGIYWLSRNLLWYIPHYFYHFSFLMAAVHCAAKPVVYFCLGSAQ
GRRLPLRLVLQRALGDEAELGARETSRRGLVDIAA

```

The following DNA sequence nGPCR-Seq42 <SEQ ID NO. 54> was identified in *H. sapiens*:

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GGAGCCTCGC CGGCCGCCAT GTGGAGCTGC AGCTGGTCA ACGGCACAGG GCTGGTGGAG
GAGCTGCCCTGCCAGGA CCTGCAGCTG GGGCTGTCAC TGTGTCGCT GCTGGGCCCTG
GTGGTGGCG TGCCAGTGGG CCTGTGCTAC AACGCCCTC TGGTGCTGGC CAACCTACAC
AGCAAGGCCA GCATGACCAT GCGGACGTG TACTTTGTCA ACATGGCAGT GGCAGGCCCTG
GTGCTCAGCG CCCTGGCCCC TGTGCACCTG CTCGGCCCC CGAGCCTCCCG GTGGGCCCTG
TGGAGTGTGG CGGGCGAAGT CCACGTGGCA CTGCAGATCC CTTCAATGT GTCCTCACTG

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GTGGCCATGT ACTCCACCGC CCTGCTGAGC CTCGACCACT ACATCGAGCG TGCACTGCCG CGGACCTACA TGGCCAGCGT GTACAAACACG CGGCACGTGT GCGGCTTCGT GTGGGGTGGC GCGCTGCTGA CCAGCTTCTC CTCGCTGCTC TTCTACATCT GCAGCCATGT GTCCACCCGC GCGCTAGAGT GCGCCAAGAT GCAGAACGCA GAAGCTGCCG ACGCCACGCT GGTGTTCATC GGCTACGTGG TGCCAGCACT GGCCACCCCTC TACCGCCTGG TGCTACTCTC CCGCGTCCGC AGGGAGGACA CGCCCTGGA CCGGGACACG GGCCGGCTGG AGCCCTCGGC ACACAGGCTG CTGGTGGCCA CCGTGTGCAC GCAGTTGGG CTCTGGACGC CACACTATCT GATCCTGCTG GGGCACACGG TCATCATCTC GCGAGGGAAG CCCGTGGACG CACACTACCT GGGGCTACTG CACTTGTGA AGGATTCTC CAAACTCTG GCCTCTCCA GCAGCTTGT GACACCACCT CTCTACCGCT ACATGAACCA GAGCTCCCC AGCAAGCTCC AACGGCTGAT GAAAAAGCTG CCTCTGCGGG ACCGGCACTG CTCCCCGGAC CACATGGGG TGCAAGCAGGT GCTGGCGTAG GCGGCCAGC CCTCTGGGG AGACGTGACT CTGGTGGACG CAGAGCACTT AGTTACCCCTG GACGCTCCCC ACATCCCTCC AGAAGGAGAC GAGCTGCTGG AAGAGAAGCA GGAGGGGTGT TTTTCTTGA GTITCCTTT TCCCACAAAT GCCACTCTG GGCCAAGGCT GTGGTCCCCG TGGCTGGCAT CTGGCTTGAG TCTCCCCGAG GCCTGTGCGT CTCCCAAACA CGCAGCTCAA GGTCCACATC CGCAAAAGCC TCCTCGCTT CAGCCTCCCT AGCATTCACT TTGTCATGA AGTGTGAAA GCTTAGAGCC AGTATTATA CTTTGTGGTT AAAATACTTG ATTCCCCCTT GTTTGTTTA CAAAAACAGA TGTTCTTAG AAAAATGACA AATAGAAAA TGAACAAAAC CCTACGAAAG AATGGCAACA GCCAGGGTGG CCGGGCCCTG CCAGTGGGGC GCGTGTGCTA GCAAGGCCTG CCGGGTGTGC CGCAGTCACC ACAGGGTTCT GAGAACATT CACAGAAGTG CCTGAGACGC GGAGACATGG CTGGTGTAA ATGGAGCTAT TCAATAGCAG TGACGCGCTC TCCTCAGCCA CCAAATGTCC CTGACACCCCT CCCCAGCCCC CACAGATAAC ATCAGCTGAG GTTTTTTCA GTATGAACCT GTCTAAATC AATTCTCAA AGTGTGCACA AAACTAAAGA ATATAAATAA ACAAAAGAAA GGCAAAAAAA AAAAAAAA

The following amino acid sequence <SEQ ID NO. 114> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 54:

MWSCSWFNGTGLVEELPACQDQLQLGLSLLSLLGLVVGVPVGLCYNALLVL
ANLHSKASMTMPDVFVNMAVAGLVL SALAPVHLLGPPSSRWALWSVGGE
VHVALQIPFNVSSLVAMYSTALLSLDHYIERALPRTYMASVYNTRHVCDF
VWGGALLTSFSSLLFYICSHVSTRALECAKMQNAAADATL VFFIGYVVPA
LATLYALVLLSRVRREDTPLDRDTGRLEPSAHRLLVATVCTQFGLWTPHY
LILLGHTVIISRGKPVDAHYLGLLFVKDFSKLLAFSSSFVTPLLYRYMN
QSFPSKLQQLMKKLPCGDRHCSPDHMGVQQVLA

The following DNA sequence nGPCR-Seq46 <SEQ ID NO. 55> was identified in *H. sapiens*:

ATGGGGAA CGATTCTGTC AGCTACGAGT ATGGGGATTA CAGCGACCTC TCGGACCGCC CTGTGGACTG CCTGGATGGC GCCTGCCTGG CCATCGACCC GCTGCGCGTG GCCCCGCTCC CACTGTATGC CGCCATCTC CTGGTGGGG TGCCGGGCAA TGCCATGGTG GCCTGGGTGG CTGGGAAGGT GGCCCGCCGG AGGGTGGGTG CCACCTGGTT GCTCCACCTG GCGTGGCGG ATTTGCTGTG CTGTTGTCT CTGCCCATCC TGGCAGTGCC CATTGCCCCTG GGAGGCCACT GGCCGTATGG TGCAGTGGGC TGCGGGCGC TGCCCTCCAT CATCCTGCTG ACCATGTATG CCAGCGTCCT GCTCTGGCA GCTCTCAGTG CCGACCTCTG CTTCTGGCT CTGGGCGCTG CCTGGTGGTC TACGGTTCA CGGGCGTGC GGGTCAGGT GGCTGTGGG GCAGCCTGGA CACTGGCCTT GCTGCTCACC GTGCCCTCCG CCATCTACCG CCGGCTGCAC CAGGAGCACT TCCCAGCCCC GCTGCAGTGT GTGGTGGACT ACGGCGGCTC CTCCAGCACC GAGAATGCGG TGACTGCCAT CCGGTTCTT TTTGGCTTCC TGGGGCCCTT GGTGGCCGTG GCCAGCTGCC ACAGTGCCCT CCGTGTGCTG GCAGCCGAC GCTGCCGGCC GCTGGGCACA GCCATTGTGG TGGGGTTTT TGTCTGCTGG GCACCCCTACC ACCTGCTGGG GCTGGTGTCTC ACTGTGGCGG CCCCCGAACTC CGCACTCCCTG GCCAGGGCCC TGCGGGCTGA ACCCCTCATC GTGGGCGCTG CCCTGCTCA CAGCTGCCCT AATCCCATGC TCTTCTGT A TTTGGGAGG GCTCAACTCC GCGGTCACT GCCAGCTGCC TGTCACTGGG CCCTGAGGGA GTCCCAGGGC CAGGACGAAA GTGTGGACAG CAAGAAATCC ACCAGCCATG ACCTGGTCTC GGAGATGGAG GTGTAG

The following amino acid sequence <SEQ ID NO. 115> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 55:

MGNDSVSYEYGDYSDLSDRPVDCLDGACLAIDPLRVAPLPLYAAIFLVGV
PGNAMVAWVAGKVARRRVGATWLHLAVADLLCCLSLPILAVPLARGHW

PYGAVGCRALPSIILLTMYASVLLAALSADLCFLALGPAWWSTVQRACG
 VQVACGAAWTALLTVPASIYRRLHQEHFPARLQCVVDYGGSSSTENAV
TAIRFLFGFLGPLVAVASCHSALLCWAARRCRPLGTAIVVGFVVCWAPYH
LLGLVLTVAAPNSALLARALRAEPLIVGLALAHSCLNPMFLYFGRAQLR
 RSLPAACHWALRESQGQDESVDKKSTSHDLVSEMEV

The following DNA sequence nGPCR-Seq48 <SEQ ID NO. 56> was identified in *H. sapiens*:

ATGGCACCT TCTCATCGGG CATCACAGGT GGGGTTTGC CCCACCCCTG AACGCCCTCT
 GTGGCGCTT CCACCCACCT GTAGGCCAG AAGGATGTCG GTCTGCTACC GTCCCCCAGG
 GAACGAGACA CTGCTGAGCT GGAAGACTTC GCGGGCCACA GGCACAGCCT TCCTGCTGCT
 GCGCGCCTG CTGGGGCTGC CTGGCACCGG CTTCGTGGTG TGGAGCTTGG CGGGCTGGCG
 GCCTGCACGG GGGCACCGC TGGCGGCCAC GCTTGTGCTG CACCTGGCGC TGGCCGACGG
 CGCGGTGCTG CTGCTCACGC CGCTTTGT GGCCTTCCTG ACCCGGCAGG CCTGGCCGCT
 GGGCCAGGCG GGCTGCAAGG CGGTGTACTA CGTGTGCGCG CTCAGCATGT ACGCCAGCGT
 GCTGCTCACC GCCTGCTCA GCCTGCAGCG CTGCTCGCA GTCACCCGCC CCTTCCTGGC
 GCCTCGGCTG CGCAGCCCCG CCCTGGCCCG CCGCCTGCTG CTGGCGGTCT GGCTGGCCGC
 CCTGTTGCTC GCGTCTCCCG CGCCGCTCA CCGCCACCTG TGGAGGGACC GCGTATGCCA
 GCTGTCACAC CCGTCGCCCG TCCACGCCGC CGCCACCTG AGCCTGGAGA CTCTGACCGC
 TTTCGTGCTT CCTTTCGGGC TGATGCTCGG CTGCTACAGC GTGACGCTGG CACGGCTGCG
 GGGCCGCCGC TGGGGCTCCG GGCGCACCGG GGCGCGGGTG GGCGGCTGG TGAGCGCCAT
 CGTGCTTGCCT TCGGCTTGC TCTGGGCCCC CTACACCGCA GTCAACCTTC TGCAGGGCGT
 CGCAGCGCTG GCTCCACCCG AAGGGGCCCTT GGCGAAGCTG GGCGGAGCCG GCCAGCGGC
 GCGAGCGGGA ACTACGGCCT TGGCCCTTCT CAGTTCTAGC GTCAACCCGG TGCTCTACGT
 CTTCACCGCT GGAGATCTGC TGCCCCGGGC AGGTCCCCGT TTCTCACCGC GGCTCTTCGA
 AGGCTCTGGG GAGGCCCGAG GGGCGGCCCG CTCTAGGGAA GGGACCATGG AGCTCCGAAC
 TACCCCTCAG CTGAAAGTGG TGGGGCAGGG CCGCGGCAAT GGAGACCCGG GGGTGGGAT
 GGAGAAGGAC GGTCCGGAAT GGGACCTTTG A

The following amino acid sequence <SEQ ID NO. 116> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 56:

MAPSHRASQVFCPTPERPLWRLPPTCRPRRMSVCYRPPGNETLLSWKTS
RATGTAFLLAALLGLPGNGFVVWSLAGWRPARGRPLAATLVLHLALADG
AVLLLTLPLFVAFLTRQAWPLGQAGCKAVYYVCALSMYASVLLTGLLSLQR
 CLAVTRPFLAPRLRSPALARRLLAVWLAALLAVPAAVYRHLWRDRVCQ
 LCHPSPVHAAAHLSLETLTAFVLPFGMLGCYSVTLARLRGARWGSGRHG
 ARVGRLVSIAVLAFLGLLWAPYHAVNLLQAVAALAPPEGALAKLGGAGQAA
 RAGTTALAFFSSSVNPVLYVFTAGDLLPRAGPRLTRLFEGSGEARGGGR
 SREGTMELRTPQLKVVGQGRNGDPGGGMEKDGPWEWDL

The following DNA sequence nGPCR-Seq51 <SEQ ID NO. 57> was identified in *H. sapiens*:

CCGCCCGCG GGGAGGACCG GAGCACCCAG CTTTAACTCCC TGGAAAGTC ACGAACAAATG
 AATCCATTTCAATGCATCTTG TTGGAACACC TCTGCCGAAC TTTTAAACAA ATCCCTGGAAAT
 AAAGAGTTG CTTATCAAAC TGCCAGTGTG GTAGATACAG TCATCCTCCC TTCCATGATT
 GGGATTATCT GTTCAACAGG GCTGGTTGGC AACATCCTCA TTGTATTAC TATAATAAGA
 TCCAGGAAA AAACAGTCCC TGACATCTAT ATCTGCAACC TGGCTGTGGC TGATTTGGTC
 CACATAGTTG GAATGCCTTT TCTTATTAC CAATGGGCC GAGGGGGAGA GTGGGTGTTT
 GGGGGCCTC TCTGCACCAT CATCACATCC CTGGATACTT GTAACCAATT TGCTGTAGT
 GCCATCATGA CTGTAATGAG TGTGGACAGG TACTTTGCC TCGTCCAACC ATTCGACTG
 ACACGGTGG AACAAGGTA CAAGACCATC CGGATCAATT TGGGCCCTTIG GGCAGCTTCC
 TTATCCTGG CATTGCCCTGT CTGGGTCTAC TCGAAGGTCA TCAAATTAA AGACGGTGT
 GAGAGTTGTG CTTTGATTT GACATCCCCT GACGATGTAC TCTGGTATAC ACTTATTTG
 ACGATAACAA CTTTTTTTCCCTCATCCC TTGATTITGG TGTGCTATAT TTTAATTAA
 TGCTATACCTT GGGAGATGTA TCAACAGAAAT AAGGATGCCA GATGCTGCAA TCCCAGTGTA
 CAAAAACAGA GAGTGTGAA GTTGACAAAG ATGGTGCTGG TGCTGGTGGT AGTCTTTATC
 CTGAGTGCTG CCCCTTATCA TGTGATACAA CTGGTGAAC TACAGATGGA ACAGCCACA
 CTGGCCTTCT ATGTGGGTTA TTACCTCTCC ATCTGCTCTCA GCTATGCCAG CAGCAGCATT
 AACCCCTTTC TCTACATCCT GCTGAGTGG AATTTCAGA AACGTCTGCC TCAAATCCAA

AGAAGAGCGA CTGAGAAGGA AATCAACAAT ATGGGAAACA CTCTGAAATC ACACITTTAG GAAAGTACAT GGATCACCAT GAGTCTAGAC ATGATTGTCT ATCTTACTGG TATTATTAGA AAGGGCAGGT GTACCGATAT GTTATGCC ATTCTTCTG TGTACTTGTG ACTCTTAGCA GCATGGAAGA GAAGTGTAAAC CATGCAAATA CAATGAGCTT AATATGCTAA CTTAGCAAG ATGTAAAATG TTGATCTATA TTGTGGGTAG GGAATGGGAT AGTCTGAGAT ACCCAGGCTT CATGATGGTG TATATTATT CAGCATATTA TAAACTAGTC ACTAATGAAA ATGCCATCC ATGACCATTG ACTCAAAACT CACCAAGGAA CCTGACCITG CCCTCCACAC TGTGGCCTCA CTGTAACAGT TTCTCAAGG TTCTAGGAG GGTATCACCT TAGAGTGAAG TCTAAAATTG GGCTATTTT TATCTATAAA AAATGTCAGT TTATATGGT CCAACTAA TACCCCTCAAC AACTAAGCCC CACCTTTAG AATAAGTTAC CATTATTGC ACACATGCAA TGTGTAAGAT TACATGTAAC AAACCTGTGA AATAAGTATT ATTACCTTG TTTGCTAAGG CTCAGAAAGG AGAAATGATA GGCTAATGC TGCAACAGCT ATCTAAGAGC TGAGCTAACAA TTCAGCTCTG CCTGTTCTT TTCTACTGCC GACCTTGACA ACCTTTACTT ATCATACTGG AGAACCCAGT AACTTGGAGT TTCTTTGCT TTCTCTGTAA GCCCTACAAG AGGAGAACTA AAGTCTGATA GAAATGAGTT GATGTTTAA GCATCATTT GGATTATCTT GTTCTCACAC CTGCTAATG TAGAAACTGG CATCTGGACT TTAATAATAA TACTTTACTT CTGGA

The following amino acid sequence <SEQ ID NO. 117> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 57:

MNPFHASCWNNTSAELLNKSWNKEFAYQTASVVDTVILPSMIGIICSTGLVGNILIVFTIIRSRKKTVP
DIYICNLAVADLV HIVGMPFLIHQWARGGEWWFGGPLCTIITS DTCNQFACSAIMTVMSVDR
YFALVQPFLTRWRTRYKTI RNLGLWAASFILALPVWVYSKVIFKFDGV
ESCAFDLTSPDDVLWYTL YLT TTTFFPLPLILVCYIILCYTWEMYQQN
KDARCCNPSVPKQRVMKLT KMVLVLLVVFILSAAPYHVIQLVNLQMEQPT
LAFYVGYYLSICL SYASSSINPFLYIILSGNFQKRLPQIQRRATEKEINN
MGNTLKS HF

The following DNA sequence nGPCR-Seq52 <SEQ ID NO. 58> was identified in *H. sapiens*:

CCATGACCAG CAATTTTCC CAACCTGTTG TGCAGCTTG CTATGAGGAT GTGAATGGAT CTTGTATTGA AACTCCCTAT TCTCTGGGT CCCGGGTAAT TCTGTACACG GCGTTTAGCT TTGGGTCTTT GCTGGCTGTA TTTGAAATC TCTTAGTAAT GACTCTGTT CTTCATTTA AGCAGCTGCA CTCTCCAACC AATTTCCTCA TTGCTCTCT GGCCTGTGCT GACTCTTGG TAGGTGTGAC TGTGATGCTT TTCAAGCATGG TCAGGACGGT GGAGAGCTGC TGGTATTTG GAGCCAATT TTGTA^TCTT CACAGTTGCT GTGATGTTGC ATTTGTTAC TCTTCTGTCC TCCACTTGTC TTCACTGTC ATCGACAGGT ACATTGTTGC TACTGATCCC CTGGTCTATG CTACCAAGTT CACCGTGTCT GTGTCGGGAA TTGCA^TTCAG CGTGTCTGG ATTCTGCCTC TCACGTACAG CGGTGCTGTG TTCTACACAG GTGTCAATGA TGATGGGCTG GAGGAATTAG TAAGTGTCT CAACTGCGTA GGTGGCTGTC AAATTATTGT AAGTCAAGGC TGGGTGTTGA TAGATTTCCTT GTTATTCCTC ATACCTACCC TTGTTATGAT AATTCTTAC AGTAAGATT TTCTTATAGC TAAACAACAA GCTATAAAAAA TTGAAACTAC TAGTAGCAA GTAGAAATCAT CCTCAGAGAG TTATAAAATC AGAGTGGCCA AGAGAGAGAG GAAAGCAGCT AAAACCCCTGG GGGTCACGGT ACTAGCATTT GTTATTCAT GGTACCGTA TACAGTTGAT ATATTAATTG ATGCCCTTAT GGGCTTCTG ACCCTGCT ATATCTATGA AATTGCTGT TGGAGTGTCTT ATTATAACTC AGCCATGAAT CCTTGATTT ATGCTCTATT TTATCCTGG TTAGGAAAG CCATAAAACT TATTITAAGT GGAGATGTT TAAAGGCTAG TTCATCAACC ATTAGTTAT TTTAGAATAA

The following amino acid sequence <SEQ ID NO. 118> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 58:

MTSNFSQPV VQLCYEDVNG SCIETPYSPG SRVILYTAFS FGSLLA^TVFGN LLVMTSVLHF KQLHSPTNFL IASLACADFL VGVTVMLFSM VRTVESCVYF GAKFCTLHSC CDVAFCYSSV LHLCFICIDR YIVVTDPLVY ATKFTVSVSG ICISVSWILP LTYSGA^TFYT GVNDG^TLEEL VSALNCVGGC QIIVSQGWVL IDFLFFFPT LVMILYSKI FLIAKQQAIK IETTSSKVES SSESYKIRVA KRERKAAKTL GVTVLAFVIS WLPYTV^TDILI DAFMGFLTPA YIYEICCWSA YYNSAMNPLI YALFYPWFRK AIKLILSGDV LKASSSTISL FLE

The following DNA sequence nGPCR-Seq49 <SEQ ID NO. 59> was identified in *H. sapiens*:

A **TGCCACTCCC TGTGCCCTCA GCGGGGGCAC AAAAGACTCC TGAGGACCAT GTCTGTCCTCC**
ATCTTGCAGG TCCCTCGCCA GCACCCCTCTG AGCCGGCCAG GATGTTGGG CTGTTCGGCC
TCTGGAGAAC CTTCGACAGT GTGGTCTTCT ACCTGACGCT GATCGTGGGC CTCGGGGGAC
CGGTAGGTA CGGGCTGGTG CTCTGGAACC TCGGCTTCCG CATCAAGAAC GGCCCCTTCT
CCATCTACCT GCTGCACCTG GCCGCCGCG ACTTCCCTGTT CCTCTCCTGC CGTGTGGGCT
TCTCCGTGGC TCAGGCTGCC CTGGGCGCCC AGGACACACT CTACTTCGTG CTCACCTTCC
TGTGGITCGC GGTGGGGCTC TGGCTGCTGG CGGCCCTCAG CGTGGAGCGC TGCCTCTCCG
ACCTCTTCCC CGCCTGCTAC CAGGGCTGCC GGCCCAGACA CGCCTCGGCC GTCCTCTGCG
CCCTGGTGTG GACCCGACCT TGCCGGCG TGCCGCTGCC CGCCAACGCC TGCGGCCCTGC
TGCGCAACAG CGCGTGCCTCC CTGGTCTGCC CGCGCTACCA CGTGGCCAGC GTCACCTGGT
TCCTGGTGCT GGCCCACGTC GCCTGGACGG CTGGCTGTT CCTCTTGTG TGGGTGACCT
GCTGCTCCAC TCGCCCGCGG CCCAGGCTCT ACGGCATCGT CCTGGGCGCG TGCTCCTGC
TCTTCTTCTG TGGCCTGCC TCGGTCTTCT ACTGGAGCCT GCAGCCCTG CTGAACCTCC
TGCTGCCCGT GTTTCCCCG CTGGCCACGC TGCTGCCCTG CGTCAACAGC AGCTCCAAGC
CCCTCATCTA CTCGGGGTTG GGCCGACAGC CGGGGAAGCG GGAGCCGCTG AGGTGGTAC
TGCGGAGGGC CCTGGGGGAG GGCGCCGAGC TGGGTGCCAG GGGACAGTCC CTGCCATGG
GTCTCCTATA A

The following amino acid sequence <SEQ ID NO. 119> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 59:

MPLPVPPAGAQKTPEDHVCLHLAGPSPAPSEPARMFGFLGLWRTFDSVVFYLTLLIVGLGGP
VGNGLVLWNLGFRIKKGPFSTIYLLHLAAADFLFLSCRVGSVAQAAALGAQDTLYFVLTFLWFAVG
LWLLAAFSVERCLSDLFPACYQGCRPRHASAVLCAVWTPTLPAVPLPA NACGLLRNSACPLVCP
RYHVASVTWFLVLARVAWTAGVVLVWVTCSTRPRPRLYGIVLGALLLFFCGLPSVF YWSLQP
LLNFLPVPFSPLATLLACVNSSSKPLIYSGLGRQPGKREPLRSVLRRALGEGAELGARGQSLPMGLL

The following DNA sequence nGPCR-Seq61 <SEQ ID NO. 60> was identified in *H. sapiens*:

ATGAACAACAATACAACATGTATTCAACCACATCTATGATCTCTCCATGGCTTACCAATC
 ATTTACATCCTCTTGTATTGTTGGTGTGGAAACACTCTCTCAATGGATATT
 TTAACAAAAATAGGTAACATCAACGACATCTACCTGTACACACCTTGACTGCA
 AACTTACTTGTGTGCAGTGCATGCCATGCCATGAGTATCTATTCTGAAAGTTCCA
 TGGGAATATCAATCTGCTCAATGCAAGAGTGTCAATTCTGGAACTCTATCCATGCA
 GCAAGTATGTTGTCACTCTTAAATTAAAGTTGGATTGCCATAAGCCCTATGCTACC
 TTAATGCAAAGGATTCTCGCAAGAGACTACTTCATGCTATGAGAAAATATTTATGG
 CATTACTGAAAAAAATTGCCAGGCCACTTTGCTAGAAAATATGCAATTACATATGG
 GGAGTTGTACTGGGCATAATCATTCCAGTTACCGTATACTACTCAGTCATAGAGGCTACA
 GAAGGAGAAGAGAGCCTATGCTACAATGGCAGATGGAACTAGGAGCCATGATCTCTCAG
 ATTGCAGGTCTCATTGGACCACATTATTGGATTCTTCTTCTAGTAGTACTAACATCA
 TACTACTCTTGTAAAGCCATCTGAGAAAAATAAGAACCTGTACGTCCATTATGGAGAAA
 GATTGACTTACAGTCTGTGAAAAGACATCTTGTACCCAGATTCTACTAATAGTT
 TGCTTCTTCTTCTATAGTATTAAACCCATTCTTATGTTCTACACCAAAGAGATAAC
 TGTCAGCAATTGAATTATTAATAGAAACAAAAACATTCTCACCTGTCTGCTTCCGGCC
 AGAAGTAGCACAGACCCATTATATTCTTATTAGACAAAACATTCAAGAACACTA
 TATAATCTTCTTACAAAGTCTAATTGACACATATGCAATCATATGGTGA

The following amino acid sequence <SEQ ID NO. 120> is the predicted amino acid sequence derived from the DNA sequence of SEQ ID NO. 60:

MNNNTTCIOPSMISSMALPIYILLCIVGVFGNTLSQWFLTKIGKKTSTHIYLSHLVT
ANLLVCSAMPFMSIYFLKGFWQEYQSAQCRVVFNLGTL SMHASMFVSLLILSWIAISRY
ATLMQKDSSQETTSCYEKIFYGHLLKKFRQPNFARKLCIYIWGVVLGIII PVTVVYSVI
EATEGEESL CYNRQMELGAMISQIAGLIGTTFIGFSFLVVLTSYYSFVSHLRKIRTCTS
IMEKDLTYSSVKRHL LVIQILLIVCFLPYSIFKPIFYVLHQRDNCQQLNYLIETKNILT
CLASARSSTDPIFL LLDKTFKKTLYNLFTKSNSAHMQSYG

Example 2: Cloning of nGPCR-x

cDNAs may be sequenced directly using an AB1377 or ABI373A fluorescence-based sequencer (Perkin Elmer/Applied Biosystems Division, PE/ABD, Foster City, CA) and the ABI PRISM Ready Dye-Deoxy Terminator kit with Taq FS polymerase. Each ABI cycle

5 sequencing reaction contains about 0.5 µg of plasmid DNA. Cycle-sequencing is performed using an initial denaturation at 98C for 1 minute, followed by 50 cycles: 98C for 30 seconds, annealing at 50C for 30 seconds, and extension at 60C for 4 minutes. Temperature cycles and times are controlled by a Perkin-Elmer 9600 thermocycler. Extension products are purified 10 using Centriflex gel filtration (Advanced Genetic Technologies Corp., Gaithersburg, MD). Each reaction product is loaded by pipette onto the column, which is then centrifuged in a swinging bucket centrifuge (Sorvall model RT6000B table top centrifuge) at 1500 x g for 4 minutes at room temperature. Column-purified samples are dried under vacuum for about 40 minutes and then dissolved in 5 µl of a DNA loading solution (83% deionized formamide, 8.3 mM EDTA, and 1.6 mg/ml Blue Dextran). The samples are then heated to 90C for three minutes and loaded 15 into the gel sample wells for sequence analysis by the ABI377 sequencer. Sequence analysis is done by importing ABI373A files into the Sequencher program (Gene Codes, Ann Arbor, MI). Generally, sequence reads of 700 bp are obtained. Potential sequencing errors are minimized by obtaining sequence information from both DNA strands and by re-sequencing difficult areas using primers at different locations until all sequencing ambiguities are removed.

20 To isolate a cDNA clone encoding full length nGPCR, a DNA fragment corresponding to a nucleotide sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60, or a portion thereof, can be used as a probe for hybridization screening of a phage cDNA library. The DNA fragment is amplified by the polymerase chain reaction (PCR) method. The PCR reaction mixture of 50 ml contains polymerase mixture (0.2 mM dNTPs, 1x PCR Buffer and 25 0.75 ml Expand High Fidelity Polymerase (Roche Biochemicals)), 1 µg of 3206491 plasmid, and 50 pmoles of forward primer and 50 pmoles of reverse primer. The primers are preferably 10 to 25 nucleotides in length and are determined by procedures well known to those skilled in the art. Amplification is performed in an Applied Biosystems PE2400 thermocycler, using the following program: 95C for 15 seconds, 52C for 30 seconds and 72C for 90 seconds; repeated 30 for 25 cycles. The amplified product is separated from the plasmid by agarose gel electrophoresis, and purified by Qiaquick gel extraction kit (Qiagen).

A lambda phage library containing cDNAs cloned into lambda ZAPII phage-vector is plated with *E. coli* XL-1 blue host, on 15 cm LB-agar plates at a density of 50,000 pfu per plate, and grown overnight at 37C; (plated as described by Sambrook et al., supra). Phage plaques are

transferred to nylon membranes (Amersham Hybond NJ), denatured for 2 minutes in denaturation solution (0.5 M NaOH, 1.5 M NaCl), renatured for 5 minutes in renaturation solution (1 M Tris pH 7.5, 1.5 M NaCl), and washed briefly in 2xSSC (20x SSC: 3 M NaCl, 0.3 M Na-citrate). Filter membranes are dried and incubated at 80C for 120 minutes to cross link the 5 phage DNA to the membranes.

The membranes are hybridized with a DNA probe prepared as described above. A DNA fragment (25 ng) is labeled with α -32P-dCTP (NEN) using Rediprime random priming (Amersham Pharmacia Biotech), according to manufacturers instructions. Labeled DNA is separated from unincorporated nucleotides by S200 spin columns (Amersham Pharmacia 10 Biotech), denatured at 95C for 5 minutes and kept on ice. The DNA-containing membranes (above) are pre-hybridised in 50 ml ExpressHyb (Clontech) solution at 68C for 90 minutes. Subsequently, the labeled DNA probe is added to the hybridization solution, and the probe is left to hybridise to the membranes at 68C for 70 minutes. The membranes are washed five times in 2x SSC, 0.1% SDS at 42C for 5 minutes each, and finally washed 30 minutes in 0.1x SSC, 0.2% 15 SDS. Filters are exposed to Kodak XAR film (Eastman Kodak Company, Rochester, N.Y., USA) with an intensifying screen at -80C for 16 hours. One positive colony is isolated from the plates, and replated with about 1000 pfu on a 15 cm LB plate. Plating, plaque lift to filters and hybridization are performed as described above. About four positive phage plaques are isolated form this secondary screening.

20 cDNA containing plasmids (pBluescript SK-) are rescued from the isolated phages by in vivo excision by culturing XL-1 blue cells co-infected with the isolated phages and with the Excision helper phage, as described by manufacturer (Stratagene). XL-blue cells containing the plasmids are plated on LB plates and grown at 37C for 16 hours. Colonies (18) from each plate are replated on LB plates and grown. One colony from each plate is stricken onto a nylon filter 25 in an ordered array, and the filter is placed on a LB plate to raise the colonies. The filter is then hybridized with a labeled probe as described above. About three positive colonies are selected and grown up in LB medium. Plasmid DNA is isolated from the three clones by Qiagen Midi Kit (Qiagen) according to the manufacturer's instructions. The size of the insert is determined by digesting the plasmid with the restriction enzymes NotI and SalI, which establishes an insert 30 size. The sequence of the entire insert is determined by automated sequencing on both strands of the plasmids.

nGPCR-70

The PCR reaction was performed in 50 μ l containing 31 μ l H₂O, 5 μ l Buffer II (PE Applied Biosystems AmpliTaq Gold system), 6 μ l 25 mM MgCl₂, 2 μ l 10 mM dNTP mix, 5 μ l

Marathon Ready whole human brain cDNA (Clontech #7400-1), 0.3 μ l primer VR70C (1 μ g/ μ l)(SEQ ID NO:121), 0.3 μ l primer VR70D (1 μ g/ μ l)(SEQ ID NO:122), and 0.4 μ l AmpliTaq GoldTM DNA Polymerase. The primer sequence for VR70C is 5'-
5' TTCAAAGCTTATGACGTCCACCTGCACC-3' (SEQ ID NO:121), corresponding to the 5'
end of the coding region and containing a HindIII restriction site. The primer sequence for
VR70D is 5'- TTCACTCGAGTCAAGGAAAAGTAGCAGAATCGTAG-3' (SEQ ID
NO:122), corresponding to the 3' end of the coding region and containing an XhoI restriction
site (Genosys).

The PCR reaction was carried out using a GeneAmp PCR 9700 thermocycler (Perkin
10 Elmer Applied Biosystems) and started with 1 cycle of 80C for 20 minutes followed by 95C for
10 minutes, then 12 cycles at 95C for 30 seconds, 72C for 2 minutes decreasing 1C each cycle,
72C for 1 minute, followed by 35 cycles at 95C for 30 seconds, 60C for 30 seconds, 72C for 1
minute. The PCR reaction was loaded on a 0.75% gel. The DNA band was excised from the gel
and the DNA was eluted from the agarose using a QIAquick gel extraction kit (Qiagen). The
15 eluted DNA was ethanol-precipitated and resuspended in 4 μ l H₂O for ligation. The ligation
reaction consisted of 4 μ l of fresh ethanol-precipitated PCR product and 1 μ l of pCRII-TOPO
vector (Invitrogen). The reaction was gently mixed and allowed to incubate for 5 minutes at
room temperature followed by the addition of 1 μ l of 6x TOPO cloning stop solution and mixing
for 10 seconds at room temperature. The sample was then placed on ice and 2 μ l was
20 transformed in 50 μ l of One Shot cells (Invitrogen) and plated onto ampicillin plates. Seven
white colonies were chosen and the presence of an insert was verified by PCR in the following
manner. Each colony was resuspended in 2 ml LB broth and incubated at 37C for 2 hours. A
500 μ l aliquot was spun down in the microfuge, the supernatant discarded, and the pellet
resuspended in 25 μ l of H₂O. A 16 μ l aliquot was removed and boiled for 5 minutes and the
25 sample was placed on ice. The sample was microfuged briefly to pellet any bacterial debris and
PCR was carried out as described above with 15 μ l of sample using primers VR70C (SEQ ID
NO:121) and VR70D (SEQ ID NO:122).

nGPCR-63

Isolation of a clone for nGPCR-63 from genomic DNA was performed by PCR in a 50
30 μ l reaction containing Herculase DNA Polymerase blend (Stratagene), with buffer
recommendations as supplied by the manufacturer, 200 ng each primers PSK16 and 17 (SEQ ID
NO:123 and SEQ ID NO:124, respectively), 150 ng of human genomic DNA (Clontech) and 4%
DMSO. The PCR reaction was performed on a Robocycler thermocycler (Stratagene)
starting with 1 cycle of 94C for 2 minutes followed by 35 cycles of 94C for 30 seconds, 65C for

30 seconds, 72C for 2 minutes. The PCR reaction was purified by the QiaQuick PCR Purification Kit (Qiagen) and eluted in TE. The PCR primer sequences were: PSK16 5'-GATCGAATTCA**T**GATGGAGCCCAGAGAAGCTGGAC-3' (SEQ ID NO:123) and PSK17 5'-GATCCTCGAGTCAGGCTGCTATGTCCACCAGGCC-3' (SEQ ID NO:124). Translation 5 initiation and termination codons are shown above in bold.

The PCR product was ligated into the pCR-BluntII-TOPO vector (Invitrogen) using the Zero Blunt Topo PCR TA cloning kit as follows. 3 μ l PCR product DNA, 1 μ l pCRII-TOPO vector, and 1 μ l TOPOII salt solution (1.2M NaCl, 0.06M MgCl₂) was incubated for 5 minutes at room temperature. To the ligation reaction one microliter of 6X TOPO Cloning Stop Solution 10 was added then the reaction was placed on ice. Two microliters of the ligation reaction was transformed in One-Shot TOP10 cells (Invitrogen), and placed on ice for 30 minutes. The cells were heat-shocked for 30 seconds at 42C, placed on ice for two minutes, 250 μ l of SOC was added, then incubated at 37C with shaking for one hour and then plated onto ampicillin plates 15 supplemented with Xgal and IPTG. Single colonies were screened by restriction digestion for the presence of the insert, and a plasmid DNA from colony 63-4-23 was purified using a Qiagen Endo-Free plasmid purification kit.

The clone containing nGPCR-63 was sequenced directly using an ABI377 fluorescence-based sequencer (Perkin Elmer/Applied Biosystems Division, PE/ABD, Foster City, CA) and the ABI BigDyeTM Terminator Cycle Sequencing Ready Reaction kit with Taq FSTM polymerase. Each ABI cycle sequencing reaction contained about 0.5 μ g of plasmid DNA. 20 Cycle-sequencing was performed using an initial denaturation at 98C for 1 minute, followed by 50 cycles: 96C for 30 seconds, annealing at 50C for 30 seconds, and extension at 60C for 4 minutes. Temperature cycles and times were controlled by a Perkin-Elmer 9600 thermocycler. Extension products were purified using AGTC (R) gel filtration block (Edge Biosystems, 25 Gaithersburg, MD). Each reaction product was loaded by pipette onto the column, which was then centrifuged in a swinging bucket centrifuge (Sorvall model RT6000B tabletop centrifuge) at 1500 x g for 4 minutes at room temperature. Column-purified samples were dried under vacuum for about 40 minutes and then dissolved in 3 μ l of a DNA loading solution (83% deionized formamide, 8.3 mM EDTA, and 1.6 mg/ml Blue Dextran). The samples were then 30 heated to 90C for 3.5 minutes and loaded into the gel sample wells for sequence analysis by the ABI377 sequencer. Sequence analysis was performed by importing ABI377 files into the Sequencer program (Gene Codes, Ann Arbor, MI).

nGPCR-42

PCR was performed in a 50 μ l reaction containing 36.7 μ l H₂O, 5 μ l 10X TT buffer (140 mM Ammonium Sulfate, 0.1 % gelatin, 0.6 M Tris-tricine pH 8.4), 5 μ l 15 mM MgSO₄, 2 μ l 10 mM dNTP, 0.3 μ l 6541947H1 DNA (1.4 Tg/Tl), 0.3 μ l of LW1579 (1 μ g/ μ l), 0.3 μ l of LW1580 (1 μ g/ μ l), 0.4 μ l High Fidelity Taq polymerase (Boehringer Mannheim). The PCR reaction was 5 performed on a Robocycler thermocycler (Stratagene) starting with 1 cycle of 94C for 2 minutes followed by 15 cycles of 94C for 30 seconds, 55C for 30 seconds, 68C for 1.3 minutes. The PCR reaction was loaded onto a 1.2 % agarose gel. The DNA band was excised from the 10 gel, placed in GenElute Agarose spin column (Supelco) and spun for 10 minutes at maximum speed in a microcentrifuge. The eluted DNA was EtOH precipitated and resuspended in 4 μ l H₂O for ligation. The forward PCR primer was: LW1579 5'-GCATAAGCTTCCATGTGGAGCTGC AGCTGGTTCAACG-3' (SEQ ID NO:125), and the reverse PCR primer was: LW1580 5'-GCATCTCGAGCCTACGCCAGCACCTGCTGCACC-3' (SEQ ID NO:126). The ligation 15 reaction used solutions from the TOPO TA Cloning Kit (Invitrogen) which consisted of 4 μ l PCR product DNA and 1 μ l pCRII-TOPO vector that was incubated for 5 minutes at room temperature. To the ligation reaction one microliter of 6X TOPO Cloning Stop Solution was added then the reaction was placed on ice. Two microliters of the ligation reaction was transformed in One-Shot TOP10 cells (Invitrogen), and placed on ice for 30 minutes. The cells were heat-shocked for 30 seconds at 42C, placed on ice for two minutes, 250 μ l of SOC was added, then incubated at 37C with shaking for one hour and then plated onto ampicillin plates. A 20 single colony containing an insert was used to inoculate a 5 ml culture of LB medium. Plasmid DNA was purified using a Concert Rapid Plasmid Miniprep System (GibcoBRL) and then sequenced.

nGPCR-46

PCR was performed in a 50 μ l reaction using components that come with PLATINUM® 25 *Pfx* DNA Polymerase (GibcoBRL) containing 30.5 μ l H₂O, 5 μ l 10X *Pfx* Amplification buffer, 5 μ l 10X Enhancer solution, 1.5 μ l 50mM MgSO₄, 2 μ l 10 mM dNTP, 5 μ l human genomic DNA (0.3 μ g/ μ l)(Clontech), 0.3 μ l of LW1626 (1 μ g/ μ l), 0.3 μ l of LW1627 (1 μ g/ μ l), 0.4 μ l PLATINUM® *Pfx* DNA Polymerase (2.5 U/ μ l). The PCR reaction was performed in a 30 Robocycler Gradient 96 (Stratagene) starting with 1 cycle of 94C for 5 minutes followed by 30 cycles at 94C for 40 seconds, 55C for 2 minutes, 68C for 3 minutes. Following the final cycle, 0.5 μ l of AmpliTaq DNA Polymerase (5 U/ μ l) was added and the tube was incubated at 72C for 5 minutes. The PCR reaction was loaded onto a 1.2 % agarose gel. The DNA band was excised from the gel, placed in GenElute Agarose spin column (Supelco) and spun for 10 minutes at maximum speed in a microcentrifuge. The eluted DNA was EtOH precipitated and resuspended

in 12 μ l H₂O for ligation. The forward PCR primer was: LW1626 5'-GCATAAGCTTCCATGGGGAACGATTCTGTCAGC-3' (SEQ ID NO:127) and the reverse PCR primer was: LW1627 5'-GCATCTCGAGCCTACACCTCCATCTCCGAGACC-3' (SEQ ID NO:128). The ligation reaction used solutions from the TOPO TA Cloning Kit (Invitrogen) which consisted of 4 μ l PCR product DNA and 1 μ l pCRII-TOPO vector that was incubated for 5 minutes at room temperature. To the ligation reaction one microliter of 6X TOPO Cloning Stop Solution was added then the reaction was placed on ice. Two microliters of the ligation reaction was transformed in One-Shot TOP10 cells (Invitrogen), and placed on ice for 30 minutes. The cells were heat-shocked for 30 seconds at 42C, placed on ice for two minutes, 250 μ l of SOC was added, then incubated at 37C with shaking for one hour and then plated onto ampicillin plates. A single colony containing an insert was used to inoculate a 5 ml culture of LB medium. Plasmid DNA was purified using a Concert Rapid Plasmid Miniprep System (GibcoBRL) and then sequenced.

nGPCR-48

15 PCR was performed in a 50 μ l reaction using components that come with PLATINUM® Pfx DNA Polymerase (GibcoBRL) containing 30.5 μ l H₂O, 5 μ l 10X Pfx Amplification buffer, 5 μ l 10X Enhancer solution, 1.5 μ l 50mM MgS0₄, 2 μ l 10 mM dNTP, 5 μ l human genomic DNA (0.3 μ g / μ l)(Clontech), 0.3 μ l of LW1572 (1 μ g/ μ l), 0.3 μ l of LW1573 (1 μ g/ μ l), 0.4 μ l PLATINUM® Pfx DNA Polymerase (2.5 U/ μ l). The PCR reaction was performed in a 20 Robocycler Gradient 96 (Stratagene) starting with 1 cycle of 94C for 5 minutes followed by 30 cycles at 94C for 40 seconds, 55C for 2 minutes, 68C for 3 minutes. Following the final cycle, 0.5 μ l of AmpliTaq DNA Polymerase (5 U/Tl) was added and the tube was incubated at 72C for 5 minutes. The PCR reaction was loaded onto a 1.2 % agarose gel. The DNA band was excised from the gel, placed in GenElute Agarose spin column (Supelco) and spun for 10 minutes at 25 maximum speed in a microcentrifuge. The eluted DNA was EtOH precipitated and resuspended in 12 μ l H₂O for ligation. The forward primer for PCR was: LW1572 5'-GATCAAGCTTGCA TGGCACCTTCTCATCGGG-3' (SEQ ID NO:129) and the reverse primer was: LW1573 5'-GATCCTCGAGTCAAAGGTCCCATTCCGGACC-3' (SEQ ID NO:130). The ligation reaction used solutions from the TOPO TA Cloning Kit (Invitrogen) which consisted of 4 μ l PCR product 30 DNA and 1 μ l pCRII-TOPO vector that was incubated for 5 minutes at room temperature. To the ligation reaction one microliter of 6X TOPO Cloning Stop Solution was added then the reaction was placed on ice. Two microliters of the ligation reaction was transformed in One-Shot TOP10 cells (Invitrogen), and placed on ice for 30 minutes. The cells were heat-shocked for 30 seconds at 42C, placed on ice for two minutes, 250 μ l of SOC was added, then incubated

at 37C with shaking for one hour and then plated onto ampicillin plates. A single colony containing an insert was used to inoculate a 5 ml culture of LB medium. Plasmid DNA was purified using a Concert Rapid Plasmid Miniprep System (GibcoBRL) and then sequenced.
nGPCR-49

5 The PCR reaction used components that come with PLATINUM® *Pfx* DNA Polymerase (GibcoBRL) containing 30.5 μ l H₂O, 5 μ l 10X *Pfx* Amplification buffer, 5 μ l 10X Enhancer Solution, 1.5 μ l 50mM MgSO₄, 2 μ l 10 mM dNTP, 5 μ l human genomic DNA (0.3 μ g/ μ l) (Clontech), 0.3 μ l of LW1726 (1 μ g/ μ l) (SEQ ID NO:131), 0.3 μ l of LW1727 (1 μ g/ μ l) (SEQ ID NO:132), 0.4 μ l PLATINUM® *Pfx* DNA Polymerase (2.5 U/ μ l). The PCR reaction was
10 performed in a Robocycler Gradient 96 (Stratagene) starting with 1 cycle of 94C for 2 minutes followed by 35 cycles at 94C for 30 seconds, 55C for 30 seconds, 68C for 2 minutes. Following the final cycle, 0.5 μ l of AmpliTaq DNA Polymerase (5 U/ μ l) was added and the tube was incubated at 72C for 5 minutes. The sequence of LW1726 is: 5'-CATAAGCTTGGATGCCA
15 CTCCCTGTGCCCCC-3' (SEQ ID NO:131) and for LW1727 is: 5'-GCATCTCGAGTTATAG
GAGACCCATGGGCAGGG-3' (SEQ ID NO:132). The underlined portion of the primer matches the 5' and 3' areas, respectively, of the coding region. The ligation reaction used
solutions from the TOPO TA Cloning Kit (Invitrogen) which consisted of 4 μ l PCR product
DNA, 1 μ l of salt solution and 1 μ l pCRII-TOPO vector that was incubated for 5 minutes at
room temperature then the reaction was placed on ice. Two microliters of the ligation reaction
20 was transformed in One-Shot TOP10 cells (Invitrogen), and placed on ice for 30 minutes. The
cells were heat-shocked for 30 seconds at 42C, placed on ice for two minutes, 250 μ l of SOC
was added, then incubated at 37C with shaking for one hour and then plated onto ampicillin
plates. A single colony containing an insert was used to inoculate a 5 ml culture of LB medium.
Plasmid DNA was purified using a Concert Rapid Plasmid Miniprep System (GibcoBRL) and
25 then sequenced.

The mutation in SEQ-49 in the plasmid above was repaired using the QuikChange Site-Directed Mutagenesis Kit (Stratagene). The PCR reaction contained 40 μ l H₂O, 5 μ l 10x Reaction buffer, 1 μ l mini-prep DNA, 1 μ l LW1741 (125 ng/ μ l) (SEQ ID NO:133), 1 μ l LW1742 (125 ng/ μ l) (SEQ ID NO:134), 1 μ l 10 mM dNTP, 1 μ l Pfu DNA polymerase. The
30 cycle conditions were 95C for 30 seconds then 14 cycles at 95C for 30 seconds, 55C for 1 minute, 68C for 12 minutes. The tube was placed on ice for 2 minutes, then 1 μ l of DpnI was added and the tube incubated at 37C for one hour. One microliter of the DpnI-treated DNA was transformed into Epicurian coli XL1-Blue supercompetent cells and the entire insert was re-sequenced. The primer sequences are for LW1741: 5'-

GGTGCTCTGGAACCTCGGCTTCCGCATCAAGAAGGGCCCC-3' (SEQ ID NO:133) and for LW1742 was: 5'-GGGGCCCTTCTTGATGCGGAAGCCGAGGTTCCAGAGCACC-3' (SEQ ID NO:134).

nGPCR-61

5 The PCR reaction was performed in 50 μ l containing 34.5 μ l H₂O, 5 μ l Buffer II (PE Applied Biosystems AmpliTaq Gold system), 6 μ l 25 mM MgCl₂, 2 μ l 10 mM dNTP mix, 1.5 μ l human genomic DNA (Clontech #6550-1, 0.1 μ g/ μ l), 0.3 μ l primer VR61C (1 μ g/ μ l) (SEQ ID NO:135), 0.3 μ l primer VR61D (1 μ g/ μ l) (SEQ ID NO:136), and 0.4 μ l AmpliTaq Gold™ DNA Polymerase. The primer sequence for VR61C was: 5'-TTCAAAGCTTATGAACAAAC
10 AATACAACATGTATTCAAC-3' (SEQ ID NO:135), corresponding to the 5' end of the coding region and containing a HindIII restriction site, and the primer sequence for VR61D was: 5'-TTCACTCGAGTCAAACATATGATTGCATATGTG-3' (SEQ ID NO:136), corresponding to the 3' end of the coding region and containing an XhoI restriction site(Generis). The PCR reaction was carried out using a GeneAmp PCR9700 thermocycler (Perkin Elmer Applied
15 Biosystems) and started with 1 cycle of 95C for 10 minutes, then 14 cycles at 95C for 30 seconds, 72C for 2 minutes decreasing 1C each cycle, 72C for 1 minute, followed by 30 cycles at 95C for 30 seconds, 60C for 30 seconds, 72C for 1 minute. The PCR reaction was loaded on a 0.75% agarose gel. The DNA band was excised from the gel and the DNA was eluted from the agarose using a QIAquick gel extraction kit (Qiagen). The eluted DNA was ethanol-precipitated
20 and resuspended in 4 μ l H₂O for ligation. The ligation reaction consisted of 4 μ l of fresh ethanol-precipitated PCR product and 1 μ l of pCRII-TOPO vector (Invitrogen). The reaction was gently mixed and allowed to incubate for 5 minutes at room temperature followed by the addition of 1 μ l of 6x TOPO cloning stop solution and mixing for 10 seconds at room temperature. The sample was then placed on ice and 2 μ l was transformed in 50 μ l of One Shot cells (Invitrogen)
25 and plated onto ampicillin plates. Four white colonies were chosen and the presence of an insert was verified by PCR in the following manner. Each colony was resuspended in 50 μ L H₂O. A 16 μ l aliquot was removed and boiled for 5 minutes and the sample was placed on ice for 5 minutes. The sample was microfuged briefly to pellet any bacterial debris and PCR was carried out with 15 μ l sample using primers VR61C and VR61D, above.

30 Colonies from the positive clones identified by PCR were used to inoculate a 4 ml culture of LB medium containing 100 μ g/ml ampicillin. Plasmid DNA was purified using the Wizard Plus Minipreps DNA purification system (Promega). Since the primers used to PCR SEQ-61 from genomic DNA were engineered to have HindIII and XhoI sites, the cDNA obtained from the minipreps was digested with these restriction enzymes. Clones were verified

as having an insert of the correct size by gel electrophoresis. cDNA from one of the clones was then submitted for sequencing. Two mutations were found (bp 939 T→C and bp1004 G→T). The mutation at bp 939 was found to be a silent mutation and was not repaired. The mutation at bp 1004 was repaired as described below.

5 The mutation at bp 1004 in SEQ-61 was repaired using the QuikChange Site-Directed Mutagenesis Kit (Stratagene). The PCR reaction contained 38.1 µl H₂O, 5 µl 10x reaction buffer, 50 ng mini-prep cDNA from above, 1.25 µl primer VR61G (100 ng/µl) (SEQ ID NO:137), 1.25 µl primer VR61H (100 ng/µl) (SEQ ID NO:138), 2 µl 10 mM dNTP mix, and 1 µl Pfu DNA polymerase. The primer sequence for VR61G was: 5'-GCACATATGCAATCATA 10 TGGTTGACTCGAGTGAAAAGGG-3' (SEQ ID NO:137) and the primer sequence for VR61H was: 5'- CCCTTTCACTCGAGTCAACCATGATTGCATATGTGC-3' (SEQ ID NO:138), where the base highlighted and underlined is the one being corrected. The PCR cycle conditions were 95C for 30 seconds, then 12 cycles at 95C for 30 seconds, 55C for 1 minute, 68C for 11 minutes. One µl of DpnI was added and the reaction was incubated at 37C for 1 15 hour. One µl of the DpnI-treated DNA was transformed into 50 µl Epicurian coli XL1-Blue supercompetent cells and plated onto ampicillin plates. Two colonies were chosen and miniprep DNA was purified as described above. The presence of an insert was verified by restriction digest/gel electrophoresis. DNA from both of the samples was submitted for sequencing. Sequencing information showed that the mutation was repaired in both clones. One of the clones 20 was chosen for use in subsequent work.

The clone described above was sequenced directly using an ABI377 fluorescence-based sequencer (Perkin Elmer/Applied Biosystems Division, PE/ABD, Foster City, CA) and the ABI BigDyeTM Terminator Cycle Sequencing Ready Reaction kit with Taq FSTM polymerase. Each ABI cycle sequencing reaction contained 0.5 µg of plasmid DNA. Cycle-sequencing was 25 performed using an initial denaturation at 98C for 1 minute, followed by 50 cycles: 96C for 30 seconds, annealing at 50C for 30 seconds, and extension at 60C for 4 minutes. Temperature cycles and times were controlled by a Perkin-Elmer 9600 thermocycler. Extension products were purified using AGTC (R) gel filtration block (Edge Biosystems, Gaithersburg, MD). Each reaction product was loaded by pipette onto the column, which was then centrifuged in a 30 swinging bucket centrifuge (Sorvall model RT6000B tabletop centrifuge) at 1500 x g for 4 minutes at room temperature. Column-purified samples were dried under vacuum for about 40 minutes and then dissolved in 3 µl of a DNA loading solution (83% deionized formamide, 8.3 mM EDTA, and 1.6 mg/ml Blue Dextran). The samples were then heated to 90C for 3.5 minutes and loaded into the gel sample wells for sequence analysis by the ABI377 sequencer. Sequence

analysis was performed by importing ABI377 files into the Sequencer program (Gene Codes, Ann Arbor, MI).

nGPCR-51

The cDNA clone for nGPCR-51 was isolated by Life Technologies, Inc. using their
5 GENETRAPPER® cDNA Positive Selection System (Cat. No. 10356-020). Double-stranded
plasmid DNA from the SUPERSCRIPT Human Fetal Brain cDNA Library (Cat. No. 10662-
013) was used as a substrate. The library was made with fetal brain cDNA directionally cloned
into the *NotI-Sall* site of pCMV-SPORT2. An oligonucleotide probe 5'-CTCCATCTGTCTC
AGCTATGCCAGCAGCAG-3' (SEQ ID NO:139) was designed from Celera sequence
10 GA_11007426 and then biotinylated for solution hybridization to single-stranded fetal brain
DNA (prepared from total cDNA library) and selection with paramagnetic beads.

Preparation of dsDNA from a Plasmid cDNA Library

One hundred ml of Terrific Broth containing 100 mg/ml ampicillin was inoculated with
cells from an amplified library. The cells were grown to saturation at 30C. The sample was
15 centrifuged at 4,800 g for 15 minutes at 4C. The supernatant was decanted, and the cell pellet
was resuspended in a total volume of 10 ml of buffer I with RNase (15 mM Tris-HCl (pH 8.0),
10 mM EDTA, RNase A(100 µg/ml), RNase T1 (1,200 units/ml)). Ten ml of buffer II (0.2 M
NaOH, 1% SDS) was added to the resuspended cells, and the sample was inverted to mix and
allowed to incubate for 5 minutes at room temperature. Ten ml of cold 7.5 M NH₄OAc was
20 added to the cell mixture, and the sample was inverted to mix and allowed to incubate on ice for
10 minutes. The sample was centrifuged at 3,000 g (4,000 rpm in an HS4 rotor) for 15 minutes
at 4C. The supernatant was poured through cheesecloth into a fresh 50-ml centrifuge tube. An
equal volume of cold isopropanol (-20C) was added to the tube, mixed well, and the sample was
25 centrifuged at 3,000 g for 15 minutes at 4C. The supernatant was discarded, and the pellet was
resuspended in 1 ml of buffer I with RNase and transferred to a microcentrifuge tube. The
solution was clarified by centrifugation at 4C for 1 minute at 14,000 g. The supernatant was
transferred to a fresh microcentrifuge tube, and incubated at 37C for 30 minutes, after which the
tube was incubated at 65C for 5 minutes. The sample was split into two equal parts (~500 µl
30 each) in 1.5-ml microcentrifuge tubes. An equal volume of phenol: chloroform:isoamyl alcohol
(25:24:1) was added to each sample and the tube was vortexed. The sample was centrifuged at
room temperature for 5 minutes at 14,000 g. Four hundred fifty µl of the upper aqueous phase
was transferred to fresh microcentrifuge tubes. The phenol:chloroform extraction procedure
above was repeated 3 times. An equal volume of isopropanol (-20C) was added to each tube.
The tube was centrifuged at 4C for 15 minutes at 14,000 g, and the supernatant was discarded.

Five hundred μ l of 70% ethanol was carefully added to each tube. The tubes were centrifuged at 4C for 5 minutes at 14,000 g. The supernatant was discarded, and the pellet was dried at room temperature for 10 minutes. The pellets were resuspended in a total of 200 μ l TE buffer.

Generation of ssDNA

5 Five μ g ds phagemid cDNA (1 μ g/ μ l), 2 μ l 10X Gene II Buffer, and sufficient autoclaved, distilled water to bring the volume to 19 μ l was added to a 1.5 ml microcentrifuge tube at room temperature. To each sample, 1 μ l of Gene II (phage F1 endonuclease) was added. The sample was vortexed and centrifuged at room temperature for 2 s at 14,000 g to collect the contents to the bottom of the tube. The tubes were incubated in a 30C water bath for 25 minutes. The mixture was heated at 65C for 5 minutes and immediately chilled on ice for 1 minute. Two μ l of Exo III were added, and the tube was vortexed and centrifuged at room temperature for 2 s at 14,000 g. The tubes were incubated at 37C for 60 minutes. An equal volume (20 μ l) of phenol:chloroform:isoamyl alcohol (25:24:1) was added to each tube, vortexed thoroughly, and centrifuged at room temperature for 5 minutes at 14,000 g to separate the phases. Eighteen μ l of the upper aqueous phase was transferred to a fresh 1.5-ml

10 microcentrifuge tube.

15

Biotinylation Reaction

The following components were added to a 1.5 ml microcentrifuge tube:

5 μ l 5X TdT Buffer; 3 μ g oligonucleotide (SEQ ID NO:139), 5 μ l Biotin-14-dCTP, sufficient autoclaved, distilled water to bring the volume to 23 μ l, and 2 μ l TdT (Terminal 20 Deoxynucleotidyl Transferase). Each tube is vortexed gently and centrifuged for 2 s at 14,000 g. The tubes were incubated for 1 hour at 30C. After 1 hour, 1 μ l of Glycogen (20 μ g/ μ l), 26 μ l of 1 M Tris-HCl (pH 7.5), and 120 μ l of ethanol were added to the oligonucleotide biotinylation reaction. Each sample was vortexed and stored on dry ice for 10 minutes. Each tube was 25 centrifuged at 4C for 30 minutes at 14,000 g. The supernatant was removed from the microcentrifuge tubes, and 200 μ l of 70% ethanol (-20C) was layered over the pellet. Each tube was centrifuged at 4C for 2 minutes at 14,000 g. The ethanol wash was removed from the microcentrifuge tubes, and the ethanol wash was repeated once. The pellets were dried at room 30 temperature for 10 minutes or until completely dry. The biotinylated oligonucleotide was resuspended in 20 μ l TE buffer.

Oligonucleotide Hybridization

The biotinylated oligonucleotide was diluted to 20 ng/ μ l in TE buffer. The 4X Hybridization Buffer was incubated for 2 minutes at 37C, mixed well, and 6 μ l was added to the remaining 17 μ l of Gene II/Exo III-treated DNA. The DNA was denatured in a 95C water bath

for 1 minutes, and then chilled immediately on ice for 1 minute. One μ l of diluted biotinylated oligonucleotide (20 ng) was added to the denatured DNA. The tube was incubated at 37C in a water bath or incubator for 1 hour.

Streptavidin Paramagnetic Bead Preparation

5 The beads were gently mixed by pipetting until the beads at the bottom of the tube were completely resuspended. For each reaction, 45 μ l of the mixed beads were transferred to the bottom of a microcentrifuge tube. The tubes were placed in the magnet and allowed to sit for 2 minutes. With the tubes still in the magnet, the supernatant was removed by pipetting. One hundred μ l of TE buffer was immediately added to the beads. The tubes were removed from the 10 magnet and the beads were resuspended by finger-tapping or vortexing at the lowest setting. The tubes were reinserted into the magnet, and after 2 min, the supernatant was removed. The beads were resuspended in 30 μ l of TE buffer.

cDNA Capture

15 The hybridization mixture was removed from the 37C water bath and centrifuged at room temperature for 2 s at 14,000 g. The prepared paramagnetic beads were pipetted into the mixture and gently mixed by pipetting. The suspension was incubated for 30 minutes at room temperature. The suspension was gently mixed frequently to resuspend the beads. The tubes were inserted into the magnet, and after 2 min, the supernatant was removed and discarded. One hundred μ l of Wash Buffer was then added to the beads. The beads were resuspended and the 20 tubes reinserted into the magnet for 2 minutes. The supernatant was removed and discarded. The washing step was repeated one more time. One hundred μ l of Wash Buffer was added to the beads. The beads were gently resuspended by pipetting up and down (not by vortexing) and the solution was transferred to a new tube. The tubes were inserted into the magnet for 5 minutes. The supernatant was removed from the paramagnetic beads and discarded. One hundred μ l of 25 Wash Buffer was immediately added and the tubes mixed by finger tapping or gently vortexing. The tubes were placed into the magnet for 5 minutes. For each elution, 14 μ l of TE buffer (pH 8.0) was mixed with 7 μ l of the 3X Elution Buffer. After the 5 minute incubation, the supernatant was removed and discarded from the paramagnetic beads; 20 μ l of 1X elution buffer was added to the beads, and mixed well. The beads were incubated for 5 minutes at room 30 temperature. During the incubation, the beads were mixed for 10 seconds every minute. The tube was inserted into the magnet and allowed to sit for 5 minutes. The supernatant (containing the captured cDNA clone) was collected and transferred to a fresh tube, and the beads were resuspended in 15 μ l of TE buffer. The tube was inserted into the magnet and allowed to sit for 5 minutes. The supernatant was transferred from the tube and combined with the previous

supernatant. The tube containing the combined supernatants was inserted into the magnet for 10 minutes to remove any remaining paramagnetic beads and the supernatant was transferred to a fresh microcentrifuge tube. To the supernatant (~35 μ l), 1 μ l of Glycogen, 18 μ l of 7.5 M NH₄OAc, and 135 μ l of ethanol (~20C) were added. The tube was mixed well and stored on ice for 5 10 minutes. The tube was centrifuged at 4C for 30 minutes at 14,000 g. The supernatant was removed from the small pellets. One hundred μ l of 70% ethanol was added to each tube. The tubes were centrifuged at room temperature for 2 minutes at 14,000 g. The ethanol was removed and the pellets dried at room temperature until dry. The pellets were resuspended in 5 μ l of TE buffer and stored at 4C.

10 *Repair of Captured cDNA: Repair Reaction*

The thermal cycler is programmed for one cycle as described below:
90C denature step for 1 minute; 55C annealing step for 30 seconds; 70C extension step for 15 minutes. A DNA primer/repair mix was prepared for each capture reaction by adding the following to the captured cDNA: captured DNA (5 μ l), autoclaved, distilled water (11 μ l), 50 ng 15 oligonucleotide (SEQ ID NO:139; not biotinylated) (1 μ l), 10 mM dNTP Mix (0.5 μ l), 10X Repair Buffer (2 μ l), Repair Enzyme (0.5 μ l). The tubes were mixed and centrifuged at room temperature for 2 s at 14,000 g. The DNA primer/repair mix was incubated at 90C for 1 minute. The mix was transferred to 55C and incubated for 30 seconds. The mix was transferred to 70C and incubated for 15 minutes to allow primer extension. The mix was centrifuged at room 20 temperature for 2 s at 14,000 g. The repaired DNA was precipitated by adding 1 μ l Glycogen, 11 μ l 7.5 M ammonium acetate, and 90 μ l of ~20C ethanol to each tube. The tubes were vortexed and placed in ice for 10 minutes. The tubes were centrifuged at 4C for 30 minutes at 14,000 g. The ethanol was removed from the small pellet and 100 μ l of 70% ethanol (~20C) was layered on the pellet. The tubes were centrifuged at 4C for 2 minutes at 14,000 g. The ethanol 25 was removed and the pellets dried at room temperature until dry. The pellets were dissolved in 10 μ l of TE buffer.

Transformation with ULTRAMAX DH5a a-FT Cells

Competent cells were removed from -70C freezer and thawed on wet ice. The required number of 17 x 100 polypropylene tubes (Falcon 2059) were placed on ice. Immediately after 30 thawing, the cells were gently mixed, then 100 μ l of cells were aliquotted into chilled polypropylene tubes. Three μ l of the DNA were mixed into each individual tube of cells. The cells were incubated on ice for 30 minutes. Cells were heat-shocked for 45 seconds in a 42C water bath, and then placed on ice for 2 minutes. To each tube, 0.9 ml of room temperature S.O.C medium was added. The tubes were shaken at 225 rpm (37C) for 1 hour. For captured or

repaired cDNA samples, 100 μ l and 200 μ l aliquots were plated onto LB plates containing 100 μ g/ml ampicillin. The plates were incubated overnight in a 37C incubator.

Identification of Desired cDNA Clones by Colony PCR

A PCR master mix was prepared with the following components: 22.5 μ l/clone of 1X
5 PCR SUPERMIX, forward and reverse primers at a final concentration of 200 nM/primer, and
autoclaved, distilled water to a final volume of 25 μ l. Using a micropipette tip or sterile
toothpick, each colony was picked and placed into an individual tube containing master mix.
The PCR reaction was as follows: 1 cycle at 94C, 1 minute, then 30 cycles of 94C, 30 seconds;
55C, 30 seconds; 72C, 1 minute. The PCR primer pairs used in colony screening were, forward
10 primer: 5'-CCATCTGTCTCAGCTATGCC-3' (SEQ ID NO:140), and the reverse primer: 5'-
TCCTTCTCAGTCGCTTTC-3' (SEQ ID NO:141).

nGPCR-52

Two microliters of a human genomic library ($\sim 10^8$ PFU/ml) (Clontech) were added to 6
ml of an overnight culture of K802 cells (Clontech), then distributed as 250 μ l aliquots into each
15 of 24 tubes. The tubes were incubated at 37C for 15 minutes. Seven milliliters of 0.8% agarose
was added to each tube, mixed, then poured onto LB agar + 10 mM MgSO₄ plates and incubated
overnight at 37C. To each plate 5 ml of SM (0.1M NaCl, 8.1 TM MgSO₄·7H₂O, 50mM Tris-Cl
(pH 7.5), 0.0001% gelatin) phage buffer was added and the top agarose was removed with a
microscope slide and placed in a 50 ml centrifuge tube. A drop of chloroform was added and the
20 tube was place in a 37C shaker for 15 minutes, then centrifuged for 20 minutes at 4000 RPM
(Sorvall RT6000 table top centrifuge) and the supernatant stored at 4C as a stock solution.

The PCR reaction was performed in 20 μ l containing 8.8 μ l H₂O, 4 T1 5X Rapid-Load
Buffer (Origene), 2 μ l 10xPCR buffer II (Perkin-Elmer), 2 μ l 25 mM MgCl₂, 0.8 μ l 10 mM
dNTP, 0.12 μ l LW1632 (SEQ ID NO:142) (1 μ g/ μ l), 0.12 μ l LW1633 (SEQ ID NO:143) (1
25 μ g/ μ l), 0.2 μ l AmpliTaq Gold polymerase (Perkin Elmer) and 2 μ l of phage from each of the 24
tubes. The PCR reaction involved 1 cycle at 80C for 20 minutes, 95C for 10 minutes, then 22
cycles at 95C for 30 seconds, 72C for 4 minutes decreasing 1C each cycle, 68C for 2 minutes,
followed by 30 cycles at 95C for 30 seconds, 55C for 30 seconds, 68C for 60 seconds. The
reaction was loaded onto a 2 % agarose gel. From the tube that gave a PCR product of the
30 correct size, 5 μ l was used to set up five 1:10 dilutions that were plated onto LB agar + 10 mM
MgSO₄ plates and incubated overnight. A BA85 nitrocellulose filter (Schleicher & Schuell) was
placed on top of each plate for 1 hour. The filter was removed, placed phage side up in a petri
dish, and covered with 4 ml of SM for 15 minutes to elute the phage. One milliliter of SM was
removed from each plate and used to set up a PCR reaction as above. The plate of the lowest

dilution to give a PCR product was subdivided, filter-lifted and the PCR reaction was repeated. The series of dilutions and subdividing of the plate was continued until a single plaque was isolated that gave a positive PCR band. Once a single plaque was isolated, 10 μ l phage supernatant was added to 100 μ l SM and 200 μ l of K802 cells per plate with a total of 8 plates 5 set up. The plates were incubated overnight at 37C. The top agarose was removed by adding 8 ml of SM, then scrapping off the agarose with a microscope slide and collected in a centrifuge tube. To the tube, 3 drops of chloroform was added, vortexed, incubated at 37C for 15 minutes then centrifuged for 20 minutes at 4000 RPM (Sorvall RT6000 table top centrifuge) to recover the phage, which was used to isolate genomic phage DNA using the Qiagen Lambda Midi Kit. 10 The sequence for primer LW1632 was 5'-CCTCCACTTGTGCTTCATC-3' (SEQ ID NO:142) and for LW1633: 5'-AAAATCTATCAACACCCAGCC-3' (SEQ ID NO:143).

For subcloning the coding region of nGPCR-52, PCR was performed in a 50 μ l reaction containing 33 μ l H₂O, 5 μ l 10X TT buffer (140 mM Ammonium Sulfate, 0.1 % gelatin, 0.6 M Tris-tricine pH 8.4), 5 μ l 15 mM MgSO₄, 2 μ l 10 mM dNTP, 4 μ l genomic phage DNA (0.1 μ g/ μ l), 0.3 μ l LW1643 (SEQ ID NO:144) (1 μ g/ μ l), 0.3 μ l LW1644 (SEQ ID NO:145) (1 μ g/ μ l), 0.4 μ l High Fidelity Taq polymerase (Boehringer Mannheim). The PCR reaction was 15 started with 1 cycle of 94C for 2 minutes followed by 15 cycles at 94C for 30 seconds, 55C for 60 seconds, and 68C for 2 minutes. The PCR reaction was loaded onto a 2 % agarose gel. The DNA band was excised from the gel, placed in GenElute Agarose spin column (Supelco) and 20 spun for 10 minutes at maximum speed. The eluted DNA was EtOH precipitated and resuspended in 12 μ l H₂O for ligation. The PCR primer sequence for LW1643: 5'-GA TCAAGCTTACCATGACCAGCAATTTTCCC-3' (SEQ ID NO:144) and for LW1644 was: 5'-GATCCTCGAGCTTATTCTAAAAAAACTAATGG-3' (SEQ ID NO:145). The ligation 25 reaction used solutions from the TOPO TA Cloning Kit (Invitrogen) which consisted of 4 μ l PCR product DNA and 1 μ l pCRII-TOPO vector that was incubated for 5 minutes at room temperature. To the ligation reaction one microliter of 6X TOPO Cloning Stop Solution was added then the reaction was placed on ice. Two microliters of the ligation reaction was 30 transformed in One-Shot TOP10 cells (Invitrogen), and placed on ice for 30 minutes. The cells were heat-shocked for 30 seconds at 42C, placed on ice for two minutes, 250 μ l of SOC was added, then incubated at 37C with shaking for one hour and then plated onto ampicillin plates. A single colony containing an insert was used to inoculate a 5 ml culture of LB medium. Plasmid DNA was purified using a Concert Rapid Plasmid Miniprep System (GibcoBRL) and then sequenced.

nGPCR-52 genomic phage DNA was sequenced using the ABI PRISM™ 310 Genetic Analyzer (PE Applied Biosystems) which uses advanced capillary electrophoresis technology and the ABI PRISM™ BigDye™ Terminator Cycle Sequencing Ready Reaction Kit. The cycle-sequencing reaction contained 14 μ l of H₂O, 16 μ l of BigDye Terminator mix, 7 μ l 5 genomic phage DNA (0.1 μ g/ μ l), and 3 μ l primer (25 ng/ μ l). The reaction was performed in a Perkin-Elmer 9600 thermocycler at 95C for 5 minutes, followed by 99 cycles of 95C for 30 seconds, 55C for 20 seconds, and 60C for 4 minutes. The product was purified using a Centriflex™ gel filtration cartridges, dried under vacuum, then dissolved in 16 μ l of Template Suppression Reagent. The samples were heated at 95C for 5 minutes then placed in the 310 10 Genetic Analyzer

Example 3: Subcloning of the Coding Region of nGPCR-x Via PCR

Additional experiments may be conducted to subclone the coding region of nGPCR and place the isolated coding region into a useful vector. Two additional PCR primers are designed based on the coding region of nGPCR, corresponding to either end. To protect against 15 exonucleolytic attack during subsequent exposure to enzymes, *e.g.*, Taq polymerase, primers are routinely synthesized with a protective run of nucleotides at the 5' end that were not necessarily complementary to the desired target.

PCR is performed in a 50 μ l reaction containing 34 μ l H₂O, 5 μ l 10X TT buffer (140 mM 20 ammonium sulfate, 0.1% gelatin, 0.6 M Tris-tricine, pH 8.4), 5 μ l 15 mM MgSO₄, 2 μ l dNTP mixture (dGTP, dATP, dTTP, and dCTP, each at 10 mM), 3 μ l genomic phage DNA (0.25 μ g/ μ l), 0.3 μ l Primer 1 (1 μ g/ μ l), 0.3 μ l Primer 2 (1 μ g/ μ l), 0.4 μ l High Fidelity Taq polymerase (Boehringer Mannheim). The PCR reaction was started with 1 cycle of 94C for 2 minutes; followed by 25 cycles at 94C for 30 seconds, 55C for 30 seconds, and 72C for 1.3 minutes.

The contents from the PCR reaction are loaded onto a 2% agarose gel and fractionated. 25 The DNA band of expected size is excised from the gel, placed in a GenElute Agarose spin column (Supelco) and spun for 10 minutes at maximum speed in a microfuge. The eluted DNA is precipitated with ethanol and resuspended in 6 μ l H₂O for ligation.

The PCR-amplified DNA fragment containing the coding region is cloned into pCR2.1 using a protocol standard in the art. In particular, the ligation reaction consists of 6 μ l of GPCR 30 DNA, 1 μ l 10X ligation buffer, 2 μ l pCR2.1 (25 ng/ μ l, Invitrogen), and 1 μ l T4 DNA ligase (Invitrogen). The reaction mixture is incubated overnight at 14C and the reaction is then stopped by heating at 65C for 10 minutes. Two microliters of the ligation reaction are transformed into One Shot cells (Invitrogen) and plated onto ampicillin plates. A single colony containing a recombinant pCR2.1 bearing an insert is used to inoculate a 5 ml culture of LB medium.

Plasmid DNA is purified using the Concert Rapid Plasmid Miniprep System (GibcoBRL) and sequenced. Following confirmation of the sequence, a 50 ml culture of LB medium is inoculated with the transformed One Shot cells, cultured, and processed using a Qiagen Plasmid Midi Kit to yield purified pCR-GPCR.

5 *nGPCR-70*

Colonies from the positive clones identified by PCR were used to inoculate a 4 ml culture of LB medium containing 100 µg/ml ampicillin. Plasmid DNA was purified using the Wizard Plus Minipreps DNA purification system (Promega). Since the primers used to PCR SEQ-70 from human brain cDNA were engineered to have HindIII and XhoI sites, the DNA 10 obtained from the minipreps was digested with these restriction enzymes. Five of the clones were verified by gel electrophoresis to give a DNA band of the correct size. cDNA from each of these clones was then submitted for sequencing. Each of the clones was found to have two or more mutations. The clone containing the fewest mutations (clone #7-2 mutations at bp 561 G→A and bp 1093 G→A) was repaired as described as below.

15 The mutations in SEQ-70 were repaired sequentially using the QuikChange Site-Directed Mutagenesis Kit (Stratagene). The mutation at bp 561 was repaired first and the primer sequences used were: VR70G 5'-CATGATCTGGGGGGCCAGCCCCAGC-3' (SEQ ID NO:146) and VR70H 5'-GCTGGGGCTGGCCCCCAGATCATG-3' (SEQ ID NO:147) where the base highlighted and underlined is the one being corrected. The PCR reaction contained 38.4 20 µl H₂O, 5 µl 10x reaction buffer, 50 ng mini-prep cDNA from clone #7, 2 µl 10 mM dNTP mix, and 1 µl Pfu DNA polymerase. The cycle conditions were 95C for 30 seconds, then 12 cycles at 95C for 30 seconds, 55C for 1 minute, 68C for 11 minutes. One µl of DpnI was added and the reaction was incubated at 37C for 1 hour. One µl of the DpnI-treated DNA was transformed into 50 µl Epicurian coli XL1-Blue supercompetent cells and plated onto ampicillin 25 plates. Four colonies were chosen and miniprep DNA was purified as described above. DNA from one of the preps was used as a template to repair the second mutation at bp 1093. The primer sequences used were: VR70I 5'-GACATCAATTCAGTGAGGGATGACGTCGAGGC AG-3' (SEQ ID NO:148) and VR70J 5'-CTGCCTCGACGTCATCCTCACTGAAATTGATG TC-3' (SEQ ID NO:149), where the base highlighted and underlined is the one being corrected. 30 The PCR and transformation reactions were carried out as described above. Mini-prep DNA was prepared from three colonies, and the presence of an insert was verified by restriction digest/gel electrophoresis. DNA from one of the samples was submitted for sequencing. Sequencing information showed that both mutations were repaired but that an additional mutation at bp 560 (G→A) had been introduced. This mutation was adjacent to the original mutation at bp 561 and

was encompassed in the sequence of the primer that was used. Since it could not be ruled out that there was an error in the synthesis of the primers, the same exact sequence corresponding to primers VR70G (SEQ ID NO:146) and VR70H (SEQ ID NO:147) was reordered and repair of the mutation with the new primers was carried out as above. DNA from 2 clones was submitted 5 for sequencing. One of the clones had no mutations and was used for subsequent work.

The clone described above was sequenced directly using an ABI377 fluorescence-based sequencer (Perkin Elmer/Applied Biosystems Division, PE/ABD, Foster City, CA) and the ABI BigDyeTM Terminator Cycle Sequencing Ready Reaction kit with Taq FSTM polymerase. Each ABI cycle sequencing reaction contained 0.5 µg of plasmid DNA. Cycle-sequencing was 10 performed using an initial denaturation at 98C for 1 minute, followed by 50 cycles: 96C for 30 seconds, annealing at 50C for 30 seconds, and extension at 60C for 4 minutes. Temperature cycles and times were controlled by a Perkin-Elmer 9600 thermocycler. Extension products were purified using AGTC (R) gel filtration block (Edge Biosystems, Gaithersburg, MD). Each reaction product was loaded by pipette onto the column, which was then centrifuged in a 15 swinging bucket centrifuge (Sorvall model RT6000B tabletop centrifuge) at 1500 x g for 4 minutes at room temperature. Column-purified samples were dried under vacuum for about 40 minutes and then dissolved in 3 µl of a DNA loading solution (83% deionized formamide, 8.3 mM EDTA, and 1.6 mg/ml Blue Dextran). The samples were then heated to 90C for 3.5 minutes and loaded into the gel sample wells for sequence analysis by the ABI377 sequencer. Sequence 20 analysis was performed by importing ABI377 files into the Sequencer program (Gene Codes, Ann Arbor, MI).

nGPCR-42

The DNA subcloned into pCRII was sequenced using the ABI PRISM™ 310 Genetic Analyzer (PE Applied Biosystems) which uses advanced capillary electrophoresis technology 25 and the ABI PRISM™ BigDye™ Terminator Cycle Sequencing Ready Reaction Kit. Each cycle-sequencing reaction contained 6 µl of H₂O, 8 µl of BigDye Terminator mix, 5 µl mini-prep DNA (0.1 µg/µl), and 1 µl primer (25 ng/µl) and was performed in a Perkin-Elmer 9600 thermocycler with 25 cycles of 96C for 10 seconds, 50C for 10 seconds, and 60C for 4 minutes. The product was purified using a Centriflex™ gel filtration cartridge, dried under vacuum, then 30 dissolved in 16 µl of Template Suppression Reagent (PE Applied Biosystems). The samples were heated at 95C for 5 minutes then placed in the 310 Genetic Analyzer.

nGPCR-46

The DNA subcloned into pCRII was sequenced using the ABI PRISM™ 310 Genetic Analyzer (PE Applied Biosystems) which uses advanced capillary electrophoresis technology

and the ABI PRISM™ BigDye™ Terminator Cycle Sequencing Ready Reaction Kit. Each cycle-sequencing reaction contained 6 μ l of H₂O, 8 μ l of BigDye Terminator mix, 5 μ l mini-prep DNA (0.1 μ g/ μ l), and 1 μ l primer (25 ng/ μ l) and was performed in a Perkin-Elmer 9600 thermocycler with 25 cycles of 96C for 10 seconds, 50C for 10 seconds, and 60C for 4 minutes.

5 The product was purified using a Centriflex™ gel filtration cartridge, dried under vacuum, then dissolved in 16 μ l of Template Suppression Reagent (PE Applied Biosystems). The samples were heated at 95C for 5 minutes then placed in the 310 Genetic Analyzer.

The mutation found in the clone was repaired using the QuikChange Site-Directed Mutagenesis Kit (Stratagene). The PCR reaction contained 39 μ l H₂O, 5 μ l 10xReaction buffer, 10 1 μ l mini-prep DNA (150 ng/ μ l), 2 μ l 10 mM dNTP, 1 μ l Pfu DNA polymerase, 1 μ l LW1638 (SEQ ID NO:150) (125 ng/ μ l), and 1 μ l LW1639 (SEQ ID NO:151) (125 ng/ μ l). The cycle conditions were 95C for 30 seconds then 12 cycles at 95C for 30 seconds, 55C for 1 minute, 68C for 12 minutes. The tube was placed on ice for 2 minutes, then 1 μ l of DpnI was added and the tube incubated at 37C for one hour. Two microliters of the *DpnI*-treated DNA was 15 transformed into Epicurian coli XL1-Blue supercompetent cells and the entire insert was re-sequenced. The forward PCR primer sequence was: LW1638 5'-CGAACTCCGCACTCCTGG CCAGGGCCCTGCGGGC-3' (SEQ ID NO:150) and the reverse PCR primer sequence was: LW1639 5'-GCCCGCAGGGCCCTGGCCAGGAGTGCAGGAGTTC-3' (SEQ ID NO:151).

nGPCR-48

20 The DNA subcloned into pCRII was sequenced using the ABI PRISM™ 310 Genetic Analyzer (PE Applied Biosystems) which uses advanced capillary electrophoresis technology and the ABI PRISM™ BigDye™ Terminator Cycle Sequencing Ready Reaction Kit. Each cycle-sequencing reaction contained 6 μ l of H₂O, 8 μ l of BigDye Terminator mix, 5 μ l mini-prep DNA (0.1 μ g/ μ l), and 1 μ l primer (25 ng/ μ l) and was performed in a Perkin-Elmer 9600 thermocycler with 25 cycles of 96C for 10 seconds, 50C for 10 seconds, and 60C for 4 minutes. 25 The product was purified using a Centriflex™ gel filtration cartridge, dried under vacuum, then dissolved in 16 μ l of Template Suppression Reagent (PE Applied Biosystems). The samples were heated at 95C for 5 minutes then placed in the 310 Genetic Analyzer.

nGPCR-49

30 The DNA subcloned into pCRII was sequenced using the ABI PRISM™ 310 Genetic Analyzer (PE Applied Biosystems) which uses advanced capillary electrophoresis technology and the ABI PRISM™ BigDye™ Terminator Cycle Sequencing Ready Reaction Kit. Each cycle-sequencing reaction contained 6 μ l of H₂O, 8 μ l of BigDye Terminator mix, 5 μ l mini-prep DNA (0.1 μ g/ μ l), and 1 μ l primer (25 ng/ μ l) (SEQ ID NO:131 and SEQ ID NO:132) and was

performed in a Perkin-Elmer 9600 thermocycler with 25 cycles of 96C for 10 seconds, 50C for 10 seconds, and 60C for 4 minutes. The product was purified using a Centriflex™ gel filtration cartridge, dried under vacuum, then dissolved in 16 µl of Template Suppression Reagent (PE Applied Biosystems). The samples were heated at 95C for 5 minutes then placed in the 310

5 Genetic Analyzer.

nGPCR-52

The DNA subcloned into pCRII was sequenced using the ABI PRISM™ 310 Genetic Analyzer (PE Applied Biosystems) which uses advanced capillary electrophoresis technology and the ABI PRISM™ BigDye™ Terminator Cycle Sequencing Ready Reaction Kit. Each 10 cycle-sequencing reaction contained 6 µl of H₂O, 8 µl of BigDye Terminator mix, 5 µl mini-prep DNA (0.1 µg/µl), and 1 µl primer (25 ng/µl) and was performed in a Perkin-Elmer 9600 thermocycler with 25 cycles of 96C for 10 seconds, 50C for 10 seconds, and 60C for 4 minutes. The product was purified using a Centriflex™ gel filtration cartridge, dried under vacuum, then dissolved in 16 µl of Template Suppression Reagent (PE Applied Biosystems). The samples 15 were heated at 95C for 5 minutes then placed in the 310 Genetic Analyzer.

Example 4: Hybridization Analysis to Demonstrate nGPCR-X Expression in Brain

The expression of nGPCR-x in mammals, such as the rat, may be investigated by *in situ* hybridization histochemistry. To investigate expression in the brain, for example, coronal and sagittal rat brain cryosections (20 µm thick) are prepared using a Reichert-Jung cryostat.

20 Individual sections are thaw-mounted onto silanized, nuclease-free slides (CEL Associates, Inc., Houston, TX), and stored at -80C. Sections are processed starting with post-fixation in cold 4% paraformaldehyde, rinsed in cold phosphate-buffered saline (PBS), acetylated using acetic anhydride in triethanolamine buffer, and dehydrated through a series of alcohol washes in 70%, 95%, and 100% alcohol at room temperature. Subsequently, sections are delipidated in 25 chloroform, followed by rehydration through successive exposure to 100% and 95% alcohol at room temperature. Microscope slides containing processed cryosections are allowed to air dry prior to hybridization. Other tissues may be assayed in a similar fashion.

A nGPCR-x-specific probe is generated using PCR. Following PCR amplification, the 30 fragment is digested with restriction enzymes and cloned into pBluescript II cleaved with the same enzymes. For production of a probe specific for the sense strand of nGPCR-x, the nGPCR-x clone in pBluescript II is linearized with a suitable restriction enzyme, which provides a substrate for labeled run-off transcripts (*i.e.*, cRNA riboprobes) using the vector-borne T7 promoter and commercially available T7 RNA polymerase. A probe specific for the antisense strand of nGPCR-x is also readily prepared using the nGPCR-x clone in pBluescript II by

cleaving the recombinant plasmid with a suitable restriction enzyme to generate a linearized substrate for the production of labeled run-off cRNA transcripts using the T3 promoter and cognate polymerase. The riboprobes are labeled with [³⁵S]-UTP to yield a specific activity of about 0.40×10^6 cpm/pmol for antisense riboprobes and about 0.65×10^6 cpm/pmol for sense-
5 strand riboprobes. Each riboprobe is subsequently denatured and added (2 pmol/ml) to hybridization buffer which contained 50% formamide, 10% dextran, 0.3 M NaCl, 10 mM Tris (pH 8.0), 1 mM EDTA, 1X Denhardt's Solution, and 10 mM dithiothreitol. Microscope slides containing sequential brain cryosections are independently exposed to 45 μ l of hybridization solution per slide and silanized cover slips are placed over the sections being exposed to
10 hybridization solution. Sections are incubated overnight (15-18 hours) at 52C to allow hybridization to occur. Equivalent series of cryosections are exposed to sense or antisense nGPCR-x-specific cRNA riboprobes.

Following the hybridization period, coverslips are washed off the slides in 1X SSC, followed by RNase A treatment involving the exposure of slides to 20 μ g/ml RNase A in a
15 buffer containing 10 mM Tris-HCl (pH 7.4), 0.5 M EDTA, and 0.5 M NaCl for 45 minutes at 37C. The cryosections are then subjected to three high-stringency washes in 0.1 X SSC at 52C for 20 minutes each. Following the series of washes, cryosections are dehydrated by consecutive exposure to 70%, 95%, and 100% ammonium acetate in alcohol, followed by air drying and exposure to Kodak BioMax™ MR-1 film. After 13 days of exposure, the film is developed.
20 Based on these results, slides containing tissue that hybridized, as shown by film autoradiograms, are coated with Kodak NTB-2 nuclear track emulsion and the slides are stored in the dark for 32 days. The slides are then developed and counterstained with hematoxylin. Emulsion-coated sections are analyzed microscopically to determine the specificity of labeling. The signal is determined to be specific if autoradiographic grains (generated by antisense probe
25 hybridization) are clearly associated with cresyl violet-stained cell bodies. Autoradiographic grains found between cell bodies indicates non-specific binding of the probe.

Expression of nGPCR-x in the brain provides an indication that modulators of nGPCR-x activity have utility for treating neurological disorders, including but not limited to, mental disorder, affective disorders, ADHD/ADD (*i.e.*, Attention Deficit-Hyperactivity
30 Disorder/Attention Deficit Disorder), and neural disorders such as Alzheimer's disease, Parkinson's disease, migraine, and senile dementia. Some other diseases for which modulators of nGPCR-x may have utility include depression, anxiety, bipolar disease, epilepsy, neuritis, neurasthenia, neuropathy, neuroses, and the like. Use of nGPCR-x modulators, including

nGPCR-x ligands and anti-nGPCR-x antibodies, to treat individuals having such disease states is intended as an aspect of the invention.

In Situ Hybridization of SEQ-51

Coronal and sagittal oriented rat brain sections were cryosectioned (20 μ m thick) using a 5 Leica CM3050 cryostat. The individual sections were thaw-mounted onto silanated, nuclease-free slides (CEL Associates, Inc., Houston, TX), and stored at -80C. The sections were processed starting with post-fixation in cold 4% paraformaldehyde, rinsed in cold PBS, acetylated using acetic anhydride in triethanolamine buffer and dehydrated through 70%, 95%, and 100% alcohols at room temperature (RT). This was followed with delipidation in 10 chloroform then rehydration in 100% and 95% alcohol at RT. Sections were allowed to air dry prior to hybridization. For nGPCR-51, a 250 bp PCR fragment (spanning nt 83 to 332) was generated that contained T7 polymerase on either the 5' end (sense) with primers LW1743 5'-GCGTAATACGACTCACTATAGGGAGACCTGCCAGTGTGGTAGATACAG-3' (SEQ ID NO:152) and LW1746 5'-GGATGTGATGATGGTGCAG-3' (SEQ ID NO:153) or the 3' end 15 (antisense) with primers LW1744 5'-GCGTAATACGACTCACTATAGGGAGACCGGATG TGATGATGGTGCAG-3' (SEQ ID NO:154) and LW1745 5'-TGCCAGTGTGGTAGATAC AG-3' (SEQ ID NO:155). nGPCR-51 was labeled with 35 S-UTP to yield a specific activity of 0.655×10^6 cpm/pmol for antisense and 0.675×10^6 cpm/pmol for sense probe. Both riboprobes were denatured and added to hybridization buffer which contained 50% formamide, 10% 20 dextran, 0.3M NaCl, 10 mM Tris, 1 mM EDTA, 1X Denhardts, and 10 mM DTT. Sequential brain cryosections were hybridized with 45 μ l/slide of the sense and antisense riboprobe hybridization mixture then coverslipped with silanized glass coverslips. The sections were hybridized overnight (15-18 hours) at 50C in an incubator.

Coverslips were washed off the slides in 1X SSC, followed by RNase A treatment, and 25 high temperature stringency washes (3X, 20 minutes at 49C) in 0.1X SSC. Slides were dehydrated with 70%, 95% and 100% NH₄OAc alcohols, air dried and exposed to Kodak BioMax MR-1 film. After 8 days of exposure, the film was developed. This was followed with coating selected tissue slides with Kodak NTB-2 nuclear track emulsion and storing the slides in the dark for 16 days. The slides were then developed and counterstained with hematoxylin. 30 Emulsion-coated sections were analyzed microscopically to determine the specificity of labeling. Presence of autoradiographic grains (generated by antisense probe hybridization) over cell bodies (versus between cell bodies) was used as an index of specific hybridization.

Sense and antisense 35 S-labeled RNA probes were generated using a 250-bp fragment of nGPCR-51 for *in situ* hybridization histochemistry. Specific labeling with the antisense probe

showed wide spread distribution of nGPCR-51 mRNA. Localization appears in the piriform ctx, habenula, bed nucleus of stria terminalis, islands of Calleja, olfactory tubercle, hippocampus, hypothalamus, PVN, red nucleus, interpeduncular nucleus, dorsal raphe, substantia nigra pars compacta, and reticular thalamus. Expression of nGPCR-51 in these brain regions provided an
5 indication that modulators of nGPCR-51 activity have utility for treating disorders, including but not limited to, schizophrenia, major depression, bipolar disease, anxiety disorder, Parkinson's disease, endocrine disorders, Alzheimer's disease and the like.

In Situ Hybridization SEQ-52

Coronal and sagittal oriented rat brain sections were cryosectioned (20 μ m thick) using a
10 Leica CM3050 cryostat. The individual sections were thaw-mounted onto silanated, nuclease-free slides (CEL Associates, Inc., Houston, TX), and stored at -80C. The sections were processed starting with post-fixation in cold 4% paraformaldehyde, rinsed in cold PBS, acetylated using acetic anhydride in triethanolamine buffer and dehydrated through 70%, 95%, and 100% alcohols at room temperature. This was followed with delipidation in chloroform then
15 rehydration in 100% and 95% alcohol at room temperature. Sections were allowed to air dry prior to hybridization. For nGPCR-52, a 292 bp PCR fragment (spanning nt 2920 to 3211) was generated that contained T7 polymerase on either the 5' end (sense) with primers LW1682 5'-GCGTAATACGACTCACTATAGGGAGACCACCAGCAATTTCCCAACC-3' (SEQ ID NO:156) and LW1685 5'-AATACCAGCAGCTCTCCAC-3' (SEQ ID NO:157) or the 3' end
20 (antisense) with primers LW1683 5'-GCGTAATACGACTCACTATAGGGAGACCAATACC AGCAGCTCTCCAC-3' (SEQ ID NO:158) and LW1784 5'-ACCAGCAATTTCCCAACC-3' (SEQ ID NO:159). nGPCR-52 was labeled with 35 S-UTP to yield a specific activity of 0.686 $\times 10^6$ cpm/pmol for the antisense probe and 0.601 $\times 10^6$ cpm/pmol for the sense probe. Both
riboprobes were denatured and added to hybridization buffer which contained 50% formamide,
25 10% dextran, 0.3M NaCl, 10 mM Tris, 1 mM EDTA, 1X Denhardts, and 10 mM DTT. Sequential brain cryosections were hybridized with 45 Tl/slide of the sense and antisense riboprobe hybridization mixture then coverslipped with silanized glass coverslips. The sections were hybridized overnight (15-18 hours) at 49C in an incubator.

Coverslips were washed off the slides in 1X SSC, followed by RNase A treatment, and
30 high temperature stringency washes (3X, 20 minutes at 48C) in 0.1X SSC. Slides were dehydrated with 70%, 95% and 100% NH₄OAc alcohols, air dried and exposed to Kodak BioMax MR-1 film. After 8 days of exposure, the film was developed. This was followed with coating selected tissue slides with Kodak NTB-2 nuclear track emulsion and storing the slides in the dark for 16 days. The slides were then developed and counterstained with hematoxylin.

Emulsion-coated sections were analyzed microscopically to determine the specificity of labeling. Presence of autoradiographic grains (generated by antisense probe hybridization) over cell bodies (versus between cell bodies) was used as an index of specific hybridization.

To determine which regions of the brain nGPCR-52 is expressed in, a 292 bp fragment 5 of nGPCR-52 was used to generate sense and antisense ^{35}S -labeled RNA probes for *in situ* hybridization histochemistry. Specific labeling with the antisense probe showed wide spread distribution of nGPCR-52 mRNA. Localization appears in the cortex, piriform ctx, habenula, islands of Calleja, hippocampus, hypothalamus, red nucleus, dorsal raphe, substantia nigra pars compacta, and reticular thalamus. The regions where nGPCR-52 is expressed are within the 10 limbic and neuroendocrine circuitry of the brain.

Example 5: Tissue Expression Profiling

A PCR-based system (RapidScan™ Gene Expression Panel, OriGene Technologies, Rockville, MD) may be used to generate a comprehensive expression profile of the putative nGPCR-x in human tissue, and in human brain regions. The RapidScan Expression Panel is 15 comprised of first-strand cDNAs from various human tissues and brain regions that are serially diluted over a 4-log range and arrayed into a multi-well PCR plate. Human tissues in the array may include: brain, heart, kidney, spleen, liver, colon, lung, small intestine, muscle, stomach, testis, placenta, salivary gland, thyroid, adrenal gland, pancreas, ovary, uterus, prostate, skin, PBL, bone marrow, fetal brain, and fetal liver.

20 Expression of nGPCR-x in various tissues is detected using PCR primers designed based on the available sequence of the receptor that will prime the synthesis of a predetermined size fragment in the presence of the appropriate cDNA.

PCR is performed in a 50 μl reaction containing 34 μl H_2O , 5 μl 10X TT buffer (140 mM ammonium sulfate, 0.1% gelatin, 0.6 M Tris-tricine, pH 8.4), 5 μl 15 mM MgSO_4 , 2 μl dNTP 25 mixture (dGTP, dATP, dTTP, and dCTP, each at 10 mM), 0.3 μl forward primer (1 $\mu\text{g}/\mu\text{l}$), 0.3 μl reverse primer (1 $\mu\text{g}/\mu\text{l}$), 0.4 μl High Fidelity Taq polymerase (Boehringer Mannheim). The PCR reaction mixture is added to each well of the PCR plate. The plate is placed in a MJ Research PTC100 thermocycler, and is then exposed to the following cycling parameters: Pre-soak 94C for 3 minutes; denaturation at 94C for 30 seconds; annealing at primer 57C for 45 30 seconds; extension 72C for 2 minutes; for 35 cycles. PCR productions are then separated and analyzed by electrophoresis on a 1.2% agarose gel stained with ethidium bromide. The 4-log dilution range of cDNA deposited on the plate ensures that the amplification reaction is within the linear range and, hence, facilitates semi-quantitative determination of relative mRNA accumulation in the various tissues or brain regions examined.

nGPCR-70

Tissue specific expression of the cDNA encoding nGPCR-70 was detected using a PCR-based method. Multiple Choice™ first strand cDNAs (OriGene Technologies, Rockville, MD) from 6 human tissues were serially diluted over a 3-log range and arrayed into a multi-well PCR plate. This array was used to generate a comprehensive expression profile of the putative GPCR in human tissues. Human tissues arrayed included: brain, heart, kidney, peripheral blood leukocytes, lung and testis. PCR primers were designed based on the available sequence of the Celera sequence GA_16417344. The forward primer used was: 5'-GTGACTAACTCTGCCT GCG-3' (SEQ ID NO:160). The reverse primer used was: 5'-TTGCGCTGCAACACTAGCG-3' (SEQ ID NO:161). This primer set primed the synthesis of a 286 base pair fragment in the presence of the appropriate cDNA. For detection of expression within brain regions, the same primer set was used with the Human Brain Rapid Scan™ Panel (OriGene Technologies, Rockville, MD). This panel represented serial dilutions over a 2 log range of first strand cDNA from the following brain regions arrayed in a 96 well format: frontal lobe, temporal lobe, cerebellum, hippocampus, substantia nigra, caudate nucleus, amygdala, thalamus, hypothalamus, pons, medulla and spinal cord. Primers were synthesized by Genosys Corp., The Woodlands, TX. PCR reactions were assembled using the components of the Expand Hi-Fi PCR System™ (Roche Molecular Biochemicals, Indianapolis, IN). Twenty-five microliters of the PCR reaction mixture was added to each well of the RapidScan PCR plate. The plate was placed in a GeneAmp 9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The following cycling program was executed: Pre-soak at (94C for 3 minutes) followed by 35 cycles of 94C for 45 seconds, 53C for 2 minutes and 72C for 45 seconds. PCR reaction products were then separated and analyzed by electrophoresis on a 2.0% agarose gel stained with ethidium bromide.

nGPCR-70 was expressed in the brain, heart and lung. Within the brain, nGPCR-70 was expressed in regions including but not limited to, cerebellum, hippocampus, substantia nigra, thalamus, frontal lobe, caudate nucleus, and spinal cord. Expression of the nGPCR-70 in the brain provides an indication that modulators of nGPCR-70 activity have utility for treating neurological disorders, including but not limited to, movement disorders, affective disorders, metabolic disorders, inflammatory disorders and cancers. Use of nGPCR-70 modulators, including nGPCR-70 ligands and anti-nGPCR-70 antibodies, to treat individuals having such disease states is intended as an aspect of the invention.

nGPCR-63

Tissue specific expression of the putative nGPCR-63 was detected using a PCR-based RapidScanTM Gene Expression Panel (OriGene Technologies, Rockville, MD). The RapidScan Expression Panel is comprised of first-strand cDNAs from 12 human tissues that are serially diluted over a 3-log range and arrayed into a multi-well PCR plate. This array was used to 5 generate a comprehensive expression profile of the putative GPCR in human tissues. Human tissues arrayed included: brain, heart, kidney, spleen, liver, colon, lung, small intestine, muscle, stomach, testis, placenta, salivary gland, thyroid, adrenal gland, pancreas, ovary, uterus, prostate, skin, PBL, bone marrow, fetal brain, fetal liver. PCR primers were designed based on the available sequence of the Celera sequence HUM_IDS|Contig|11000258115466. The 10 forward primer used was: 5'-ACAGCCCCAAAGCCAAACAC-3' (SEQ ID NO:162). The reverse primer was: 5'-CCGCAGGAGCAATGAAAATCAG-3' (SEQ ID NO:163). This primer set primed the synthesis of a 220 base pair fragment in the presence of the appropriate cDNA. For detection of expression within brain regions, the same primer set was used with the Human 15 Brain RapidScanTM Panel (OriGene Technologies, Rockville, MD). This panel represents serial dilutions over a 2 log range of first strand cDNA from the following brain regions arrayed in a 96 well format: frontal lobe, temporal lobe, cerebellum, hippocampus, substantia nigra, caudate nucleus, amygdala, thalamus, hypothalamus, pons, medulla and spinal cord. Primers were synthesized by Genosys Corp., The Woodlands, TX. PCR reactions were assembled using the components of the Expand Hi-Fi PCR SystemTM (Roche Molecular Biochemicals, Indianapolis, 20 IN). Twenty-five microliters of the PCR reaction mixture was added to each well of the RapidScan PCR plate. The plate was placed in a GeneAmp 9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The following cycling program was executed: Pre-soak at (94 for 3minutes) followed by 35 cycles of 94C for 45 seconds, 54C for 2 minutes, 72C for 45 seconds. PCR reaction products were then separated and analyzed by electrophoresis on a 2.0% agarose 25 gel, stained with ethidium bromide.

nGPCR-63 was expressed in the brain, heart, kidney, liver, muscle, ovary, prostate, small intestine, spleen, testis, peripheral blood leukocytes, and lung. Within the brain, nGPCR-63 was expressed in regions including but not limited to, cerebellum, amygdala, hypothalamus, medulla, temporal lobe, pons, hippocampus, substantia nigra, thalamus, frontal lobe, caudate 30 nucleus, and spinal cord. Expression of the nGPCR-63 in the brain provides an indication that modulators of nGPCR-63 activity have utility for treating neurological disorders, including but not limited to, movement disorders, affective disorders, metabolic disorders, inflammatory disorders and cancers. Use of nGPCR-63 modulators, including nGPCR-63 ligands and anti-

nGPCR-63 antibodies, to treat individuals having such disease states is intended as an aspect of the invention.

nGPCR-42

Expression of nGPCR-42 in the various tissues was detected by using PCR primers 5 designed based on the sequence of the receptor that prime the synthesis of a 110 bp fragment in the presence of the appropriate cDNA. The forward primer used to detect expression of nGPCR-42 was: 5'-TCTCCAAACTCCTGGCCTTC-3' (SEQ ID NO:164) and the reverse primer was: 5'-GCAGGGCAGCTTTCATCC-3' (SEQ ID NO:165). Primers were synthesized by Genosys Corp., The Woodlands, TX. The primer set was assembled into a PCR reaction using 10 the components of the Expand Hi-Fi PCR System™ (Roche Molecular Biochemicals). Twenty-five microliters of the PCR reaction mixture was added to each well of the RapidScan PCR plate. The plate was placed in a GeneAmp PCR9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The plate was then exposed to the following cycling parameters: Pre-soak 94C for 3 minutes; denaturation at 94C for 30 seconds; annealing at primer T_m for 45 seconds; extension 15 72C for 2 minutes; for 35 cycles. PCR productions were then separated and analyzed by electrophoresis on a 2.0% agarose gel.

The 4-log dilution range of cDNA deposited on the plate ensured that the amplification reaction was within the linear range and, facilitated semi-quantitative determination of relative mRNA accumulation in the various tissues or brain regions examined.

20 nGPCR-42 was expressed in the brain, peripheral blood leukocytes, bone marrow, placenta, salivary gland, liver, ovary, uterus, testis, fetal liver, heart, thyroid gland, kidney, adrenal gland, spleen, pancreas, colon, lung, prostate, small intestine, skin, muscle, fetal brain, and stomach. Within the brain, GPCR-42 was expressed in the temporal lobe, cerebellum, substantia nigra, caudate nucleus, amygdala, frontal lobe, thalamus, hippocampus, 25 hypothalamus, pons, medulla, and spinal cord. Expression of the nGPCR-42 in the brain provided an indication that modulators of nGPCR-42 activity have utility for treating disorders, including but not limited to, schizophrenia, affective disorders, metabolic disorders, inflammatory disorders, cancers, ADHD/ADD (i.e., Attention Deficit-Hyperactivity Disorder/Attention Deficit Disorder), and neural disorders such as Alzheimer's disease, 30 Parkinson's disease, migraine, and senile dementia. Some other diseases for which modulators of nGPCR-42 may have utility include depression, anxiety, bipolar disease, epilepsy, neuritis, neurasthenia, neuropathy, neuroses, and the like. Use of nGPCR-42 modulators, including nGPCR-42 ligands and anti-nGPCR-42 antibodies, to treat individuals having such disease states is intended as an aspect of the invention.

nGPCR-46

Tissue specific expression of the cDNA encoding nGPCR-46 was detected using a PCR-based method. Multiple ChoiceTM first strand cDNAs (OriGene Technologies, Rockville, MD) from 12 human tissues are serially diluted over a 3-log range and arrayed into a multi-well PCR plate. This array is used to generate a comprehensive expression profile of nGPCR-46 in human tissues. Human tissues arrayed include: brain, heart, kidney, peripheral blood leukocytes, liver, lung, muscle, ovary, prostate, small intestine, spleen and testis. The forward PCR primer was: 5'-ACGCCCGCTGAACCGTATAC-3' (SEQ ID NO:166) and the reverse primer used was: 5'-GGGTGCCACCTGGTTGCTC-3' (SEQ ID NO:167). This primer set will prime the synthesis of a 242 base pair fragment in the presence of the appropriate cDNA. For detection of expression within brain regions, the same primer set was used with the Human Brain Rapid ScanTM Panel (OriGene Technologies, Rockville, MD). This panel represents serial dilutions over a 2 log range of first strand cDNA from the following brain regions arrayed in a 96 well format: frontal lobe, temporal lobe, cerebellum, hippocampus, substantia nigra, caudate nucleus, amygdala, thalamus, hypothalamus, pons, medulla and spinal cord. Primers were synthesized by Genosys Corp., The Woodlands, TX. PCR reactions were assembled using the components of the Expand Hi-Fi PCR SystemTM (Roche Molecular Biochemicals, Indianapolis, IN). Twenty-five microliters of the PCR reaction mixture was added to each well of the RapidScan PCR plate. The plate was placed in a GeneAmp 9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The following cycling program was executed: Pre-soak at (94C for 3 minutes) followed by 35 cycles of 94C for 45 seconds, 53C for 2 minutes, 72C for 45 seconds. PCR reaction products were then separated and analyzed by electrophoresis on a 2.0% agarose gel stained with ethidium bromide.

nGPCR-46 was expressed in the brain, peripheral blood lymphocytes, testis, heart, kidney, spleen, prostate, ovary, liver, lung, small intestine, and muscle. Within the brain, nGPCR-46 was expressed in frontal lobe, temporal lobe, cerebellum, hippocampus, substantia nigra, caudate nucleus, amygdala, thalamus, hypothalamus, pons, medulla, and spinal cord. Expression of the nGPCR-46 in the brain indicates that modulators of nGPCR-46 activity have utility for treating neurological disorders, including but not limited to, movement disorders, affective disorders, metabolic disorders, inflammatory disorders and cancers. Use of nGPCR-46 modulators, including nGPCR-46 ligands and anti-nGPCR-46 antibodies, to treat individuals having such disease states is intended as an aspect of the invention.

nGPCR-48

Tissue specific expression of the cDNA encoding nGPCR-48 was detected using a PCR-based RapidScan™ Gene Expression Panel (OriGene Technologies, Rockville, MD). The RapidScan Expression Panel is comprised of first-strand cDNAs from 24 human tissues that are serially diluted over a 3-log range and arrayed into a multi-well PCR plate. This array is used to generate a comprehensive expression profile of the putative GPCR in human tissues. Human tissues arrayed include: brain, heart, kidney, spleen, liver, colon, lung, small intestine, muscle, stomach, testis, placenta, salivary gland, thyroid, adrenal gland, pancreas, ovary, uterus, prostate, skin, PBL, bone marrow, fetal brain, fetal liver. The forward PCR primer was 5'-ATGGCACCTTCTCATCGG-3' (SEQ ID NO:168) and the reverse primer was 5'- ACGTAGT ACACCGCCTTG-3' (SEQ ID NO:169). This primer set will prime the synthesis of a 392 base pair fragment in the presence of the appropriate cDNA. For detection of expression within brain regions, the same primer set was used with the Human Brain Rapid Scan™ Panel (OriGene Technologies, Rockville, MD). This panel represents serial dilutions over a 2 log range of first strand cDNA from the following brain regions arrayed in a 96 well format: frontal lobe, temporal lobe, cerebellum, hippocampus, substantia nigra, caudate nucleus, amygdala, thalamus, hypothalamus, pons, medulla and spinal cord. Primers were synthesized by Genosys Corp., The Woodlands, TX. PCR reactions were assembled using the components of the Expand Hi-Fi PCR System™ (Roche Molecular Biochemicals, Indianapolis, IN). Twenty-five microliters of the PCR reaction mixture was added to each well of the RapidScan PCR plate. The plate was placed in a GeneAmp 9700 PCR Thermocycler (PE Applied Biosystems). The following cycling program was executed: Pre-soak at (94°C for 3min.) followed by 35 cycles of 94C for 45 seconds, 53C for 2 minutes, 72C for 45 seconds. PCR reaction products were then separated and analyzed by electrophoresis on a 2.0% agarose gel, stained with ethidium bromide.

The 4-log dilution range of cDNA deposited on the plate ensured that the amplification reaction was within the linear range and, facilitated semi-quantitative determination of relative mRNA accumulation in the various tissues or brain regions examined.

nGPCR-48 was expressed in the ovary, prostate, skin, stomach bone marrow, placenta, fetal liver, brain, heart, spleen, liver, colon, uterus, lung, small intestine, peripheral blood leukocytes, testis, and fetal brain. Within the brain, nGPCR-48 was expressed in the substantia nigra, frontal lobe, temporal lobe, cerebellum, hippocampus, caudate nucleus, thalamus and spinal cord. Expression of nGPCR-48 in the brain provided an indication that modulators of nGPCR-48 activity have utility for treating disorders, including but not limited to, schizophrenia, affective disorders, ADHD/ADD (i.e., Attention Deficit-Hyperactivity Disorder/Attention Deficit Disorder), neural disorders such as Alzheimer's disease, Parkinson's

disease, migraine, senile dementia, depression, anxiety, bipolar disease, epilepsy, neuritis, neurasthenia, neuropathy, neuroses, metabolic disorders, inflammatory disorders, cancers and the like. Use of nGPCR-48 modulators, including nGPCR-48 ligands and anti-nGPCR-48 antibodies, to treat individuals having such disease states is intended as an aspect of the

5 invention.

nGPCR-49

Tissue specific expression of the cDNA encoding SEQ-49 was detected using a PCR-based method. Multiple ChoiceTM first strand cDNAs (OriGene Technologies, Rockville, MD) from 6 human tissues were serially diluted over a 3-log range and arrayed into a multi-well PCR 10 plate. This array was used to generate a comprehensive expression profile of the putative GPCRs in human tissues. Human tissues arrayed included: brain, heart, kidney, peripheral blood leukocytes, lung, and testis. PCR primers were designed based on the available sequence of the Celera sequence GA_11585051. The forward primer used was 5'-AGCAGGTAGATGGAGAA 15 GG-3' (SEQ ID NO:170). The reverse primer used was 5'-GACTCCTGAGGACCATGTC- 3' (SEQ ID NO:171). This primer set primed the synthesis of a 320 base pair fragment in the presence of the appropriate cDNA. PCR reactions were performed in a GeneAmp 9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The following cycling program was executed: Pre-soak at 94C for 3minutes followed by 35 cycles of 94C for 45 seconds, 53.5C for 2 minutes, 20 72C for 45 seconds. PCR reaction products were then separated and analyzed by electrophoresis on a 2.0% agarose gel stained with ethidium bromide.

nGPCR-49 was expressed in the brain, heart, testis, peripheral blood leukocytes, kidney, liver, muscle, ovary, prostate, small intestine, spleen, and lung. Within the brain, nGPCR-49 was expressed in regions including but not limited to, cerebellum, hippocampus, substantia nigra, thalamus, hypothalamus, frontal lobe, temporal lobe, amygdala, pons, medulla, caudate nucleus, 25 and spinal cord. Expression of the nGPCR-49 in the brain provides an indication that modulators of nGPCR-49 activity have utility for treating neurological disorders, including but not limited to, movement disorders, affective disorders, metabolic disorders, inflammatory disorders and cancers. Use of nGPCR-49 modulators, including nGPCR-49 ligands and anti-nGPCR-49 30 antibodies, to treat individuals having such disease states is intended as an aspect of the invention.

nGPCR-61

Tissue specific expression of the cDNA encoding SEQ-61 was detected using a PCR-based method. Multiple ChoiceTM first strand cDNAs (OriGene Technologies, Rockville, MD) from 6 human tissues were serially diluted over a 3-log range and arrayed into a multi-well PCR

plate. This array was used to generate a comprehensive expression profile of the putative GPCRs in human tissues. Human tissues arrayed included: brain, heart, kidney, peripheral blood leukocytes, lung, and testis. PCR primers were designed based on the available sequence of the Celera sequence GA_13368549. The forward primer used was 5'-CATTGGAACCACTTAT 5 TGG-3' (SEQ ID NO:172). The reverse primer used was 5'-AAGCAAGACAGGTGAGAATG- 3' (SEQ ID NO:173). This primer set primed the synthesis of a 284 base pair fragment in the presence of the appropriate cDNA. PCR reactions were performed in a GeneAmp 9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The following cycling program was executed: Pre-soak at 94C for 3 minutes followed by 35 cycles of 94C for 45 seconds, 53.5C for 2 minutes, 10 72C for 45 seconds. PCR reaction products were then separated and analyzed by electrophoresis on a 2.0% agarose gel stained with ethidium bromide.

15 nGPCR-61 was expressed in the brain, heart, testis, peripheral blood leukocytes, and lung. Expression of the nGPCR-61 in the brain provides an indication that modulators of nGPCR-61 activity have utility for treating neurological disorders, including but not limited to, movement disorders, affective disorders, metabolic disorders, inflammatory disorders and cancers. Use of nGPCR-61 modulators, including nGPCR-61 ligands and anti-nGPCR-61 antibodies, to treat individuals having such disease states is intended as an aspect of the invention.

nGPCR-51

20 Tissue specific expression of the cDNA encoding SEQ-51 was detected using a PCR-based method. Multiple ChoiceTM first strand cDNAs (OriGene Technologies, Rockville, MD) from 12 human tissues were serially diluted over a 3-log range and arrayed into a multi-well PCR plate. This array was used to generate a comprehensive expression profile of the putative GPCR in human tissues. Human tissues arrayed include: brain, heart, kidney, peripheral blood 25 leukocytes, liver, lung, muscle, ovary, prostate, small intestine, spleen and testis. PCR primers were designed based on the sequence of nGPCR-51 provided herein as SEQ ID NO:39 and SEQ ID NO:57. The forward primer used was 5'-CCATCTGTCTCAGCTATGCC-3' (SEQ ID NO:174), corresponding base pairs 929 through 948 of SEQ ID NO:57. The reverse primer used was 5'-TCCTTCTCAGTCGCTCTC-3' (SEQ ID NO:175) corresponding to base pairs 1022 30 through 1038 of SEQ ID NO:57. This primer set primes the synthesis of a 112 base pair fragment in the presence of the appropriate cDNA. For detection of expression within brain regions, the same primer set was used with the Human Brain Rapid ScanTM Panel (OriGene Technologies, Rockville, MD). This panel represents serial dilutions over a 3 log range of first strand cDNA from the following brain regions arrayed in a 96 well format: frontal lobe,

temporal lobe, cerebellum, hippocampus, substantia nigra, caudate nucleus, amygdala, thalamus, hypothalamus, pons, medulla and spinal cord. Primers were synthesized by Genosys Corp., The Woodlands, TX. PCR reactions were assembled using the components of the Expand Hi-Fi PCR System™ (Roche Molecular Biochemicals, Indianapolis, IN). Twenty-five microliters of 5 the PCR reaction mixture was added to each well of the RapidScan PCR plate. The plate was placed in a GeneAmp 9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The following cycling program was executed: Pre-soak at 94C for 3 minutes followed by 35 cycles of 94C for 45 seconds, 52C for 2 minutes, 72C for 45 seconds. PCR reaction products were then separated and analyzed by electrophoresis on a 2.0% agarose gel stained with ethidium bromide.

10 nGPCR-51 was expressed in the brain, heart, peripheral blood leukocytes, liver, prostate, testis, lung, small intestine, and spleen. Within the brain, nGPCR-51 was expressed in the cerebellum, hippocampus, substantia nigra, thalamus, hypothalamus, pons, frontal lobe, temporal lobe, caudate nucleus, medulla, spinal cord, and amygdala. Expression of the nGPCR- 15 51 in the brain provides an indication that modulators of nGPCR-51 activity have utility for treating neurological disorders, including but not limited to, movement disorders, affective disorders, metabolic disorders, inflammatory disorders, cancers, attention disorders, anxiety, depression, and obesity. Use of nGPCR-51 modulators, including nGPCR-51 ligands and anti- nGPCR-51 antibodies, to treat individuals having such disease states is intended as an aspect of the invention.

20 *nGPCR-52*

Tissue specific expression of the cDNA encoding SEQ-52 was detected using a PCR-based method. Multiple Choice™ first strand cDNAs (OriGene Technologies, Rockville, MD) from 12 human tissues were serially diluted over a 3-log range and arrayed into a multi-well 25 PCR plate. This array was used to generate a comprehensive expression profile of nGPCR-52 in human tissues. Human tissues arrayed include: brain, heart, kidney, peripheral blood leukocytes, liver, lung, muscle, ovary, prostate, small intestine, spleen and testis. The forward primer used was 5'-CAAACAAACAAACAGCAGAAC-3' (SEQ ID NO:176) and the reverse primer was 5'-TCACAGTCACACCTACCAAG-3' (SEQ ID NO:177). This primer set primed the synthesis of a 274 base pair fragment in the presence of the appropriate cDNA. For detection of 30 expression within brain regions, the same primer set was used with the Human Brain Rapid Scan™ Panel (OriGene Technologies, Rockville, MD). This panel represents serial dilutions over a 2 log range of first strand cDNA from the following brain regions arrayed in a 96 well format: frontal lobe, temporal lobe, cerebellum, hippocampus, substantia nigra, caudate nucleus, amygdala, thalamus, hypothalamus, pons, medulla and spinal cord. Primers were synthesized

by Genosys Corp., The Woodlands, TX. PCR reactions were assembled using the components of the Expand Hi-Fi PCR SystemTM (Roche Molecular Biochemicals, Indianapolis, IN).

Twenty-five microliters of the PCR reaction mixture was added to each well of the RapidScan PCR plate. The plate was placed in a GeneAmp 9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The following cycling program was executed: Pre-soak at 94C for 3 minutes followed by 35 cycles of 94C for 45 seconds, 53C for 2 minutes, 72C for 45 seconds. PCR reaction products were then separated and analyzed by electrophoresis on a 2.0% agarose gel stained with ethidium bromide.

nGPCR-52 was expressed in the brain, the lungs, muscle, small intestine, spleen, testis, heart, peripheral blood leukocytes, and liver. Within the brain, nGPCR-52 was expressed in the frontal lobe, temporal lobe, cerebellum, hippocampus, substantia nigra, caudate nucleus, amygdala, hypothalamus, and medulla. Expression of nGPCR-52 in the brain provides an indication that modulators of nGPCR-52 activity have utility for treating neurological disorders, including but not limited to, movement disorders, affective disorders, metabolic disorders, inflammatory disorders and cancers. Use of nGPCR-52 modulators, including nGPCR-52 ligands and anti-nGPCR-52 antibodies, to treat individuals having such disease states is intended as an aspect of the invention.

Example 6: Chromosomal Localization

nGPCR-51

The chromosomal location of the gene encoding SEQ51 was determined using the Stanford G3 Radiation Hybrid Panel (Research Genetics, Inc., Huntsville, AL). This panel contains 83 radiation hybrid clones of the entire human genome created by the Stanford Human Genome Center. PCR reactions were assembled containing 25ng of DNA from each clone and the components of the Expand Hi-Fi PCR SystemTM (Roche Molecular Biochemicals, Indianapolis, IN) in a final reaction volume of 15 ul. PCR primers were synthesized by Genosys Corp., The Woodlands, TX. PCR primers were designed based on the available sequence of the Celera sequence GA_11007426. The forward primer used was 5'-CCATCTGTCTCAGCTAT GCC-3' (SEQ ID NO:178) corresponding base pairs 241 through 260 of GA_11007426. The reverse primer used was 5'-TCCTTCTCAGTCGCTCTTC-3' (SEQ ID NO:179) corresponding to base pairs 334 through 352 of GA_11007426. This primer set will prime the synthesis of a 112 base pair fragment in the presence of the appropriate genomic DNA. PCR reactions were incubated in a GeneAmp 9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The following cycling program was executed: Pre-soak at 94C for 3 minutes, and 35 cycles at 94C for 30 seconds, 52C for 60 seconds, 72C for 2 minutes. PCR reaction products were then

separated and analyzed by electrophoresis on a 2.0% agarose gel, stained with ethidium bromide. Lanes were scored for the presence or absence of the expected PCR product and the results submitted to the Stanford Human Genome Center via e-mail for analysis (world wide web of the Internet at, for example, shgc.stanford.edu./RH/rhserverformnew.html). G3

5 Radiation Hybrid Panel Analysis of the SEQ51 places it on chromosome 6, most nearly linked to Stanford marker SHGC-34355 with a LOD score of 12.32. This marker lies at position 6q21. Chromosome 6q21-q22.3 contains a highly significant disease locus for schizophrenia (Cao *et al.*, *Genomics* 43: 1-8, (1997)). Genes which map to this region of the chromosome are candidate genes for schizophrenia. Any genes localized to chromosomal regions in linkage with 10 schizophrenia are candidate genes for disease susceptibility. Genes in these regions with the potential to play a biochemical/functional role in the disease process (like G protein coupled receptors) have a high probability of being a disease modifying locus. nGPCR-51, because of its chromosomal location, is an attractive target therefor for screening ligands useful in modulating cellular processes involved in schizophrenia.

15 *nGPCR-52*

The chromosomal location of the gene encoding Seq-52 was determined using the Stanford G3 Radiation Hybrid Panel (Research Genetics, Inc., Huntsville, AL). This panel contains 83 radiation hybrid clones of the entire human genome created by the Stanford Human Genome Center. PCR reactions were assembled containing 25ng of DNA from each clone and 20 the components of the Expand Hi-Fi PCR SystemTM (Roche Molecular Biochemicals, Indianapolis, IN) in a final reaction volume of 15 μ l. PCR primers were synthesized by Genosys Corp., The Woodlands, TX. The forward primer used was RRH421: 5'-GTCTATG CTACCAAGTTCAACC-3' (SEQ ID NO:180) corresponding to base pairs 3328 through 3348. The reverse primer used was RRH422: 5'-ATTCCTCCAGCCCCATCATC-3' (SEQ ID NO:181) 25 corresponding to base pairs 3433 through 3451. This primer set will prime the synthesis of a 124 base pair fragment in the presence of the appropriate genomic DNA. PCR reactions were incubated in a GeneAmp 9700 PCR thermocycler (Perkin Elmer Applied Biosystems). The following cycling program was executed: Pre-soak at 94C for 3 minutes, and 35 cycles of 94C for 30 seconds, 55C for 60 seconds, 72C for 2 minutes. PCR reaction products were then 30 separated and analyzed by electrophoresis on a 2.0% agarose gel, stained with ethidium bromide. Lanes were scored for the presence or absence of the expected PCR product and the results submitted to the Stanford Human Genome Center via e-mail for analysis (world wide web of the Internet at, for example, shgc.stanford.edu./RH/rhserverformnew.html). The chromosomal location of the gene encoding nGPCR-52 was determined using the Stanford G3

Radiation Hybrid Panel (Research Genetics, Inc., Huntsville, AL). This panel contains 83 radiation hybrid clones of the entire human genome created by the Stanford Human Genome Center. Lanes were scored for the presence or absence of the expected PCR product and the results submitted to the Stanford Human Genome Center via e-mail for analysis. The analysis of 5 nGPCR-52 places it on chromosome 6, most nearly linked to Stanford marker SHGC-1836 with a LOD score of 15.65. This marker lies at position 6q21. Chromosome 6q21-q22.3 contains a highly significant disease locus for schizophrenia (Cao *et al.*, *Genomics* 43: 1-8, (1997)). Genes which map to this region of the chromosome are candidate genes for schizophrenia. Any genes localized to chromosomal regions in linkage with schizophrenia are candidate genes for disease 10 susceptibility. Genes in these regions with the potential to play a biochemical/functional role in the disease process (like G protein coupled receptors) have a high probability of being a disease modifying locus. nGPCR-52 because of its chromosomal location is an attractive target therefore for screening ligands useful in modulating cellular processes involved in schizophrenia.

Example 7: Northern Blot Analysis

15 Northern blots are performed to examine the expression of nGPCR-x mRNA. The sense orientation oligonucleotide and the antisense-orientation oligonucleotide, described above, are used as primers to amplify a portion of the GPCR-x cDNA sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60. Multiple human tissue northern blots from Clontech (Human II # 7767-1) are hybridized with the probe. Pre-hybridization is carried out at 20 42C for 4 hours in 5xSSC, 1X Denhardt's reagent, 0.1% SDS, 50% formamide, 250 mg/ml salmon sperm DNA. Hybridization is performed overnight at 42C in the same mixture with the addition of about 1.5×10^6 cpm/ml of labeled probe. The probe is labeled with α -³²P-dCTP by Rediprime™ DNA labeling system (Amersham Pharmacia), purified on Nick Column™ (Amersham Pharmacia) and added to the hybridization solution. The filters are washed several 25 times at 42C in 0.2x SSC, 0.1% SDS. Filters are exposed to Kodak XAR film (Eastman Kodak Company, Rochester, N.Y., USA) with intensifying screen at -80C.

Example 8: Recombinant Expression of nGPCR-x in Eukaryotic Host Cells

Expression of nGPCR-x in Mammalian Cells

To produce nGPCR-x protein, a nGPCR-x-encoding polynucleotide is expressed in a 30 suitable host cell using a suitable expression vector and standard genetic engineering techniques. For example, the nGPCR-x-encoding sequence described in Example 1 is subcloned into the commercial expression vector pzeoSV2 (Invitrogen, San Diego, CA) and transfected into Chinese Hamster Ovary (CHO) cells using the transfection reagent FuGENE6™ (Boehringer-Mannheim) and the transfection protocol provided in the product insert. Other

eukaryotic cell lines, including human embryonic kidney (HEK293) and COS cells, are suitable as well. Cells stably expressing nGPCR-x are selected by growth in the presence of 100 µg/ml zeocin (Stratagene, LaJolla, CA). Optionally, nGPCR-x may be purified from the cells using standard chromatographic techniques. To facilitate purification, antisera is raised against one or 5 more synthetic peptide sequences that correspond to portions of the nGPCR-x amino acid sequence, and the antisera is used to affinity purify nGPCR-x. The nGPCR-x also may be expressed in-frame with a tag sequence (e.g., polyhistidine, hemagglutinin, FLAG) to facilitate purification. Moreover, it will be appreciated that many of the uses for nGPCR-x polypeptides, such as assays described below, do not require purification of nGPCR-x from the host cell.

10 *Expression of nGPCR-x in 293 cells*

For expression of nGPCR-x in mammalian cells HEK293 (transformed human, primary embryonic kidney cells), a plasmid bearing the relevant nGPCR-x coding sequence is prepared, using vector pSecTag2A (Invitrogen). Vector pSecTag2A contains the murine IgK chain leader sequence for secretion, the c-myc epitope for detection of the recombinant protein with the anti-myc antibody, a C-terminal polyhistidine for purification with nickel chelate chromatography, and a Zeocin resistant gene for selection of stable transfecants. The forward primer for 15 amplification of this GPCR cDNA is determined by routine procedures and preferably contains a 5' extension of nucleotides to introduce the *HindIII* cloning site and nucleotides matching the GPCR sequence. The reverse primer is also determined by routine procedures and preferably contains a 5' extension of nucleotides to introduce an *XhoI* restriction site for cloning and 20 nucleotides corresponding to the reverse complement of the nGPCR-x sequence. The PCR conditions are 55C as the annealing temperature. The PCR product is gel purified and cloned into the *HindIII-XhoI* sites of the vector.

The DNA is purified using Qiagen chromatography columns and transfected into 293 25 cells using DOTAP™ transfection media (Boehringer Mannheim, Indianapolis, IN). Transiently transfected cells are tested for expression after 24 hours of transfection, using western blots probed with anti-His and anti-nGPCR-x peptide antibodies. Permanently transfected cells are selected with Zeocin and propagated. Production of the recombinant protein is detected from both cells and media by western blots probed with anti-His, anti-Myc or anti-GPCR peptide 30 antibodies.

Expression of nGPCR-x in COS cells

For expression of the nGPCR-x in COS7 cells, a polynucleotide molecule having a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60 can be cloned into vector p3-CI. This vector is a pUC18-derived plasmid that contains the HCMV (human

cytomegalovirus) promoter-intron located upstream from the bGH (bovine growth hormone) polyadenylation sequence and a multiple cloning site. In addition, the plasmid contains the dhfr (dihydrofolate reductase) gene which provides selection in the presence of the drug methotrexane (MTX) for selection of stable transformants.

5 The forward primer is determined by routine procedures and preferably contains a 5' extension which introduces an *Xba*I restriction site for cloning, followed by nucleotides which correspond to a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60. The reverse primer is also determined by routine procedures and preferably contains 5'-extension of nucleotides which introduces a *Sal*I cloning site followed by nucleotides which 10 correspond to the reverse complement of a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60. The PCR consists of an initial denaturation step of 5 minutes at 95C, 30 cycles of 30 seconds denaturation at 95C, 30 seconds annealing at 58C and 30 seconds extension at 72C, followed by 5 minutes extension at 72C. The PCR product is gel purified and ligated into the *Xba*I and *Sal*I sites of vector p3-Cl. This construct is transformed into *E. coli* 15 cells for amplification and DNA purification. The DNA is purified with Qiagen chromatography columns and transfected into COS 7 cells using Lipofectamine™ reagent from BRL, following the manufacturer's protocols. Forty-eight and 72 hours after transfection, the media and the cells are tested for recombinant protein expression.

20 nGPCR-x expressed from a COS cell culture can be purified by concentrating the cell-growth media to about 10 mg of protein/ml, and purifying the protein by, for example, chromatography. Purified nGPCR-x is concentrated to 0.5 mg/ml in an Amicon concentrator fitted with a YM-10 membrane and stored at -80C.

Expression of nGPCR-x in Insect Cells

25 For expression of nGPCR-x in a baculovirus system, a polynucleotide molecule having a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60 can be amplified by PCR. The forward primer is determined by routine procedures and preferably contains a 5' extension which adds the *Nde*I cloning site, followed by nucleotides which correspond to a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60. The reverse primer is also determined by routine procedures and preferably contains a 5' 30 extension which introduces the *Kpn*I cloning site, followed by nucleotides which correspond to the reverse complement of a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60.

The PCR product is gel purified, digested with *Nde*I and *Kpn*I, and cloned into the corresponding sites of vector pACHTL-A (Pharmingen, San Diego, CA). The pAcHTL

expression vector contains the strong polyhedrin promoter of the *Autographa californica* nuclear polyhedrosis virus (AcMNPV), and a 6XHis tag upstream from the multiple cloning site. A protein kinase site for phosphorylation and a thrombin site for excision of the recombinant protein precede the multiple cloning site is also present. Of course, many other baculovirus 5 vectors could be used in place of pAcHTL-A, such as pAc373, pVL941 and pAcIM1. Other suitable vectors for the expression of GPCR polypeptides can be used, provided that the vector construct includes appropriately located signals for transcription, translation, and trafficking, such as an in-frame AUG and a signal peptide, as required. Such vectors are described in Luckow *et al.*, *Virology* 170:31-39, among others. The virus is grown and isolated using 10 standard baculovirus expression methods, such as those described in Summers *et al.* (*A Manual of Methods for Baculovirus Vectors and Insect Cell Culture Procedures*, Texas Agricultural Experimental Station Bulletin No. 1555 (1987)).

In a preferred embodiment, pAcHTL-A containing nGPCR-x gene is introduced into baculovirus using the "BaculoGold™" transfection kit (Pharmingen, San Diego, CA) using 15 methods established by the manufacturer. Individual virus isolates are analyzed for protein production by radiolabeling infected cells with 35 S-methionine at 24 hours post infection. Infected cells are harvested at 48 hours post infection, and the labeled proteins are visualized by SDS-PAGE. Viruses exhibiting high expression levels can be isolated and used for scaled up expression.

20 For expression of a nGPCR-x polypeptide in a Sf9 cells, a polynucleotide molecule having a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60 can be amplified by PCR using the primers and methods described above for baculovirus expression. The nGPCR-x cDNA is cloned into vector pAcHTL-A (Pharmingen) for expression in Sf9 insect. The insert is cloned into the *NdeI* and *KpnI* sites, after elimination of an internal *NdeI* 25 site (using the same primers described above for expression in baculovirus). DNA is purified with Qiagen chromatography columns and expressed in Sf9 cells. Preliminary Western blot experiments from non-purified plaques are tested for the presence of the recombinant protein of the expected size which reacted with the GPCR-specific antibody. These results are confirmed after further purification and expression optimization in HiG5 cells.

30 **Example 9: Interaction Trap/Two-Hybrid System**

In order to assay for nGPCR-x-interacting proteins, the interaction trap/two-hybrid library screening method can be used. This assay was first described in Fields *et al.*, *Nature*, 1989, 340, 245, which is incorporated herein by reference in its entirety. A protocol is published in Current Protocols in Molecular Biology 1999, John Wiley & Sons, NY, and Ausubel, F. M. *et*

al. 1992, Short protocols in molecular biology, Fourth edition, Greene and Wiley-interscience, NY, each of which is incorporated herein by reference in its entirety. Kits are available from Clontech, Palo Alto, CA (Matchmaker Two-Hybrid System 3).

A fusion of the nucleotide sequences encoding all or partial nGPCR-x and the yeast transcription factor GAL4 DNA-binding domain (DNA-BD) is constructed in an appropriate plasmid (*i.e.*, pGBKT7) using standard subcloning techniques. Similarly, a GAL4 active domain (AD) fusion library is constructed in a second plasmid (*i.e.*, pGADT7) from cDNA of potential GPCR-binding proteins (for protocols on forming cDNA libraries, see Sambrook *et al.* 1989, Molecular cloning: a laboratory manual, second edition, Cold Spring Harbor Press, Cold Spring Harbor, NY), which is incorporated herein by reference in its entirety. The DNA-BD/nGPCR-x fusion construct is verified by sequencing, and tested for autonomous reporter gene activation and cell toxicity, both of which would prevent a successful two-hybrid analysis. Similar controls are performed with the AD/library fusion construct to ensure expression in host cells and lack of transcriptional activity. Yeast cells are transformed (*ca.* 105 transformants/mg DNA) with both the nGPCR-x and library fusion plasmids according to standard procedures (Ausubel *et al.*, 1992, Short protocols in molecular biology, fourth edition, Greene and Wiley-interscience, NY, which is incorporated herein by reference in its entirety). *In vivo* binding of DNA-BD/nGPCR-x with AD/library proteins results in transcription of specific yeast plasmid reporter genes (*i.e.*, lacZ, HIS3, ADE2, LEU2). Yeast cells are plated on nutrient-deficient media to screen for expression of reporter genes. Colonies are dually assayed for β -galactosidase activity upon growth in Xgal (5-bromo-4-chloro-3-indolyl- β -D-galactoside) supplemented media (filter assay for β -galactosidase activity is described in Breeden *et al.*, Cold Spring Harb. Symp. Quant. Biol., 1985, 50, 643, which is incorporated herein by reference in its entirety). Positive AD-library plasmids are rescued from transformants and reintroduced into the original yeast strain as well as other strains containing unrelated DNA-BD fusion proteins to confirm specific nGPCR-x/library protein interactions. Insert DNA is sequenced to verify the presence of an open reading frame fused to GAL4 AD and to determine the identity of the nGPCR-x-binding protein.

Example 10: Mobility Shift DNA-Binding Assay Using Gel Electrophoresis

A gel electrophoresis mobility shift assay can rapidly detect specific protein-DNA interactions. Protocols are widely available in such manuals as Sambrook *et al.* 1989, *Molecular cloning: a laboratory manual*, second edition, Cold Spring Harbor Press, Cold Spring Harbor, NY and Ausubel, F. M. *et al.*, 1992, *Short Protocols in Molecular Biology*, fourth edition, Greene and Wiley-interscience, NY, each of which is incorporated herein by reference in its entirety.

Probe DNA(<300 bp) is obtained from synthetic oligonucleotides, restriction endonuclease fragments, or PCR fragments and end-labeled with ^{32}P . An aliquot of purified nGPCR-x (ca. 15 μg) or crude nGPCR-x extract (ca. 15 ng) is incubated at constant temperature (in the range 22-37C) for at least 30 minutes in 10-15 μl of buffer (*i.e.* TAE or TBE, pH 8.0-8.5) 5 containing radiolabeled probe DNA, nonspecific carrier DNA (ca. 1 μg), BSA (300 $\mu\text{g}/\text{ml}$), and 10% (v/v) glycerol. The reaction mixture is then loaded onto a polyacrylamide gel and run at 30-35 mA until good separation of free probe DNA from protein-DNA complexes occurs. The gel is then dried and bands corresponding to free DNA and protein-DNA complexes are detected by autoradiography.

10 **Example 11: Antibodies to nGPCR-x**

Standard techniques are employed to generate polyclonal or monoclonal antibodies to the nGPCR-x receptor, and to generate useful antigen-binding fragments thereof or variants thereof, including "humanized" variants. Such protocols can be found, for example, in 15 Sambrook *et al.* (1989) and Harlow *et al.* (Eds.), *Antibodies A Laboratory Manual*; Cold Spring Harbor Laboratory; Cold Spring Harbor, NY (1988). In one embodiment, recombinant nGPCR-x polypeptides (or cells or cell membranes containing such polypeptides) are used as antigen to generate the antibodies. In another embodiment, one or more peptides having amino acid 20 sequences corresponding to an immunogenic portion of nGPCR-x (*e.g.*, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, or more amino acids) are used as antigen. Peptides corresponding to extracellular portions of nGPCR-x, especially hydrophilic extracellular portions, are preferred. The antigen may be mixed with an adjuvant or linked to a hapten to increase antibody production.

Polyclonal or Monoclonal antibodies

As one exemplary protocol, recombinant nGPCR-x or a synthetic fragment thereof is 25 used to immunize a mouse for generation of monoclonal antibodies (or larger mammal, such as a rabbit, for polyclonal antibodies). To increase antigenicity, peptides are conjugated to Keyhole Lympet Hemocyanin (Pierce), according to the manufacturer's recommendations. For an initial injection, the antigen is emulsified with Freund's Complete Adjuvant and injected subcutaneously. At intervals of two to three weeks, additional aliquots of nGPCR-x antigen are 30 emulsified with Freund's Incomplete Adjuvant and injected subcutaneously. Prior to the final booster injection, a serum sample is taken from the immunized mice and assayed by western blot to confirm the presence of antibodies that immunoreact with nGPCR-x. Serum from the immunized animals may be used as polyclonal antisera or used to isolate polyclonal antibodies

that recognize nGPCR-x. Alternatively, the mice are sacrificed and their spleen removed for generation of monoclonal antibodies.

To generate monoclonal antibodies, the spleens are placed in 10 ml serum-free RPMI 1640, and single cell suspensions are formed by grinding the spleens in serum-free RPMI 1640, 5 supplemented with 2 mM L-glutamine, 1 mM sodium pyruvate, 100 units/ml penicillin, and 100 µg/ml streptomycin (RPMI) (Gibco, Canada). The cell suspensions are filtered and washed by centrifugation and resuspended in serum-free RPMI. Thymocytes taken from three naive Balb/c mice are prepared in a similar manner and used as a Feeder Layer. NS-1 myeloma cells, kept in log phase in RPMI with 10% fetal bovine serum (FBS) (Hyclone Laboratories, Inc., Logan, 10 Utah) for three days prior to fusion, are centrifuged and washed as well.

To produce hybridoma fusions, spleen cells from the immunized mice are combined with NS-1 cells and centrifuged, and the supernatant is aspirated. The cell pellet is dislodged by tapping the tube, and 2 ml of 37C PEG 1500 (50% in 75 mM HEPES, pH 8.0) (Boehringer-Mannheim) is stirred into the pellet, followed by the addition of serum-free RPMI. Thereafter, 15 the cells are centrifuged, resuspended in RPMI containing 15% FBS, 100 µM sodium hypoxanthine, 0.4 µM aminopterin, 16 µM thymidine (HAT) (Gibco), 25 units/ml IL-6 (Boehringer-Mannheim) and 1.5 x 10⁶ thymocytes/ml, and plated into 10 Corning flat-bottom 96-well tissue culture plates (Corning, Corning New York).

On days 2, 4, and 6 after the fusion, 100 µl of medium is removed from the wells of the 20 fusion plates and replaced with fresh medium. On day 8, the fusions are screened by ELISA, testing for the presence of mouse IgG that binds to nGPCR-x. Selected fusion wells are further cloned by dilution until monoclonal cultures producing anti-nGPCR-x antibodies are obtained.

Humanization of anti-nGPCR-x monoclonal antibodies

The expression pattern of nGPCR-x as reported herein and the proven track record of 25 GPCRs as targets for therapeutic intervention suggest therapeutic indications for nGPCR-x inhibitors (antagonists). nGPCR-x-neutralizing antibodies comprise one class of therapeutics useful as nGPCR-x antagonists. Following are protocols to improve the utility of anti-nGPCR-x monoclonal antibodies as therapeutics in humans by “humanizing” the monoclonal antibodies to improve their serum half-life and render them less immunogenic in human hosts (*i.e.*, to prevent 30 human antibody response to non-human anti-nGPCR-x antibodies).

The principles of humanization have been described in the literature and are facilitated by the modular arrangement of antibody proteins. To minimize the possibility of binding complement, a humanized antibody of the IgG4 isotype is preferred.

For example, a level of humanization is achieved by generating chimeric antibodies comprising the variable domains of non-human antibody proteins of interest with the constant domains of human antibody molecules. (See, *e.g.*, Morrison *et al.*, *Adv. Immunol.*, 44:65-92 (1989)). The variable domains of nGPCR-x-neutralizing anti-nGPCR-x antibodies are cloned 5 from the genomic DNA of a B-cell hybridoma or from cDNA generated from mRNA isolated from the hybridoma of interest. The V region gene fragments are linked to exons encoding human antibody constant domains, and the resultant construct is expressed in suitable mammalian host cells (*e.g.*, myeloma or CHO cells).

To achieve an even greater level of humanization, only those portions of the variable 10 region gene fragments that encode antigen-binding complementarity determining regions ("CDR") of the non-human monoclonal antibody genes are cloned into human antibody sequences. (See, *e.g.*, Jones *et al.*, *Nature* 321:522-525 (1986); Riechmann *et al.*, *Nature* 332:323-327 (1988); Verhoeyen *et al.*, *Science* 239:1534-36 (1988); and Tempest *et al.*, *Bio/Technology* 9: 266-71 (1991)). If necessary, the β -sheet framework of the human antibody 15 surrounding the CDR3 regions also is modified to more closely mirror the three dimensional structure of the antigen-binding domain of the original monoclonal antibody. (See Kettleborough *et al.*, *Protein Engin.*, 4:773-783 (1991); and Foote *et al.*, *J. Mol. Biol.*, 224:487-499 (1992)).

In an alternative approach, the surface of a non-human monoclonal antibody of interest 20 is humanized by altering selected surface residues of the non-human antibody, *e.g.*, by site-directed mutagenesis, while retaining all of the interior and contacting residues of the non-human antibody. See Padlan, *Molecular Immunol.*, 28(4/5):489-98 (1991).

The foregoing approaches are employed using nGPCR-x-neutralizing anti-nGPCR-x monoclonal antibodies and the hybridomas that produce them to generate humanized nGPCR-x 25 neutralizing antibodies useful as therapeutics to treat or palliate conditions wherein nGPCR-x expression or ligand-mediated nGPCR-x signaling is detrimental.

Human nGPCR-x-Neutralizing Antibodies from Phage Display

Human nGPCR-x-neutralizing antibodies are generated by phage display techniques such as those described in Aujame *et al.*, *Human Antibodies* 8(4):155-168 (1997); 30 Hoogenboom, *TIBTECH* 15:62-70 (1997); and Rader *et al.*, *Curr. Opin. Biotechnol.* 8:503-508 (1997), each of which is incorporated herein by reference in its entirety. For example, antibody variable regions in the form of Fab fragments or linked single chain Fv fragments are fused to the amino terminus of filamentous phage minor coat protein pIII. Expression of the fusion protein and incorporation thereof into the mature phage coat results in phage particles that

present an antibody on their surface and contain the genetic material encoding the antibody. A phage library comprising such constructs is expressed in bacteria, and the library is screened for nGPCR-x-specific phage-antibodies using labeled or immobilized nGPCR-x as antigen-probe.

Human nGPCR-x-neutralizing antibodies from transgenic mice

5 Human nGPCR-x-neutralizing antibodies are generated in transgenic mice essentially as described in Bruggemann *et al.*, *Immunol. Today* 17(8):391-97 (1996) and Bruggemann *et al.*, *Curr. Opin. Biotechnol.* 8:455-58 (1997). Transgenic mice carrying human V-gene segments in germline configuration and that express these transgenes in their lymphoid tissue are immunized with a nGPCR-x composition using conventional immunization protocols. Hybridomas are
10 generated using B cells from the immunized mice using conventional protocols and screened to identify hybridomas secreting anti-nGPCR-x human antibodies (e.g., as described above).

Example 12: Assays to Identify Modulators of nGPCR-x Activity

Set forth below are several nonlimiting assays for identifying modulators (agonists and antagonists) of nGPCR-x activity. Among the modulators that can be identified by these assays
15 are natural ligand compounds of the receptor; synthetic analogs and derivatives of natural ligands; antibodies, antibody fragments, and/or antibody-like compounds derived from natural antibodies or from antibody-like combinatorial libraries; and/or synthetic compounds identified by high-throughput screening of libraries; and the like. All modulators that bind nGPCR-x are useful for identifying nGPCR-x in tissue samples (e.g., for diagnostic purposes, pathological
20 purposes, and the like). Agonist and antagonist modulators are useful for up-regulating and down-regulating nGPCR-x activity, respectively, to treat disease states characterized by abnormal levels of nGPCR-x activity. The assays may be performed using single putative modulators, and/or may be performed using a known agonist in combination with candidate antagonists (or visa versa).

25 *A. cAMP Assays*

In one type of assay, levels of cyclic adenosine monophosphate (cAMP) are measured in nGPCR-x-transfected cells that have been exposed to candidate modulator compounds. Protocols for cAMP assays have been described in the literature. (See, e.g., Sutherland *et al.*, *Circulation* 37: 279 (1968); Frandsen *et al.*, *Life Sciences* 18: 529-541 (1976); Dooley *et al.*,
30 *Journal of Pharmacology and Experimental Therapeutics* 283 (2): 735-41 (1997); and George *et al.*, *Journal of Biomolecular Screening* 2 (4): 235-40 (1997)). An exemplary protocol for such an assay, using an Adenylyl Cyclase Activation FlashPlate® Assay from NEN™ Life Science Products, is set forth below.

Briefly, the nGPCR-x coding sequence (*e.g.*, a cDNA or intronless genomic DNA) is subcloned into a commercial expression vector, such as pzeoSV2 (Invitrogen), and transiently transfected into Chinese Hamster Ovary (CHO) cells using known methods, such as the transfection protocol provided by Boehringer-Mannheim when supplying the FuGENE 6 transfection reagent. Transfected CHO cells are seeded into 96-well microplates from the FlashPlate® assay kit, which are coated with solid scintillant to which antisera to cAMP has been bound. For a control, some wells are seeded with wild type (untransfected) CHO cells. Other wells in the plate receive various amounts of a cAMP standard solution for use in creating a standard curve.

One or more test compounds (*i.e.*, candidate modulators) are added to the cells in each well, with water and/or compound-free medium/diluent serving as a control or controls. After treatment, cAMP is allowed to accumulate in the cells for exactly 15 minutes at room temperature. The assay is terminated by the addition of lysis buffer containing [¹²⁵I]-labeled cAMP, and the plate is counted using a Packard Topcount™ 96-well microplate scintillation counter. Unlabeled cAMP from the lysed cells (or from standards) and fixed amounts of [¹²⁵I]-cAMP compete for antibody bound to the plate. A standard curve is constructed, and cAMP values for the unknowns are obtained by interpolation. Changes in intracellular cAMP levels of cells in response to exposure to a test compound are indicative of nGPCR-x modulating activity. Modulators that act as agonists of receptors which couple to the G_s subtype of G proteins will stimulate production of cAMP, leading to a measurable 3-10 fold increase in cAMP levels. Agonists of receptors which couple to the G_{i/o} subtype of G proteins will inhibit forskolin-stimulated cAMP production, leading to a measurable decrease in cAMP levels of 50-100%. Modulators that act as inverse agonists will reverse these effects at receptors that are either constitutively active or activated by known agonists.

25 nGPCR-51

Modulators that act as agonists at receptors which couple to the G_s subtype of G proteins will activate adenylyl cyclase leading to a 3-10 fold increase in cyclic adenosine monophosphate (cAMP). Compounds to be tested for the ability to activate nGPCR-51 were assayed for cAMP using an Adenylyl Cyclase Activation FlashPlate® Assay from NEN™ Life Science Products. Briefly, nGPCR-51 cDNA is subcloned into the commercial expression vector pCMV Sport (Gibco/Life Technologies) and transiently transfected into CHO or COS 7 cells using the transfection reagent FuGENE 6 (Boehringer-Mannheim) and the transfection protocol provided in the product insert. 24 hours post transfection the cells are harvested by dislodging from the culture flask using Versene (Gibco/BRL). The cells are counted and prepared as a suspension in

a buffer included in the assay kit that contains the phosphodiesterase inhibitor isobutylmethylxanthine. The assay is conducted in a special 96 well microplate included in the kit which is coated with solid scintillant to which antisera to cAMP has been bound. Dilutions of test compounds to be tested for activation of nGPCR-51 are added to assay wells. Several wells 5 on the plate receive various amounts of cAMP standard solution. After the addition of cells transiently expressing nGPCR-51, cAMP is allowed to accumulate for exactly 15 minutes at room temperature. The assay is terminated by the addition of lysis buffer containing [¹²⁵I] cAMP, and the plate is covered and allowed to incubate at room temperature for 2-24 hours. The plate is then counted using a Packard Topcount™ 96-well microplate scintillation counter.

10 Unlabelled cAMP from cells (or standards) competes with fixed amounts of [¹²⁵I] cAMP for antibody bound to the plate. A standard curve is constructed and cAMP values for the unknowns are obtained by interpolation. Data were analyzed using GraphPad Prism (San Diego, CA). There was no increase in cAMP observed after stimulation with the test compounds at concentrations up to 10 uM.

15 *B. Aequorin Assays*

In another assay, cells (e.g., CHO cells) are transiently co-transfected with both a nGPCR-x expression construct and a construct that encodes the photoprotein apoaequorin. In the presence of the cofactor coelenterazine, apoaequorin will emit a measurable luminescence that is proportional to the amount of intracellular (cytoplasmic) free calcium. (See generally, Cobbold, 20 et al. "Aequorin measurements of cytoplasmic free calcium," In: McCormack J.G. and Cobbold P.H., eds., Cellular Calcium: A Practical Approach. Oxford:IRL Press (1991); Stables et al., Analytical Biochemistry 252: 115-26 (1997); and Haugland, Handbook of Fluorescent Probes and Research Chemicals, Sixth edition. Eugene OR: Molecular Probes (1996).)

In one exemplary assay, nGPCR-x is subcloned into the commercial expression vector 25 pzeoSV2 (Invitrogen) and transiently co-transfected along with a construct that encodes the photoprotein apoaequorin (Molecular Probes, Eugene, OR) into CHO cells using the transfection reagent FuGENE 6 (Boehringer-Mannheim) and the transfection protocol provided in the product insert.

The cells are cultured for 24 hours at 37C in MEM (Gibco/BRL, Gaithersburg, MD) 30 supplemented with 10% fetal bovine serum, 2 mM glutamine, 10 U/ml penicillin and 10 µg/ml streptomycin, at which time the medium is changed to serum-free MEM containing 5 µM coelenterazine (Molecular Probes, Eugene, OR). Culturing is then continued for two additional hours at 37C. Subsequently, cells are detached from the plate using VERSEN (Gibco/BRL), washed, and resuspended at 200,000 cells/ml in serum-free MEM.

Dilutions of candidate nGPCR-x modulator compounds are prepared in serum-free MEM and dispensed into wells of an opaque 96-well assay plate at 50 μ l/well. Plates are then loaded onto an MLX microtiter plate luminometer (Dynex Technologies, Inc., Chantilly, VA). The instrument is programmed to dispense 50 μ l cell suspensions into each well, one well at a time, and immediately read luminescence for 15 seconds. Dose-response curves for the candidate modulators are constructed using the area under the curve for each light signal peak. Data are analyzed with SlideWrite, using the equation for a one-site ligand, and EC₅₀ values are obtained. Changes in luminescence caused by the compounds are considered indicative of modulatory activity. Modulators that act as agonists at receptors which couple to the G_q subtype of G proteins give an increase in luminescence of up to 100 fold. Modulators that act as inverse agonists will reverse this effect at receptors that are either constitutively active or activated by known agonists.

nGPCR-51

Agonist activation of receptors that couple to the G_q subtype of G proteins will lead to the release of intracellular calcium. The photoprotein aequorin emits a characteristic luminescence in the presence of calcium and may be expressed in cells along with the receptor of interest in order to report agonist signalling. Peptide A was tested for the ability to activate nGPCR-51 using an assay for aequorin. Peptide A, a cyclic peptide via a disulfide bond between the Cys residues, has the following amino acid sequence: Asp Phe Asp Met Leu Arg Cys Met Leu Gly Arg Val Tyr Arg Pro Cys Trp Gln Val (SEQ ID NO:183). Preferably, Peptide A is used in a solution containing 0.17M acetic acid, or the like, and a stabilizer such as a 0.2% BSA:acetonitrile 90:10. Briefly, nGPCR-51 cDNA is subcloned into the commercial expression vector pCMV Sport (Gibco/Life Technologies) and transiently transfected along with the expression construct mtAequorin (Molecular Probes, Eugene, OR) into COS 7 cells using the transfection reagent FuGENE 6 (Boehringer-Mannheim) and the transfection protocol provided in the product insert. 24 hours post transfection the cells are harvested by dislodging from the culture flask using Versene (Gibco/BRL) and prepared as a suspension in assay buffer (Dulbecco's Modified Eagle's Medium with high glucose, pyridoxine HCl, L-glutamine, sodium pyruvate, and 0.1% fetal bovine serum (Gibco/BRL)) and containing the cofactor coelenterazine (Molecular Probes). The cell suspension is incubated for 4 hours at room temperature with gentle stirring. After the coelenterazine loading incubation, the cells are counted and diluted to 1 \times 10⁶ cells/ml in assay buffer. Dilutions of test compound are prepared in assay buffer and pipetted into wells of an opaque 96-well assay plate, 50 μ l/well. Plates are loaded onto an MLX microtiter plate luminometer (Dynex Technologies, Inc., Chantilly, VA). The instrument is

programmed to dispense 50 μ l cell suspension into each well, one well at a time, and immediately read luminescence for 20 seconds. Dose response curves are constructed using the area under the curve for each light signal peak. Data are analyzed with SlideWrite, using the equation $y = a^0 * a^1/(x + a^1) + a^2$ where a^0 is the maximum response minus the baseline; a^1 is the 5 EC_{50} ; and a^2 is the maximum response.

Peptide A stimulation of nGPCR-51 expressing COS 7 cells resulted in a dose dependent increase in aequorin luminescence with an EC_{50} of approximately 40 nM and a maximum response of 52-fold over baseline.

C. *Luciferase Reporter Gene Assay*

10 The photoprotein luciferase provides another useful tool for assaying for modulators of nGPCR-x activity. Cells (e.g., CHO cells or COS 7 cells) are transiently co-transfected with both a nGPCR-x expression construct (e.g., nGPCR-x in pzeoSV2) and a reporter construct which includes a gene for the luciferase protein downstream from a transcription factor binding site, such as the cAMP-response element (CRE), AP-1, or NF-kappa B. Agonist binding to 15 receptors coupled to the G_s subtype of G proteins leads to increases in cAMP, thereby activating the CRE transcription factor and resulting in expression of the luciferase gene. Agonist binding to receptors coupled to the G_q subtype of G protein leads to production of diacylglycerol that activates protein kinase C, which activates the AP-1 or NF-kappa B transcription factors, in turn resulting in expression of the luciferase gene. Expression levels of luciferase reflect the 20 activation status of the signaling events. (See generally, George *et al.*, *Journal of Biomolecular Screening*, 2(4): 235-240 (1997); and Stratowa *et al.*, *Current Opinion in Biotechnology* 6: 574-581 (1995)). Luciferase activity may be quantitatively measured using, e.g., luciferase assay reagents that are commercially available from Promega (Madison, WI).

In one exemplary assay, CHO cells are plated in 24-well culture dishes at a density of 25 100,000 cells/well one day prior to transfection and cultured at 37C in MEM (Gibco/BRL) supplemented with 10% fetal bovine serum, 2 mM glutamine, 10 U/ml penicillin and 10 μ g/ml streptomycin. Cells are transiently co-transfected with both a nGPCR-x expression construct and a reporter construct containing the luciferase gene. The reporter plasmids CRE-luciferase, AP-1-luciferase and NF-kappaB-luciferase may be purchased from Stratagene (LaJolla, CA). 30 Transfections are performed using the FuGENE 6 transfection reagent (Boehringer-Mannheim) according to the supplier's instructions. Cells transfected with the reporter construct alone are used as a control. Twenty-four hours after transfection, cells are washed once with PBS pre-warmed to 37C. Serum-free MEM is then added to the cells either alone (control) or with one or more candidate modulators and the cells are incubated at 37C for five hours. Thereafter,

cells are washed once with ice-cold PBS and lysed by the addition of 100 μ l of lysis buffer per well from the luciferase assay kit supplied by Promega. After incubation for 15 minutes at room temperature, 15 μ l of the lysate is mixed with 50 μ l of substrate solution (Promega) in an opaque-white, 96-well plate, and the luminescence is read immediately on a Wallace model 5 1450 MicroBeta scintillation and luminescence counter (Wallace Instruments, Gaithersburg, MD).

Differences in luminescence in the presence versus the absence of a candidate modulator compound are indicative of modulatory activity. Receptors that are either constitutively active or activated by agonists typically give a 3 to 20-fold stimulation of luminescence compared to cells 10 transfected with the reporter gene alone. Modulators that act as inverse agonists will reverse this effect.

nGPCR-51

nGPCR-51 was transfected in N7 cells, stable HOS (human osteosarcoma) cells expressing NFkB-Luciferase reporter gene Briefly, LipofectAMINE, PLUS reagent, and 4 μ g 15 DNA were added to cells in Opti-MEM. After 4 hours, serum-containing medium was added to each plate. Twenty-four hour after transfection the cells were trypsinized and seeded in COSTAR white 96-well plate at a cell density of about 20,000 cells per well. Another 24 hours later the culture medium was replaced with fresh growth medium and Peptide A was added. After 5-hr incubation with Peptide A, the medium was aspirated and 100 μ L diluted Steady Glo 20 was added. The plates were gently shake at room temperature for 30 minutes and luminescence determined by TopCount at the SPC mode.

The dose-response effect of Peptide A on luciferase expression in Seq51-transfected N7 cells was tested in 2 separate experiments. The EC₅₀ value of Peptide A for the 2 experiments was 1.6 nM and 1.7 nM, respectively. Salmon Peptide B (Asp Thr Met Arg Cys Met Val Gly 25 Arg Val Tyr Arg Pro Cys Trp Glu Val; SEQ ID NO:184), a variant of Peptide A, and another variant of Peptide A were also tested in the second experiment. The EC₅₀ value for salmon Peptide B was approximately 25 nM while variant of Peptide A had no effect on these Seq51-transfected cells at concentration up to 1 μ M.

Peptide A, a cyclic heptadecapeptide, is a prominent hypothalamic neuropeptide 30 expressed in cells which are part of an extensive network of neurons projecting to multiple parts of the brain. Despite the large numbers of hypothalamic Peptide A-expressing cells, the Peptide A cell groups are not part of the major hypothalamic nuclei (paraventricular, supraoptic, dorsomedial, ventromedial, arcuate, lateral tuberal or mammillary) (Knigge *et al.*, *J. Peptides*, 1996, 17, 1063-1073), but they have terminal projections to most of these hypothalamic nuclei.

Additionally, Peptide A projections from the hypothalamus to the rest of the brain are extensive, including to the cerebellar cortex, subcortical nuclei, limbic areas, thalamus, brain stem and spinal cord.

Peptide A projections are included in the circuitry involved in processing visual and
5 auditory information, suggesting a general role in arousal and sensorimotor integration (Hervieu
et al., *Eur. J. Neurosci.*, 2000, 12, 1194-1216). Intracerebroventricular injection of Peptide A
causes a decrease in the conditioning amplitude of the second of a pair of auditory evoked
potentials recorded from the hippocampus (hippocampal auditory gating paradigm) and reverses
the increases in the conditioning amplitude induced by alpha MSH (Miller *et al.*, *Peptides*, 1993,
10 14, 431-440). These data would suggest that Peptide A inhibits the normal auditory gating
process. Schizophrenics also present with a loss of auditory gating which may contribute to an
inability to filter inappropriate information. Dysregulation of Peptide A may contribute to this
phenotype in humans and antagonists active at the Peptide A receptors may effectively treat
these deficits in schizophrenia.

15 Major portions of the Peptide A cell group are in areas considered to be part of the
extrapyramidal motor circuits. It has been suggested that this location would afford the Peptide
A neuronal system the capability of coordinating hypothalamic visceral activity with appropriate
motor activity (Knigge *et al.*, 1996).

Peptide A localization in the hippocampal formation, the amygdaloid regions, the bed
20 nucleus of the stria terminalis all suggest a role in cognition and learning and in support of this
Peptide A has been shown to alter passive avoidance performance (McBride *et al.*, *Peptides*,
1994, 15, 757-759).

Central injections of Peptide A in rats leads to a reduction in peak circadian ACTH
levels and inhibit stress-induced ACTH secretion (Bluet Pajot *et al.*, *J. Neuroendocrinol.*, 1995,
25 7, 297-303; and Ludwig *et al.*, *Am. J. Physiology*, 1998, 274, E627-E633). However, injections
of Peptide A into the medial preoptic area of the hypothalamus of rats result in decreases in time
spent in open arms in the elevated plus maze assay indicative of increased anxiety. Injections of
the peptide also stimulated female sexual receptivity (Gonzalez *et al.*, *Peptides*, 1996, 17, 171-
177).

30 The lateral hypothalamus has a prominent role in feeding-related behaviors and in fact
Peptide A has been shown to be involved in energy balance and food intake. Ob/ob mice express
more Peptide A mRNA compared with lean littermates (Qu *et al.*, *Nature*, 1996, 380, 243-247).
Additionally, fasting increases hypothalamic Peptide A mRNA levels (Qu *et al.*, 1996).

Intracerebroventricular injection of Peptide A leads to an increase in food intake in rats (Qu *et al.*, *Endocrinology*, 1997, 138, 351-355).

Targeted deletion of the gene which encodes the preprohormone for Peptide A resulted in mice which have reduced body weight and are lean due to reduced feeding. Additionally these mice have an increased metabolic rate in the presence of decreased leptin levels and POMC levels (Shimada *et al.*, *Nature*, 1998, 396, 670-673). These mice also have a loss of expression of the NEI peptide as well as Peptide A. Nevertheless, the prominent phenotypic feature of these mice is their reduced size. Antagonists of Peptide A receptors may be effective in the treatment of obesity.

The SLC-1 gene product was recently identified as a G-protein coupled receptor which is potently activated by Peptide A (Chambers *et al.*, *Nature*, 1999, 400, 261-265; and Saito *et al.*, *Nature*, 1999, 400, 265-269). nGPCR-51 has been identified as a gene encoding a second Peptide A receptor which is also potently activated by Peptide A. Like that of the SLC-1 gene, expression of nGPCR-51 is extensive throughout the brain. The multiple actions of Peptide A could be differentially mediated by each of the two Peptide A receptors. Selective agonism or antagonism at nGPCR-51 could result in a therapeutic effective in treating schizophrenia, attention disorders, anxiety, depression, obesity, and the like.

D. Intracellular calcium measurement using FLIPR

Changes in intracellular calcium levels are another recognized indicator of G protein-coupled receptor activity, and such assays can be employed to screen for modulators of nGPCR-x activity. For example, CHO cells stably transfected with a nGPCR-x expression vector are plated at a density of 4×10^4 cells/well in Packard black-walled, 96-well plates specially designed to discriminate fluorescence signals emanating from the various wells on the plate. The cells are incubated for 60 minutes at 37C in modified Dulbecco's PBS (D-PBS) containing 36 mg/L pyruvate and 1 g/L glucose with the addition of 1% fetal bovine serum and one of four calcium indicator dyes (Fluo-3TM AM, Fluo-4TM AM, Calcium GreenTM-1 AM, or Oregon GreenTM 488 BAPTA-1 AM), each at a concentration of 4 μ M. Plates are washed once with modified D-PBS without 1% fetal bovine serum and incubated for 10 minutes at 37C to remove residual dye from the cellular membrane. In addition, a series of washes with modified D-PBS without 1% fetal bovine serum is performed immediately prior to activation of the calcium response.

A calcium response is initiated by the addition of one or more candidate receptor agonist compounds, calcium ionophore A23187 (10 μ M; positive control), or ATP (4 μ M; positive control). Fluorescence is measured by Molecular Device's FLIPR with an argon laser (excitation

at 488 nm). (See, e.g., Kuntzweiler et al., *Drug Development Research*, 44(1):14-20 (1998)). The F-stop for the detector camera was set at 2.5 and the length of exposure was 0.4 milliseconds. Basal fluorescence of cells was measured for 20 seconds prior to addition of candidate agonist, ATP, or A23187, and the basal fluorescence level was subtracted from the 5 response signal. The calcium signal is measured for approximately 200 seconds, taking readings every two seconds. Calcium ionophore A23187 and ATP increase the calcium signal 200% above baseline levels. In general, activated GPCRs increase the calcium signal approximately 10-15% above baseline signal.

nGPCR-51

10 HEK293 cells were transiently transfected with an expression vector for nGPCR-51 and empty vector using Lipofectamine plus (Gibco) according to the manufacturer's instructions. The next day, the cells were seeded into 96-well plates at 25,000 cells per well. The following day, cells were loaded with 1 μ M Fluo-4-acetoxymethyl fluorescent indicator dye (Molecular Probes) in MEM (minimal essential media) containing 0.1% bovine serum albumin, 0.04% 15 pluronic acid and 2.5 mM probenecid for 30 minutes at 37C. The cells were washed with pre-warmed (37C) assay buffer (Hanks buffer containing 15 mM HEPES, 2.5 mM probenecid and 0.1% bovine serum albumin). Assay buffer (100 μ l) was added to each well and plates were incubated at 37C for 15 minutes. Various concentrations (0.03 pM-10 nM) of human Peptide A or salmon Peptide B were added and fluorescence produced by fluo-4 (a calcium sensitive dye) 20 was measured every second for 150 seconds on a fluorometric imaging plate reader (FLIPR1; Molecular Devices). Human Peptide A and salmon Peptide B-induced calcium mobilization in nGPCR-51-transfected cells with EC₅₀ values of 3 nM and 6 nM, respectively. Human Peptide A and salmon Peptide B had no effect on vector-transfected cells.

E. Mitogenesis Assay

25 In a mitogenesis assay, the ability of candidate modulators to induce or inhibit nGPCR-x-mediated cell division is determined. (See, e.g., Lajiness et al., *Journal of Pharmacology and Experimental Therapeutics* 267(3): 1573-1581 (1993)). For example, CHO cells stably expressing nGPCR-x are seeded into 96-well plates at a density of 5000 cells/well and grown at 37°C in MEM with 10% fetal calf serum for 48 hours, at which time the cells are rinsed twice 30 with serum-free MEM. After rinsing, 80 μ l of fresh MEM, or MEM containing a known mitogen, is added along with 20 μ l MEM containing varying concentrations of one or more candidate modulators or test compounds diluted in serum-free medium. As controls, some wells on each plate receive serum-free medium alone, and some receive medium containing 10% fetal

bovine serum. Untransfected cells or cells transfected with vector alone also may serve as controls.

After culture for 16-18 hours, 1 μ Ci of [3 H]-thymidine (2 Ci/mmol) is added to the wells and cells are incubated for an additional 2 hours at 37C. The cells are trypsinized and collected 5 on filter mats with a cell harvester (Tomtec); the filters are then counted in a Betaplate counter. The incorporation of [3 H]-thymidine in serum-free test wells is compared to the results achieved in cells stimulated with serum (positive control). Use of multiple concentrations of test compounds permits creation and analysis of dose-response curves using the non-linear, least squares fit equation: $A = B \times [C / (D + C)] + G$ where A is the percent of serum stimulation; B is 10 the maximal effect minus baseline; C is the EC₅₀; D is the concentration of the compound; and G is the maximal effect. Parameters B, C and G are determined by Simplex optimization.

Agonists that bind to the receptor are expected to increase [3 H]-thymidine incorporation into cells, showing up to 80% of the response to serum. Antagonists that bind to the receptor will inhibit the stimulation seen with a known agonist by up to 100%.

15 **F. [35 S]GTP γ S Binding Assay**

Because G protein-coupled receptors signal through intracellular G proteins whose activity involves GTP binding and hydrolysis to yield bound GDP, measurement of binding of the non-hydrolyzable GTP analog [35 S]GTP γ S in the presence and absence of candidate modulators provides another assay for modulator activity. (See, e.g., Kowal et al., 20 *Neuropharmacology* 37:179-187 (1998).)

In one exemplary assay, cells stably transfected with a nGPCR-x expression vector are grown in 10 cm tissue culture dishes to subconfluence, rinsed once with 5 ml of ice-cold 25 $\text{Ca}^{2+}/\text{Mg}^{2+}$ -free phosphate-buffered saline, and scraped into 5 ml of the same buffer. Cells are pelleted by centrifugation (500 x g, 5 minutes), resuspended in TEE buffer (25 mM Tris, pH 7.5, 5 mM EDTA, 5 mM EGTA), and frozen in liquid nitrogen. After thawing, the cells are homogenized using a Dounce homogenizer (one ml TEE per plate of cells), and centrifuged at 1,000 x g for 5 minutes to remove nuclei and unbroken cells.

The homogenate supernatant is centrifuged at 20,000 x g for 20 minutes to isolate the membrane fraction, and the membrane pellet is washed once with TEE and resuspended in 30 binding buffer (20 mM HEPES, pH 7.5, 150 mM NaCl, 10 mM MgCl₂, 1 mM EDTA). The resuspended membranes can be frozen in liquid nitrogen and stored at -70C until use.

Aliquots of cell membranes prepared as described above and stored at -70C are thawed, homogenized, and diluted into buffer containing 20 mM HEPES, 10 mM MgCl₂, 1 mM EDTA, 120 mM NaCl, 10 μ M GDP, and 0.2 mM ascorbate, at a concentration of 10-50 μ g/ml. In a final

volume of 90 μ l, homogenates are incubated with varying concentrations of candidate modulator compounds or 100 μ M GTP for 30 minutes at 30C and then placed on ice. To each sample, 10 μ l guanosine 5'-O-(3[³⁵S]thio) triphosphate (NEN, 1200 Ci/mmol; [³⁵S]-GTP γ S), was added to a final concentration of 100-200 pM. Samples are incubated at 30C for an additional 30 minutes, 5 1 ml of 10mM HEPES, pH 7.4, 10 mM MgCl₂, at 4C is added and the reaction is stopped by filtration.

Samples are filtered over Whatman GF/B filters and the filters are washed with 20 ml ice-cold 10 mM HEPES, pH 7.4, 10 mM MgCl₂. Filters are counted by liquid scintillation 10 spectroscopy. Nonspecific binding of [³⁵S]-GTP γ S is measured in the presence of 100 μ M GTP and subtracted from the total. Compounds are selected that modulate the amount of [³⁵S]-GTP γ S binding in the cells, compared to untransfected control cells. Activation of receptors by agonists gives up to a five-fold increase in [³⁵S]GTP γ S binding. This response is blocked by antagonists.

G. *MAP Kinase Activity Assay*

Evaluation of MAP kinase activity in cells expressing a GPCR provides another assay to 15 identify modulators of GPCR activity. (See, e.g., Lajiness et al., *Journal of Pharmacology and Experimental Therapeutics* 267(3):1573-1581 (1993) and Boulton et al., *Cell* 65:663-675 (1991).)

In one embodiment, CHO cells stably transfected with nGPCR-x are seeded into 6-well plates at a density of 70,000 cells/well 48 hours prior to the assay. During this 48-hour period, 20 the cells are cultured at 37C in MEM medium supplemented with 10% fetal bovine serum, 2mM glutamine, 10 U/ml penicillin and 10 μ g/ml streptomycin. The cells are serum-starved for 1-2 hours prior to the addition of stimulants.

For the assay, the cells are treated with medium alone or medium containing either a candidate agonist or 200 nM Phorbol ester- myristoyl acetate (i.e., PMA, a positive control), and 25 the cells are incubated at 37C for varying times. To stop the reaction, the plates are placed on ice, the medium is aspirated, and the cells are rinsed with 1 ml of ice-cold PBS containing 1mM EDTA. Thereafter, 200 μ l of cell lysis buffer (12.5 mM MOPS, pH 7.3, 12.5 mM glycerophosphate, 7.5mM MgCl₂, 0.5mM EGTA, 0.5 mM sodium vanadate, 1mM benzamidine, 1mM dithiothreitol, 10 μ g/ml leupeptin, 10 μ g/ml aprotinin, 2 μ g/ml pepstatin A, and 1 μ M 30 okadaic acid) is added to the cells. The cells are scraped from the plates and homogenized by 10 passages through a 23 3/4 G needle, and the cytosol fraction is prepared by centrifugation at 20,000 x g for 15 minutes.

Aliquots (5-10 μ l containing 1-5 μ g protein) of cytosol are mixed with 1 mM MAPK Substrate Peptide (APRTPGGRR; SEQ ID NO:182), Upstate Biotechnology, Inc., N.Y.) and

50 μ M [γ -³²P]ATP (NEN, 3000 Ci/mmol), diluted to a final specific activity of ~2000 cpm/pmol, in a total volume of 25 μ l. The samples are incubated for 5 minutes at 30C, and reactions are stopped by spotting 20 μ l on 2 cm² squares of Whatman P81 phosphocellulose paper. The filter squares are washed in 4 changes of 1% H₃PO₄, and the squares are subjected to liquid 5 scintillation spectroscopy to quantitate bound label. Equivalent cytosolic extracts are incubated without MAPK substrate peptide, and the bound label from these samples are subtracted from the matched samples with the substrate peptide. The cytosolic extract from each well is used as a separate point. Protein concentrations are determined by a dye binding protein assay (Bio-Rad Laboratories). Agonist activation of the receptor is expected to result in up to a five-fold 10 increase in MAPK enzyme activity. This increase is blocked by antagonists.

H. ³H]Arachidonic Acid Release

The activation of GPCRs also has been observed to potentiate arachidonic acid release in cells, providing yet another useful assay for modulators of GPCR activity. (See, e.g., Kanterman et al., Molecular Pharmacology 39:364-369 (1991).) For example, CHO cells that 15 are stably transfected with a nGPCR-x expression vector are plated in 24-well plates at a density of 15,000 cells/well and grown in MEM medium supplemented with 10% fetal bovine serum, 2 mM glutamine, 10 U/ml penicillin and 10 μ g/ml streptomycin for 48 hours at 37C before use. Cells of each well are labeled by incubation with [³H]-arachidonic acid (Amersham Corp., 210 Ci/mmol) at 0.5 μ Ci/ml in 1 ml MEM supplemented with 10mM HEPES, pH 7.5, and 0.5% 20 fatty-acid-free bovine serum albumin for 2 hours at 37C. The cells are then washed twice with 1 ml of the same buffer.

Candidate modulator compounds are added in 1 ml of the same buffer, either alone or with 10 μ M ATP and the cells are incubated at 37C for 30 minutes. Buffer alone and mock-transfected cells are used as controls. Samples (0.5 ml) from each well are counted by liquid 25 scintillation spectroscopy. Agonists which activate the receptor will lead to potentiation of the ATP-stimulated release of [³H]-arachidonic acid. This potentiation is blocked by antagonists.

I. Extracellular Acidification Rate

In yet another assay, the effects of candidate modulators of nGPCR-x activity are assayed by monitoring extracellular changes in pH induced by the test compounds. (See, e.g., 30 Dunlop et al., Journal of Pharmacological and Toxicological Methods 40(1):47-55 (1998).) In one embodiment, CHO cells transfected with a nGPCR-x expression vector are seeded into 12 mm capsule cups (Molecular Devices Corp.) at 4 x 10⁵ cells/cup in MEM supplemented with 10% fetal bovine serum, 2 mM L-glutamine, 10 U/ml penicillin, and 10 μ g/ml streptomycin. The cells are incubated in this medium at 37C in 5% CO₂ for 24 hours.

Extracellular acidification rates are measured using a Cytosensor microphysiometer (Molecular Devices Corp.). The capsule cups are loaded into the sensor chambers of the microphysiometer and the chambers are perfused with running buffer (bicarbonate-free MEM supplemented with 4 mM L-glutamine, 10 units/ml penicillin, 10 μ g/ml streptomycin, 26 mM NaCl) at a flow rate of 100 μ l/minute. Candidate agonists or other agents are diluted into the running buffer and perfused through a second fluid path. During each 60-second pump cycle, the pump is run for 38 seconds and is off for the remaining 22 seconds. The pH of the running buffer in the sensor chamber is recorded during the cycle from 43-58 seconds, and the pump is re-started at 60 seconds to start the next cycle. The rate of acidification of the running buffer during the recording time is calculated by the Cytosoft program. Changes in the rate of acidification are calculated by subtracting the baseline value (the average of 4 rate measurements immediately before addition of a modulator candidate) from the highest rate measurement obtained after addition of a modulator candidate. The selected instrument detects 61 mV/pH unit. Modulators that act as agonists of the receptor result in an increase in the rate of extracellular acidification compared to the rate in the absence of agonist. This response is blocked by modulators which act as antagonists of the receptor.

J. Radioligand Binding Assay

HEK 293 or COS7 cells transiently expressing nGPCR-51, or CHO K-1 cells stable expressing nGPCR-51, were grown to sub-confluence (2 days post-transfection for transients), harvested from flasks in Dulbecco's PBS and pelleted by low speed centrifugation (2500 rpm) for 10 minutes. Cell pellets were homogenized in 10 ml tissue buffer (50 mM Hepes, 10 mM MgCl₂, 2 mM EGTA, 1 μ g/ml aprotinin, 1 μ g/ml leupeptin hemisulfate, 1 μ g/ml pepstatin A, pH 7.0) using a dounce, 10 strokes. Homogenate was centrifuged at 47,000 x g for 15 minutes. Membrane pellet was resuspended in 1 ml tissue buffer using the dounce, 10 strokes. An aliquot of the membrane preparation was used to determine protein concentration. For measurement of saturation binding, 10 μ g of cell membranes were incubated with various concentrations of [¹²⁵I] Peptide A (iodinated by routine procedures via the Tyr residue) in 300 μ l binding assay buffer (tissue buffer plus 0.15 mM Bacitracin and 0.1% ovalbumin) for 90 minutes at room temperature in 96-well plates. Non-specific binding was defined by the inclusion of 1 μ M Peptide A. After the binding incubation, plates were harvested onto GF/C filters presoaked in 0.3% non-fat dry milk. Filters were dried, and 30 μ l scintillant was added to each well. Filter plates were then counted in a Packard Topcount™ 96-well microplate scintillation counter. Data were analyzed using GraphPad Prism (San Diego, CA) and Kd and Bmax values were calculated.

In a saturation binding experiment using COS 7 cells transiently expressing nGPCR-51 the Kd for [¹²⁵I] Peptide A was determined to be 0.254 nM, with a Bmax of 75 fmol/mg. HEK 293 cells transiently expressing nGPCR-51 were shown to have an estimated Bmax of 108 fmol/mg. Untransfected cells have no specific radioligand binding for [¹²⁵I] Peptide A.

5 **Example 13: Using nGPCR-x Proteins to Isolate Neurotransmitters**

The isolated nGPCR-x proteins, particularly nGPCR-42, 46, 48, 49, 51, 52, 61, 63, or 70 (SEQ ID Numbers 61, 62, 68, 91, 94, 96, 97, 99, 100, and 111-120) can be used to isolate novel or known neurotransmitters (Saito et al., *Nature*, 400: 265-269, 1999). The cDNAs that encode the isolated nGPCR-x can be cloned into mammalian expression vectors and used to stably or 10 transiently transfet mammalian cells including CHO, Cos or HEK293 cells. Receptor expression can be determined by Northern blot analysis of transfected cells and identification of an appropriately sized mRNA band (predicted size from the cDNA). Brain regions shown by mRNA analysis to express each of the nGPCR-x proteins could be processed for peptide extraction using any of several protocols ((Reinscheidk R.K. et al., *Science* 270: 243-247, 1996; 15 Sakurai, T., et al., *Cell* 92; 573-585, 1998; Hinuma, S., et al., *Nature* 393: 272-276, 1998). Chromotographic fractions of brain extracts could be tested for ability to activate nGPCR-x proteins by measuring second messenger production such as changes in cAMP production in the presence or absence of forskolin, changes in inositol 3-phosphate levels, changes in intracellular calcium levels or by indirect measures of receptor activation including receptor stimulated 20 mitogenesis, receptor mediated changes in extracellular acidification or receptor mediated changes in reporter gene activation in response to cAMP or calcium (these methods should all be referenced in other sections of the patent). Receptor activation could also be monitored by co-transfeting cells with a chimeric G_i_{q/13} to force receptor coupling to a calcium stimulating pathway (Conklin et al., *Nature* 363; 274-276, 1993). Neurotransmitter mediated activation of 25 receptors could also be monitored by measuring changes in [³⁵ S]-GTPKS binding in membrane fractions prepared from transfected mammalian cells. This assay could also be performed using baculoviruses containing nGPCR-x proteins infected into SF9 insect cells.

The neurotransmitter which activates nGPCR-x proteins can be purified to homogeneity through successive rounds of purification using nGPCR-x proteins activation as a measurement 30 of neurotransmitter activity. The composition of the neurotransmitter can be determined by mass spectrometry and Edman degradation if peptidergic. Neurotransmitters isolated in this manner will be bioactive materials which will alter neurotransmission in the central nervous system and will produce behavioral and biochemical changes.

Example 14: Using nGPCR-x Proteins to Isolate and Purify G Proteins

cDNAs encoding nGPCR-x proteins are epitope-tagged at the amino terminus end of the cDNA with the cleavable influenza-hemagglutinin signal sequence followed by the FLAG epitope (IBI, New Haven, CT). Additionally, these sequences are tagged at the carboxyl

5 terminus with DNA encoding six histidine residues. (Amino and Carboxyl Terminal Modifications to Facilitate the Production and Purification of a G Protein-Coupled Receptor, B.K. Kobilka, Analytical Biochemistry, Vol. 231, No. 1, Oct 1995, pp. 269-271). The resulting sequences are cloned into a baculovirus expression vector such as pVL1392 (Invitrogen). The baculovirus expression vectors are used to infect SF-9 insect cells as described (Guan *et al.*, 10 (1992) J. Biol. Chem. 267, 21995-21998). Infected SF-9 cells could be grown in 1000-ml cultures in SF900 II medium (Life Technologies, Inc.) containing 5% fetal calf serum (Gemini, Calabasas, CA) and 0.1 mg/ml gentamicin (Life Technologies, Inc.) for 48 hours at which time the cells could be harvested. Cell membrane preparations could be separated from soluble proteins following cell lysis. nGPCR-x protein purification is carried out as described for 15 purification of the 92 receptor (Kobilka, Anal. Biochem., 231 (1): 269-271, 1995) including solubilization of the membranes in 0.8-1.0 % n-dodecyl-D-maltoside (DM) (CalBiochem, La Jolla, CA) in buffer containing protease inhibitors followed by Ni-column chromatography using chelating Sepharose™ (Pharmacia, Uppsala, Sweden). The eluate from the Ni-column is further purified on an M1 anti-FLAG antibody column (IBI). Receptor containing fractions are 20 monitored by using receptor specific antibodies following western blot analysis or by SDS-PAGE analysis to look for an appropriate sized protein band (appropriate size would be the predicted molecular weight of the protein). This method of purifying G protein is particularly useful to isolate G proteins that bind to the nGPCR-x proteins in the absence of an activating ligand.

25 Some of the preferred embodiments of the invention described above are outlined below and include, but are not limited to, the following embodiments. As those skilled in the art will appreciate, numerous changes and modifications may be made to the preferred embodiments of the invention without departing from the spirit of the invention. It is intended that all such variations fall within the scope of the invention. The entire disclosure of each publication cited 30 herein is incorporated herein by reference in its entirety.

Example 15: Clone Deposit Information

In accordance with the Budapest Treaty, clones of the present invention have been deposited at the Agricultural Research Culture Collection (NRRL) International Depository

Authority, 1815 N. University Street, Peoria, Illinois 61604, U.S.A. Accession numbers and deposit dates are provided below in Table 6.

Table 6: Deposit Information

Clone	SEQ ID NO:	NRRL Accession Number	Budapest Treaty Deposit Date
5	nGPCR-42	31, 54	B-30246
	nGPCR-48	36,56	B30263
	nGPCR-46	34, 55	B-30269
10	nGPCR-52	40, 58	B-30270
	nGPCR-70	8, 52	B-30300
	nGPCR-61	1, 60	B-30301
	nGPCR-49	37, 59	B-30313
	nGPCR-51	39, 57	B-30314
15	nGPCR-63	3, 51, 53	B-30315

What is claimed is:

1. An isolated nucleic acid molecule comprising a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence homologous to sequences selected from the group consisting of: SEQ ID NO:61 to SEQ ID NO:120, said nucleic acid molecule encoding at least a portion of nGPCR-x.
5
2. The isolated nucleic acid molecule of claim 1 comprising a sequence that encodes a polypeptide comprising a sequence selected from the group consisting of SEQ ID NO:61 to SEQ ID NO:120.
10
3. The isolated nucleic acid molecule of claim 1 comprising a sequence homologous to a sequence selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:60.
15
4. The isolated nucleic acid molecule of claim 1 comprising a sequence selected from the group of sequences consisting of SEQ ID NO:1 to SEQ ID NO:60.
20
5. The isolated nucleic acid molecule of claim 4 comprising a sequence selected from the group of sequences consisting of SEQ ID NO:1 to SEQ ID NO:60.
25
6. The isolated nucleic acid molecule of claim 4 wherein said nucleotide sequence is selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:8, SEQ ID NO:31, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, and SEQ ID NO:60.
30
7. The isolated nucleic acid molecule of claim 1 wherein said nucleic acid molecule is DNA.
35
8. The isolated nucleic acid molecule of claim 1 wherein said nucleic acid molecule is RNA.
40
9. An expression vector comprising a nucleic acid molecule of any one of claims 1 to 5.
45

10. The expression vector of claim 9 wherein said nucleic acid molecule comprises a sequence selected from the group of sequences consisting of SEQ ID NO:1 to SEQ ID NO:60.

11. The expression vector of claim 9 wherein said nucleic acid molecule comprises a 5 nucleotide sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:8, SEQ ID NO:31, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, and SEQ ID NO:60.

10 12. The expression vector of claim 9 wherein said vector is a plasmid.

13. The expression vector of claim 9 wherein said vector is a viral particle.

14. The expression vector of claim 13 wherein said vector is selected from the group 15 consisting of adenoviruses, baculoviruses, parvoviruses, herpesviruses, poxviruses, adeno-associated viruses, Semliki Forest viruses, vaccinia viruses, and retroviruses.

15. The expression vector of claim 9 wherein said nucleic acid molecule is operably connected to a promoter selected from the group consisting of simian virus 40, mouse mammary 20 tumor virus, long terminal repeat of human immunodeficiency virus, maloney virus, cytomegalovirus immediate early promoter, Epstein Barr virus, rous sarcoma virus, human actin, human myosin, human hemoglobin, human muscle creatine, and human metallothionein.

16. A host cell transformed with an expression vector of claim 9.

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17. The transformed host cell of claim 16 wherein said cell is a bacterial cell.

18. The transformed host cell of claim 17 wherein said bacterial cell is *E. coli*.

30 19. The transformed host cell of claim 16 wherein said cell is yeast.

20. The transformed host cell of claim 19 wherein said yeast is *S. cerevisiae*.

21. The transformed host cell of claim 16 wherein said cell is an insect cell.

22. The transformed host cell of claim 21 wherein said insect cell is *S. frugiperda*.

23. The transformed host cell of claim 16 wherein said cell is a mammalian cell.

5 24. The transformed host cell of claim 23 wherein mammalian cell is selected from the group consisting of chinese hamster ovary cells, HeLa cells, African green monkey kidney cells, human 293 cells, and murine 3T3 fibroblasts.

10 25. An isolated nucleic acid molecule comprising a nucleotide sequence complementary to at least a portion of a sequence selected from the group of sequences consisting of SEQ ID NO:1 to SEQ ID NO:60, said portion comprising at least 10 nucleotides.

15 26. The nucleic acid molecule of claim 25 wherein said molecule is an antisense oligonucleotide directed to a region of a sequence selected from the group of sequences consisting of SEQ ID NO:1 to SEQ ID NO:60.

20 27. The nucleic acid molecule of claim 26 wherein said oligonucleotide is directed to a regulatory region of a sequence selected from the group of sequences consisting of SEQ ID NO:1 to SEQ ID NO:60.

25 28. The nucleic acid molecule of claim 25 wherein said molecule is an antisense oligonucleotide directed to a region of nucleotide sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:8, SEQ ID NO:31, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, and SEQ ID NO:60.

30 29. A composition comprising a nucleic acid molecule of any one of claims 1 to 5 or 25 and an acceptable carrier or diluent.

30 30. A composition comprising a recombinant expression vector of claim 9 and an acceptable carrier or diluent.

31. A method of producing a polypeptide that comprises a sequence selected from the group of sequences consisting SEQ ID NO:61 to SEQ ID NO:120, and homologs thereof, said method comprising the steps of:

- 5 a) introducing a recombinant expression vector of claim 10 into a compatible host cell;
- b) growing said host cell under conditions for expression of said polypeptide; and
- c) recovering said polypeptide.

32. The method of claim 31 wherein said host cell is lysed and said polypeptide is recovered from the lysate of said host cell.

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33. The method of claim 31 wherein said polypeptide is recovered by purifying the culture medium without lysing said host cell.

34. An isolated polypeptide encoded by a nucleic acid molecule of claim 1.

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35. The polypeptide of claim 34 wherein said polypeptide comprises a sequence selected from the group of sequences consisting of SEQ ID NO:61 to SEQ ID NO:120.

36. The polypeptide of claim 34 wherein said polypeptide comprises an amino acid sequence 20 homologous to a sequence selected from the group of sequences consisting of SEQ ID NO:61 to SEQ ID NO:120.

37. The polypeptide of claim 34 wherein said sequence homologous to a sequence selected 25 from the group of sequences consisting of SEQ ID NO:61 to SEQ ID NO:120 comprises at least one conservative amino acid substitution compared to the sequences in the group of sequences consisting of SEQ ID NO:61 to SEQ ID NO:120.

38. The polypeptide of claim 34 wherein said polypeptide comprises an allelic variant of a 30 polypeptide with a sequence selected from the group of sequences consisting of SEQ ID NO:61 to SEQ ID NO:120.

39. The polypeptide of claim 34 wherein said polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:68, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:100,

SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, and SEQ ID NO:120.

40. A composition comprising a polypeptide of claim 34 and an acceptable carrier or diluent.

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41. An isolated antibody which binds to an epitope on a polypeptide of claim 34.

42. The antibody of claim 41 wherein said antibody is a monoclonal antibody.

10 43. A composition comprising an antibody of claim 41 and an acceptable carrier or diluent.

44. A method of inducing an immune response in a mammal against a polypeptide of claim 34 comprising administering to said mammal an amount of said polypeptide sufficient to induce said immune response.

15

45. A method for identifying a compound which binds nGPCR-x comprising the steps of:
a) contacting nGPCR-x with a compound; and
b) determining whether said compound binds nGPCR-x.

20 46. The method of claim 45 wherein the nGPCR-x comprises an amino acid sequence selected from the group consisting of SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:68, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, and SEQ ID NO:120.

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47. The method of claim 45 wherein binding of said compound to nGPCR-x is determined by a protein binding assay.

30 48. The method of claim 45 wherein said protein binding assay is selected from the group consisting of a gel-shift assay, Western blot, radiolabeled competition assay, phage-based expression cloning, co-fractionation by chromatography, co-precipitation, cross linking, interaction trap/two-hybrid analysis, southwestern analysis, and ELISA.

49. A compound identified by the method of claim 45.

50. A method for identifying a compound which binds a nucleic acid molecule encoding nGPCR-x comprising the steps of:

- a) contacting said nucleic acid molecule encoding nGPCR-x with a compound; and
- b) determining whether said compound binds said nucleic acid molecule.

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51. The method of claim 50 wherein binding is determined by a gel-shift assay.

52. A compound identified by the method of claim 50.

10 53. A method for identifying a compound which modulates the activity of nGPCR-x comprising the steps of:

- a) contacting nGPCR-x with a compound; and
- b) determining whether nGPCR-x activity has been modulated.

15 54. The method of claim 53 wherein the nGPCR-x comprises an amino acid sequence selected from the group consisting of SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:68, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, and SEQ ID NO:120.

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55. The method of claim 53 wherein said activity is neuropeptide binding.

56. The method of claim 53 wherein said activity is neuropeptide signaling.

25 57. A compound identified by the method of claim 53.

58. A method of identifying an animal homolog of nGPCR-x comprising the steps:

- a) comparing the nucleic acid sequences of the animal with a sequence selected from the group of sequence consisting of SEQ ID NO:1 to SEQ ID NO:60, and portions thereof, said portions being at least 10 nucleotides; and
- b) identifying nucleic acid sequences of the animal that are homologous to said sequence selected from the group sequence consisting of SEQ ID NO:1 to SEQ ID NO:60, and portions thereof.

59. The method of claim 58 wherein comparing the nucleic acid sequences of the animal with a sequence selected from the group of sequences consisting of SEQ ID NO:1 to SEQ ID NO:60, and portions thereof, said portions being at least 10 nucleotides, is performed by DNA hybridization.

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60. The method of claim 58 wherein comparing the nucleic acid sequences of the animal with a sequence selected from the group of sequences consisting of SEQ ID NO:1 to SEQ ID NO:60, and portions thereof, said portions being at least 10 nucleotides, is performed by computer homology search.

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61. A method of screening a human subject to diagnose a disorder affecting the brain or genetic predisposition therefor, comprising the steps of:

a) assaying nucleic acid of a human subject to determine a presence or an absence of a mutation altering an amino acid sequence, expression, or biological activity of at least one nGPCR that is expressed in the brain, wherein the nGPCR comprises an amino acid sequence selected from the group consisting of SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:68, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, and SEQ ID NO:120, and allelic variants thereof, and wherein the nucleic acid corresponds to a gene encoding the nGPCR; and

b) diagnosing the disorder or predisposition from the presence or absence of said mutation, wherein the presence of a mutation altering the amino acid sequence, expression, or biological activity of the nGPCR in the nucleic acid correlates with an increased risk of developing the disorder.

62. The method of claim 61, wherein the nGPCR is nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70, or an allelic variant thereof.

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63. The method of claim 61, wherein the nGPCR is nGPCR-51 or nGPCR-52 or an allelic variant thereof.

64. The method of claim 61, wherein the disease is a mental disorder.

65. The method of claim 61, wherein the assaying step comprises at least one procedure selected from the group consisting of:

a) comparing nucleotide sequences from the human subject and reference sequences and determining a difference of either

5 at least a nucleotide of at least one codon between the nucleotide sequences from the human subject that encodes an nGPCR-42 allele and an nGPCR-42 reference sequence;

at least a nucleotide of at least one codon between the nucleotide sequences from the human subject that encodes an nGPCR-46 allele and an nGPCR-46 reference sequence;

10 at least a nucleotide of at least one codon between the nucleotide sequences from the human subject that encodes an nGPCR-48 allele and an nGPCR-48 reference sequence;

15 at least a nucleotide of at least one codon between the nucleotide sequences from the human subject that encodes an nGPCR-49 allele and an nGPCR-49 reference sequence;

at least a nucleotide of at least one codon between the nucleotide sequences from the human subject that encodes an nGPCR-51 allele and an nGPCR-51 reference sequence;

20 at least a nucleotide of at least one codon between the nucleotide sequences from the human subject that encodes an nGPCR-52 allele and an nGPCR-52 reference sequence;

25 at least a nucleotide of at least one codon between the nucleotide sequences from the human subject that encodes an nGPCR-61 allele and an nGPCR-61 reference sequence;

at least a nucleotide of at least one codon between the nucleotide sequences from the human subject that encodes an nGPCR-63 allele and an nGPCR-63 reference sequence; or

30 at least a nucleotide of at least one codon between the nucleotide sequences from the human subject that encodes an nGPCR-70 allele and an nGPCR-70 reference sequence;

b) performing a hybridization assay to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences;

c) performing a polynucleotide migration assay to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences; and

5 d) performing a restriction endonuclease digestion to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences.

66. The method of claim 65 wherein the assaying step comprises: performing a polymerase chain reaction assay to amplify nucleic acid comprising nGPCR-42, nGPCR-46, nGPCR-48, 10 nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 coding sequence, and determining nucleotide sequence of the amplified nucleic acid.

67. A method of screening for an nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 hereditary mental disorder genotype in a 15 human patient, comprising the steps of:

a) providing a biological sample comprising nucleic acid from said patient, said nucleic acid including sequences corresponding to alleles of nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70; and

20 b) detecting the presence of one or more mutations in the nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 allele; wherein the presence of a mutation in a nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 allele is indicative of a hereditary mental disorder genotype.

25 68. The method of claim 67 wherein said biological sample is a cell sample.

69. The method of claim 67 wherein said detecting the presence of a mutation comprises sequencing at least a portion of said nucleic acid, said portion comprising at least one codon of said nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, 30 nGPCR-63, or nGPCR-70 allele.

70. The method of claim 67 wherein said nucleic acid is DNA.

71. The method of claim 67 wherein said nucleic acid is RNA.

72. A kit for screening a human subject to diagnose a mental disorder or a genetic predisposition therefor, comprising, in association:

5 a) an oligonucleotide useful as a probe for identifying polymorphisms in a human nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 gene, the oligonucleotide comprising 6-50 nucleotides in a sequence that is identical or complementary to a sequence of a wild type human nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 gene sequence or nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 coding sequence, except for one sequence difference selected
10 from the group consisting of a nucleotide addition, a nucleotide deletion, or nucleotide substitution; and

15 b) a media packaged with the oligonucleotide, said media containing information for identifying polymorphisms that correlate with mental disorder or a genetic predisposition therefor, the polymorphisms being identifiable using the oligonucleotide as a probe.

73. A method of identifying a nGPCR allelic variant that correlates with a mental disorder, comprising the steps of:

20 a) providing a biological sample comprising nucleic acid from a human patient diagnosed with a mental disorder, or from the patient's genetic progenitors or progeny;

25 b) detecting in the nucleic acid the presence of one or more mutations in an nGPCR that is expressed in the brain, wherein the nGPCR comprises an amino acid sequence selected from the group consisting of SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:68, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, and SEQ ID NO:120, and allelic variants thereof, and wherein the nucleic acid includes sequence corresponding to the gene or genes encoding nGPCR;

30 wherein the one or more mutations detected indicates an allelic variant that correlates with a mental disorder.

74. The method of claim 73 wherein the nGPCR is nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70, or an allelic variant thereof.

75. A purified and isolated polynucleotide comprising a nucleotide sequence encoding a nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 allelic variant identified according to claim 74.

5 76. A host cell transformed or transfected with a polynucleotide according to claim 75 or with a vector comprising the polynucleotide.

77. A purified polynucleotide comprising a nucleotide sequence encoding nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or 10 nGPCR-70 of a human with a mental disorder;
wherein said polynucleotide hybridizes to the complement of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:8, SEQ ID NO:31, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, SEQ ID NO:59, and SEQ ID 15 NO:60 under the following hybridization conditions:

- a) hybridization for 16 hours at 42C in a hybridization solution comprising 50% formamide, 1% SDS, 1 M NaCl, 10% dextran sulfate; and
- b) washing 2 times for 30 minutes at 60C in a wash solution comprising 0.1x SSC and 1% SDS;

20 wherein the polynucleotide that encodes nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 amino acid sequence of the human differs from SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:8, SEQ ID NO:31, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:53, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, 25 SEQ ID NO:58, SEQ ID NO:59, and SEQ ID NO:60 by at least one residue.

78. A vector comprising a polynucleotide according to claim 77.

79. A host cell that has been transformed or transfected with a polynucleotide according to 30 claim 77 and that expresses the nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 protein encoded by the polynucleotide.

80. The host cell of claim 79 that has been co-transfected with a polynucleotide encoding and expressing the nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52,

nGPCR-61, nGPCR-63, or nGPCR-70 amino acid sequence set forth in SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:68, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, and SEQ ID NO:120.

81. A method for identifying a modulator of biological activity of nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 comprising the steps of:

- 10 a) contacting a cell according to claim 79 in the presence and in the absence of a putative modulator compound; and
- b) measuring nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 biological activity in the cell;
wherein decreased or increased nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 biological activity in the presence versus absence of the putative modulator is indicative of a modulator of biological activity.

82. A method to identify compounds useful for the treatment of a mental disorder, said 20 method comprising the steps of:

- a) contacting a composition comprising nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 with a compound suspected of binding nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70; and
- 25 b) detecting binding between nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 and the compound suspected of binding nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70;
wherein compounds identified as binding nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 are candidate compounds 30 useful for the treatment of a mental disorder.

83. A method for identifying a compound useful as a modulator of binding between nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or

nGPCR-70 and a binding partner of nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 comprising the steps of:

- 5 a) contacting the binding partner and a composition comprising nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 in the presence and in the absence of a putative modulator compound; and
- b) detecting binding between the binding partner and nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70;
wherein decreased or increased binding between the binding partner and nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or 10 nGPCR-70 in the presence of the putative modulator, as compared to binding in the absence of the putative modulator is indicative a modulator compound useful for the treatment of a mental disorder.

84. The method of claim 82 or claim 83 wherein the composition comprises a cell expressing nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70 on its surface.

85. The method of claim 84 wherein the composition comprises a cell transformed or transfected with a polynucleotide that encodes nGPCR-42, nGPCR-46, nGPCR-48, nGPCR-49, 20 nGPCR-51, nGPCR-52, nGPCR-61, nGPCR-63, or nGPCR-70.

86. A method of purifying a G protein from a sample containing said G protein comprising the steps of:

- 25 a) contacting said sample with a polypeptide of claim 1 for a time sufficient to allow said G protein to form a complex with said polypeptide;
- b) isolating said complex from remaining components of said sample;
- c) maintaining said complex under conditions which result in dissociation of said G protein from said polypeptide; and
- d) isolating said G protein from said polypeptide.

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87. The method of claim 86 wherein said sample comprises an amino acid sequence selected from the group of sequences consisting of SEQ ID NO:61 to SEQ ID NO:120.

88. The method of claim 86 wherein said polypeptide comprises an amino acid sequence homologous to a sequence selected from the group of sequences consisting of SEQ ID NO:61 to SEQ ID NO:120.

5 89. The method of claim 86 wherein said polypeptide comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:61, SEQ ID NO:62, SEQ ID NO:68, SEQ ID NO:91, SEQ ID NO:94, SEQ ID NO:96, SEQ ID NO:97, SEQ ID NO:99, SEQ ID NO:100, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116, SEQ ID NO:117, SEQ ID NO:118, SEQ ID NO:119, and SEQ ID NO:120.

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90. An isolated nucleic acid molecule comprising a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence homologous to SEQ ID NO:117.

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91. The nucleic acid molecule of claim 90 wherein said polypeptide comprises SEQ ID NO:117.

92. An isolated nucleic acid molecule comprising SEQ ID NO:57.

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93. An isolated polypeptide comprising an amino acid sequence homologous to SEQ ID NO:117.

94. The polypeptide of claim 93 comprising SEQ ID NO:117.

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95. A method of identifying a compound that binds to nGPCR-51 comprising the steps of:

a) contacting a composition comprising nGPCR-51, or a polypeptide homologous thereto, and a polypeptide comprising SEQ ID NO:183, or a polypeptide homologous thereto, with said test compound; and

b) determining whether said test compound binds to nGPCR-51.

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96. The method of claim 95 wherein said determining whether said test compound binds to nGPCR-51 is by measuring the displacement of said polypeptide comprising SEQ ID NO:183 from a complex between said polypeptide and nGPCR-51.

97. The method of claim 95 wherein said polypeptide comprising SEQ ID NO:183 is radiolabeled.

98. The method of claim 97 wherein step b) is determined by comparatively measuring 5 radioactivity of nGPCR-51 bound to said radiolabeled polypeptide comprising SEQ ID NO:183 with the radioactivity of nGPCR-51 in the presence of the test compound.

99. The method of claim 95 wherein step b) comprises a binding assay selected from the 10 group consisting of filter binding, scintillation proximity assay, gel-shift assay, radiolabeled competition assay, and ELISA.

100. A method for identifying a compound that modulates the activity of nGPCR-51 comprising the steps of :

15 a) contacting nGPCR-51 with a compound; and
b) determining whether nGPCR-51 activity has been modulated.

101. The method of claim 100 wherein said activity is neuropeptide binding.

102. The method of claim 101 wherein said neuropeptide binding is determined by binding to 20 a polypeptide comprising SEQ ID NO:183.

103. The method of claim 101 wherein said neuropeptide binding is determined by binding to a polypeptide comprising SEQ ID NO:184.

25 104. The method of claim 100 wherein said activity is neuropeptide signaling.

105. The method of claim 104 wherein said neuropeptide signalling is determined by binding to a polypeptide comprising SEQ ID NO:183.

30 106. The method of claim 104 wherein said neuropeptide signalling is determined by binding to a polypeptide comprising SEQ ID NO:184.

107. A method of screening a human subject to diagnose a disorder affecting the brain or genetic predisposition therefor, comprising the steps of:

a) assaying nucleic acid of a human subject to determine a presence or an absence of a mutation altering an amino acid sequence, expression, or biological activity of nGPCR-51, or an allelic variant thereof; and

5 b) diagnosing the disorder or predisposition from the presence or absence of said mutation, wherein the presence of a mutation altering the amino acid sequence, expression, or biological activity of the nGPCR in the nucleic acid correlates with an increased risk of developing the disorder.

108. The method of claim 107 wherein the disorder is schizophrenia.

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109. The method of claim 107 wherein the disorder is an attention disorder.

110. The method of claim 107 wherein the disorder is anxiety.

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111. The method of claim 107 wherein the disorder is depression.

112. The method of claim 107 wherein the disorder is obesity.

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113. A kit for screening a human subject to diagnose a mental disorder or a genetic predisposition therefor, comprising, in association:

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a) an oligonucleotide useful as a probe for identifying polymorphisms in a human nGPCR-51, the oligonucleotide comprising 6-50 nucleotides in a sequence that is identical or complementary to a sequence of a wild type human nGPCR-51 gene sequence or nGPCR-51 coding sequence, except for one sequence difference selected from the group consisting of a nucleotide addition, a nucleotide deletion, or nucleotide substitution; and

b) a media packaged with the oligonucleotide, said media containing information for identifying polymorphisms that correlate with mental disorder or a genetic predisposition therefor, the polymorphisms being identifiable using the oligonucleotide as a probe.

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114. A method of identifying a nGPCR allelic variant that correlates with a mental disorder, comprising the steps of:

a) providing a biological sample comprising nucleic acid from a human patient diagnosed with a mental disorder, or from the patient's genetic progenitors or progeny;

b) detecting in the nucleic acid the presence of one or more mutations in an nGPCR that is expressed in the brain, wherein the nGPCR comprises SEQ ID NO:117 or an allelic variant thereof, and wherein the nucleic acid includes sequence corresponding to the gene or genes encoding nGPCR;

5 wherein the one or more mutations detected indicates an allelic variant that correlates with a mental disorder.

115. A method for identifying a modulator of biological activity of nGPCR-51 comprising the steps of:

10 a) contacting a cell according to claim 79 in the presence and in the absence of a putative modulator compound; and

 b) measuring nGPCR-51 biological activity in the cell;

 wherein decreased or increased nGPCR-51 biological activity in the presence versus absence of the putative modulator is indicative of a modulator of biological activity.

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116. A method of identifying a compound useful for the treatment of a mental disorder, said method comprising the steps of:

 a) contacting a composition comprising nGPCR-51 with a compound suspected of binding nGPCR-51; and

20 b) detecting binding between nGPCR-51 and the compound suspected of binding nGPCR-51;

 wherein a compound identified as binding nGPCR-51 is a candidate compound useful for the treatment of a mental disorder.

25 117. A method for identifying a compound useful as a modulator of binding between nGPCR-51 and a binding partner of nGPCR-51 comprising the steps of:

 a) contacting the binding partner and a composition comprising nGPCR-51 in the presence and in the absence of a putative modulator compound; and

 b) detecting binding between the binding partner and nGPCR-51;

30 wherein decreased or increased binding between the binding partner and nGPCR-51 in the presence of the putative modulator, as compared to binding in the absence of the putative modulator is indicative a modulator compound useful for the treatment of a mental disorder.

118. An isolated nucleic acid molecule comprising a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence homologous to SEQ ID NO:118.

119. The nucleic acid molecule of claim 118 wherein said polypeptide comprises SEQ ID 5 NO:118.

120. An isolated nucleic acid molecule comprising SEQ ID NO:58.

121. An isolated polypeptide comprising an amino acid sequence homologous to SEQ ID 10 NO:118.

122. The polypeptide of claim 93 comprising SEQ ID NO:118.

123. A method of identifying a compound that binds to nGPCR-52 comprising the steps of:

15 a) contacting a composition comprising nGPCR-52, or a polypeptide homologous thereto, and a polypeptide comprising SEQ ID NO:183, or a polypeptide homologous thereto, with said test compound; and

b) determining whether said test compound binds to nGPCR-52.

20 124. The method of claim 123 wherein said determining whether said test compound binds to nGPCR-52 is by measuring the displacement of said polypeptide comprising SEQ ID NO:183 from a complex between said polypeptide and nGPCR-52.

125. The method of claim 123 wherein said polypeptide comprising SEQ ID NO:183 is 25 radiolabeled.

126. The method of claim 125 wherein step b) is determined by comparatively measuring radioactivity of nGPCR-52 bound to said radiolabeled polypeptide comprising SEQ ID NO:183 with the radioactivity of nGPCR-52 in the presence of the test compound.

30 127. The method of claim 123 wherein step b) comprises a binding assay selected from the group consisting of filter binding, scintillation proximity assay, gel-shift assay, radiolabeled competition assay, and ELISA.

128. A method for identifying a compound that modulates the activity of nGPCR-52 comprising the steps of :

- a) contacting nGPCR-52 with a compound; and
- b) determining whether nGPCR-52 activity has been modulated.

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129. The method of claim 128 wherein said activity is neuropeptide binding.

130. The method of claim 129 wherein said neuropeptide binding is determined by binding to a polypeptide comprising SEQ ID NO:183.

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131. The method of claim 129 wherein said neuropeptide binding is determined by binding to a polypeptide comprising SEQ ID NO:184.

132. The method of claim 128 wherein said activity is neuropeptide signaling.

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133. The method of claim 132 wherein said neuropeptide signalling is determined by binding to a polypeptide comprising SEQ ID NO:183.

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134. The method of claim 132 wherein said neuropeptide signalling is determined by binding to a polypeptide comprising SEQ ID NO:184.

135. A method of screening a human subject to diagnose a disorder affecting the brain or genetic predisposition therefor, comprising the steps of:

- a) assaying nucleic acid of a human subject to determine a presence or an absence of a mutation altering an amino acid sequence, expression, or biological activity of nGPCR-52, or an allelic variant thereof; and
- b) diagnosing the disorder or predisposition from the presence or absence of said mutation, wherein the presence of a mutation altering the amino acid sequence, expression, or biological activity of the nGPCR in the nucleic acid correlates with an increased risk of developing the disorder.

136. The method of claim 135 wherein the disorder is schizophrenia.

137. A kit for screening a human subject to diagnose a mental disorder or a genetic predisposition therefor, comprising, in association:

- a) an oligonucleotide useful as a probe for identifying polymorphisms in a human nGPCR-52, the oligonucleotide comprising 6-50 nucleotides in a sequence that is identical or 5 complementary to a sequence of a wild type human nGPCR-52 gene sequence or nGPCR-52 coding sequence, except for one sequence difference selected from the group consisting of a nucleotide addition, a nucleotide deletion, or nucleotide substitution; and
- b) a media packaged with the oligonucleotide, said media containing information for identifying polymorphisms that correlate with mental disorder or a genetic predisposition

10 therefore, the polymorphisms being identifiable using the oligonucleotide as a probe.

138. A method of identifying a nGPCR allelic variant that correlates with a mental disorder, comprising the steps of:

- a) providing a biological sample comprising nucleic acid from a human patient 15 diagnosed with a mental disorder, or from the patient's genetic progenitors or progeny;
- b) detecting in the nucleic acid the presence of one or more mutations in an nGPCR that is expressed in the brain, wherein the nGPCR comprises SEQ ID NO:118 or an allelic variant thereof, and wherein the nucleic acid includes sequence corresponding to the gene or genes encoding nGPCR;

20 wherein the one or more mutations detected indicates an allelic variant that correlates with a mental disorder.

139. A method for identifying a modulator of biological activity of nGPCR-52 comprising the steps of:

- a) contacting a cell according to claim 79 in the presence and in the absence of a putative modulator compound; and
- b) measuring nGPCR-52 biological activity in the cell;

wherein decreased or increased nGPCR-52 biological activity in the presence versus absence of the putative modulator is indicative of a modulator of biological activity.

30 140. A method of identifying a compound useful for the treatment of a mental disorder, said method comprising the steps of:

- a) contacting a composition comprising nGPCR-52 with a compound suspected of binding nGPCR-52; and

b) detecting binding between nGPCR-52 and the compound suspected of binding nGPCR-52;

wherein a compound identified as binding nGPCR-52 is a candidate compound useful for the treatment of a mental disorder.

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141. A method for identifying a compound useful as a modulator of binding between nGPCR-52 and a binding partner of nGPCR-52 comprising the steps of:

a) contacting the binding partner and a composition comprising nGPCR-52 in the presence and in the absence of a putative modulator compound; and

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b) detecting binding between the binding partner and nGPCR-52;

wherein decreased or increased binding between the binding partner and nGPCR-52 in the presence of the putative modulator, as compared to binding in the absence of the putative modulator is indicative a modulator compound useful for the treatment of a mental disorder.

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